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(71) Applicant: **United Biomedical Inc.**
2 Nevada Drive
Lake Success New York 11042(US)

(72) Inventor: **Chang Yi Wang**
159 Hill Park Avenue
Great Neck, N.Y. 11021(US)
Inventor: **Hoseln, Barbara**
196 E. 75th Street
New York, N.Y. 10021(US)

(74) Representative: **Hansmann, Axel et al**
Albert-Rosshaupter-Strasse 65
W-8000 München 70(DE)

(54) **Synthetic peptides specific for the detection of antibodies to HCV, diagnosis of HCV infection and prevention thereof as vaccines.**

(57) The present invention relates to peptides which are immunoreactive to antibodies to HCV or NANBHV and a method of detecting the presence of HCV or NANBHV antibodies in body fluids by using the peptides as the antigen. The peptides are selected from both the envelope and non-structural protein regions of the HCV or NANBHV. The detection method includes enzyme linked immunosorbent assay or other immunoassay procedures. The peptides and conjugates or polymers thereof are also useful as immunogens in generating high titer antibodies to HCV or in vaccines.

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INTRODUCTION

The present invention relates to peptides specific for the diagnosis and prevention of hepatitis C virus (HCV) infection, or non-A non-B hepatitis (NANBH). More particularly, the present invention is directed to synthetic peptides which are specific for the detection of antibodies to HCV in body fluids and immunoassays using the same. The invention also includes the use of the synthetic peptides in compositions as antigens for eliciting the production of monoclonal and polyclonal antibodies against HCV and as immunogens in vaccines for the prevention of NANBH or HCV infection.

In recent years, non-A, non-B hepatitis (NANBH) has become the most common form of post-transfusion hepatitis. Studies involving the experimental inoculation of chimpanzees provided evidence that the infectious agent was a lipid-containing virus resembling members of the *Togaviridae* family.

Recently, this etiological agent, termed hepatitis C virus (HCV) has been shown to be an RNA virus with a genome size of ~ 10 kilobases encoding a single polyprotein which can be further processed into several structural and nonstructural proteins (1-4). Additional computer-assisted protein analysis demonstrates that HCV shares sequence similarity with the polyproteins of animal pestiviruses and flaviviruses as well as members of two plant virus supergroups (5).

More recently, a number of reports have led to an increasingly coherent understanding of the function of various regions of the virus and of the relationships among genomic fragments isolated from variants or closely related viruses.

A summary of the HCV structure, beginning at the N terminus of the virus, follows. The HCV comprises a structural protein region and nonstructural (NS) protein regions. The structural protein region is further divided into capsid and envelop proteins. The NS protein regions are further divided into NS-1 to NS-5 regions (3).

The postulated capsid region (AA1-AA120) has been shown to contain highly immunoreactive conserved epitopes with enhanced sensitivity in the detection of hepatitis C infection (6-8). The region appears to consist of two segments of equal length (AA1-61, AA62-AA120), which are homologous to one another, perhaps as a result of a gene duplication, and are also homologous to the N terminal core region of yellow fever virus (9), also a flavivirus (Table 1A). Both halves, as represented by peptides VIIIE (AA2-AA62) and IXD (AA65-AA119), disclosed in application serial No. 558,799, have been shown to be immunoreactive. A genomic fragment of a NANBH virus cloned by Arima et al. (10), designated clone 2, contains a Gly-Pro-Arg-Leu-Gly sequence identical to residues 39-43 in peptide VIIIE (Table 1B), placing this clone 2 fragment in the putative core region of a related virus. Two other sequences from NANBH viruses, cloned by Reyes et al. (11) and by Arima et al. (clone 1) (12), show sequence similarities with the capsid region of yellow fever virus (Table 1C). Thus, there appears to be a number of related viruses, all of which have highly immunogenic capsid regions, as evidenced by the ease of cloning. Variants of hepatitis C (J, J-1, J-4) are also highly conserved in this region (2-4), so the other clones mentioned by Arima et al. may be from different viruses, rather than from variants of HCV.

Mishiro and colleagues have isolated a cDNA clone from the plasma of a chimpanzee infected with NANBH which codes for a host cellular sequence bearing an epitope which is reactive with sera from individuals who are PCR positive for HCV (13). The sequence of the immunoreactive peptide (GOR epitope) is not encoded by HCV and was reported not to resemble a published sequence of HCV spanning three-quarters of the genome (1) or the 5'-terminal sequence of HCV (2) covering the upstream quarter of the genome. However, inspection of the GOR epitope sequence revealed 47% homology with an N-terminal fragment covered by peptide VIIIE described in UBI Applications Serial No. 558,799. Lesser degrees of homology were obtained from comparison with the N-terminus of the yellow fever virus capsid protein (33%) (9) and the protein segment corresponding to clone 1 of Arima et al. (37.5%) (12) (See Table 1D).

The presence of antibodies which are cross-reactive with the GOR epitope sequence in HCV infected individuals may be explained by structural similarity of the GOR epitope with the corresponding region of the HCV capsid protein. Compared with anti-C100, antibodies to the C100 region, previously identified by Houghton et al.; antibodies to peptide VIIIE share the following characteristics with anti-GOR: they both are present in some but not all anti-C100 positive sera; they can be detected in anti-C100 negative sera from both acute and chronic NANBH patients; they appear earlier than anti-C100 in the seroconversion series; they are detected in more seroconversion panels than anti-C100 (13); and they are present in 1-2% of normal controls and 15-20% of HBsAg positive individuals. Early NANBH assays reported to react with host-determinant cytoplasmic antigens may in fact have detected anti-HCV capsid protein cross-reactivity.

The postulated envelope (env) region consists of amino acids 120 to 400. The env glycoproteins of flaviviruses are key targets for immunization because the env region is a major antigen of free viral particles and plays a central role in flavivirus biology. The env region mediates binding to cell receptors and

probably facilitates fusion to membranes. It also induces protective immune responses after vaccination or natural infection with a flavivirus (14,15) and stimulates cell-mediated immunity (16). The type-specific epitopes on env are the ones most closely associated with protective immune responses to flaviviruses (17-19). There are a number of hypervariable regions in the HCV env region, based on a comparison of US and Japanese strains (2), which may indicate epitopes for strain specific reactivity.

The non-structural protein NS-1, in addition to the small M protein of the envelope, has been shown to contribute to protective immunity in dengue fever (20,21). Inspection of sequences and hydrophobicity profiles shows that the HCV NS-1 region contains two similar domains (Table 1E). A dominant motif in this region is cysteine pairs separated by five or more amino acids.

The NS-2 region is of unknown function and little has been reported on its characteristics.

By analogy with yellow fever virus, the HCV NS-3 region may contain protease activity required for viral replication (22). A trypsin-like serine protease active site has been localized in yellow fever virus by means of site-directed mutagenesis of NS-3 to a catalytic triad consisting of His-53, Asp-77 and Ser-138. The corresponding region in HCV is the N-terminal third of NS-3, with the critical residues being His-1103, Asp-1127 and Ser-1188. The remainder of the HCV NS-3 region consists of a region which shows immunoreactivity. This region appears to consist of three subregions homologous to one another (Table 1F) and these subregions bear a distant relationship to the repeated segments of the NS-1 region.

The most widely studied region to date is the NS-4 nonstructural region. Although its function is unknown, it contains highly immunoreactive regions, primarily in the region designated as C100 by Houghton et al. (1), which became the basis for a HCV diagnostic test using recombinant technology. A high degree of structural homology is observed between part of the C100 HCV sequence with a corresponding region in the yellow fever virus (Table 1G). While this region detects antibody to the virus primarily responsible for NANBH (23), experimentally it has been shown in prior United Biomedical Inc.'s application Serial No. 558,799 and numerous recent reports that there are shortcomings in both sensitivity and specificity in the tests relying on the C100 polypeptide as an antigen. However, synthetic peptides from the NS-4 region described in prior application Serial No. 558,799 overcome the problem of non-specific reactivity.

The nonstructural region proximal to the C terminus of HCV is NS-5, the site of polymerase (pol) activity. The Gly-Asp-Asp sequence in this region is conserved across many viruses(11). Maeno et al. have isolated a clone corresponding to a sequence upstream of the pol site in the NS-5 region which is immunoreactive and which reacts specifically with sera from patients in the chronic phase of NANBH(24).

Through an extensive series of experiments involving serological validation using select specimens chosen from the screening of thousands of sera with hundreds of carefully designed synthetic peptides, we have further characterized the capsid protein related immunoreactive peptides and have identified additional immunoreactive epitopes contained within the envelope, NS-1, NS-2, NS-3, and NS-5 protein regions.

Synthetic peptides have been increasingly used to map antigenic or immunogenic sites on the surface of proteins, an approach recently termed "site-directed-serology". We, at United Biomedical, have taken this approach to identify and characterize highly antigenic epitopes on the envelope and core proteins of HIV and to develop sensitive and specific immunoassays for the detection of antibodies to HIV (previously designated HTLV-III) (25-27). See U.S. Patent 4,735,896, issued April 5, 1988 and (U.S. Patent 4,879,212 issued Nov. 7, 1989, the contents of which are, hereby, fully incorporated by reference (28,29). Subsequently, a series of finely mapped and well-characterized HTLV-III related synthetic peptides were employed in the development of synthetic peptide-based diagnostic assays for the detection of HTLV-III antibodies in infected individuals (30,31). See also U.S. Patent 4,833,071 issued May 23, 1989, U.S.S.N. 07/297,635 filed January 13, 1989 and USSN 07/469,294 filed January 24, 1990. These assays have provided superior sensitivity, excellent specificity, and, in certain cases, an unmatched capability to differentiate infections between two closely related viruses, thus overcoming many of the existing problems associated with biologically-derived tests based on either viral lysates or recombinant DNA-derived proteins.

It is, therefore, an objective of the present invention to employ the identified and characterized immunoreactive HCV peptides in the development of a detection or diagnostic procedure to identify and monitor HCV infection.

A further objective is to chemically synthesize a test reagent which can then be used to detect the presence of antibodies to HCV in body fluids and to diagnose NANBH.

Another objective is to develop a vaccine which, when introduced into healthy mammals, including humans, will stimulate production of efficacious antibodies to HCV, thereby providing protection against HCV infection.

A further objective is to provide a synthetic immunogen which can be used in mammals for the development of monoclonal and polyclonal antibodies to HCV.

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Amino acid sequence (single letter code) derived from the N-terminal capsid protein of the Yellow fever virus (AAS AAG9, upper line; Ref. V), another WABVHV sequence cloned by Reyes et al. (AAL-AA55, middle line; Fig. 3, Ref. 11) and a third WABVHV sequence cloned by Arino et al. (AAS-AA35, lower line; Ref. 12) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids, that have been inserted to optimize the alignment.

GOR Epitope Sequence (Ref.13)

AA3-AA19 Segment of HCV Capsid Peptide VIII
of prior application serial no. 558,799
Arima et al. Clone 1 (AA22-AA37, Ref.12)

Yellow Fever Virus (AA3-AA19, Ref.9)

Amino acid sequences (single letter code) derived from the GOR Eclipse (upper line; Ref.139), a segment of the HCV capsid peptide VIII representing HCV AA4-AA19 of prior application (second line), AA22-AA37 of the NANBHV sequence (clone 1) reported by Arlino et al (third line; Ref. 12) and a segment of the Yellow fever Virus N-terminal capsid protein (AA2-AA19, Ref.9) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

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Table 1E

HCV-NS1(J-1)	C	R	R	-	L	-	T	D	F	O	G	W	G	P	I	S	A	N	G	S	G	P	D	Q	R	P	Y	C	-	-	W	H	Y	P	P	K	P	C	G	-	I	V	-	P	A	-	-	K	S	V	C	G	P	V	Y	C
	D	R	S	G	A	P	I	-	Y	-	-	S	W	G	E	N	D	T	D	V	F	V	L	N	T	R	P	L	G	N	U	-	F	G	-	-	C	T	M	M	S	T	G	F	T	K	-	V	C	G	A	P	P	C		

Amino acid sequences (single letter code) derived from two segments of the HCV NS-1 protein (upper line, AA59-AA508; and lower line, AA520-AA569; are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

HCV-NS-1(J-1)	C	R	L	T	D	F	D	O	G	W	G	P	I	S	A	N	G	S	G	P	D	Q	R	P	Y	C	W	Y	P	P	K	P	C	G	I	V	P	A	K	S	V	C	G	P	V	Y	C		
HCV-NS-1(J-4)	C	R	P	I	D	W	F	A	Q	G	W	G	P	I	T	Y	T	E	P	D	S	P	D	Q	R	P	Y	C	W	Y	A	P	R	P	C	G	I	V	P	A	S	Q	V	C	G	P	V	Y	C
HCV-NS-1(J)	C	R	P	I	D	E	F	A	Q	G	W	G	P	I	T	N	D	M	P	E	S	D	Q	R	P	Y	C	W	Y	A	P	R	P	C	G	I	V	P	A	S	Q	V	C	G	P	V	Y	C	

Amino acid sequences (single letter code) derived from three HCV strains (J-1, J-4 and J) for a segment of the NS-1 protein (AA59-AA508); are aligned for comparison of homology.

HCV-NS-1(PT)	D	R	S	G	A	P	T	Y	S	W	G	E	N	D	T	V	D	F	V	L	N	T	R	P	P	L	G	N	U	F	G	C	T	W	M	N	S	T	G	F	T	K	V	C	G	A	P	P	C
HCV-NS-1(J)	D	R	F	G	A	P	T	Y	S	W	G	E	N	E	T	D	V	L	L	S	N	T	R	P	P	Q	G	N	U	F	G	C	T	W	M	N	S	T	G	F	T	K	T	C	G	G	P	P	C

Amino acid sequences (single letter code) derived from two HCV strains (PT and J) for a segment of the NS-1 protein (AA520-AA569) are aligned for comparison of homology.

HCV-NS-1(PT)	D	R	S	G	A	P	T	Y	S	W	G	E	N	D	T	V	D	F	V	L	N	T	R	P	P	L	G	N	U	F	G	C	T	W	M	N	S	T	G	F	T	K	V	C	G	A	P	P	C
HCV-NS-1(J)	D	R	F	G	A	P	T	Y	S	W	G	E	N	E	T	D	V	L	L	S	N	T	R	P	P	Q	G	N	U	F	G	C	T	W	M	N	S	T	G	F	T	K	T	C	G	G	P	P	C

Amino acid sequences (single letter code) derived from two HCV strains (PT and J) for a segment of the NS-1 protein (AA520-AA569) are aligned for comparison of homology.

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Table 1F

MCV-NS3	D	F	I	P	-	-	V	E	N	L	E	T	T	M	R	S	P	V	F	T	D	M	S	S	-	P	P	V	-	-	V	-	P	Q	-	S	-	F	Q	V	A	H	L	H	A	P	T	G	S	G	K	S	T	-	-	K	V	P	
	P	N	I	R	T	G	V	R	T	I	-	T	T	G	-	S	P	I	-	T	T	-	S	T	Y	G	K	F	L	A	D	-	G	G	C	S	G	G	A	Y	D	-	-	I	I	I	C	D	E	C	H	S	T	D	A	T			
	P	N	I	E	E	-	V	A	-	L	S	T	T	G	E	I	P	-	F	-	Y	G	K	A	-	I	P	-	L	E	V	I	K	G	G	R	N	L	I	F	C	-	-	N	S	K	K	K	C	D	E	L	-	A	-	-	A	K	L

Amino acid sequences (single letter code) derived from three segments of the HCV NS-3 protein (AA119-1241, upper line; AA1276-AA1324, middle line; and AA1360-AA1407, lower line) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

HCY	V	V	L	A	T	A	T	P	P	G	S	V	T
BVD	V	V	A	M	T	A	T	P	A	G	S	V	T
HOG	V	V	A	M	T	A	T	P	A	G	T	V	T
YFV	T	I	L	M	I	A	T	P	P	G	T	S	D

HCY	Q	R	R	G	R	I	G	R	G	K	P	G	I	-	Y	R
BVD	Q	R	R	G	R	V	G	R	V	K	P	G	R	Y	Y	R
HOG	Q	R	R	G	R	V	G	R	V	K	P	G	R	Y	Y	R
YFV	Q	R	R	G	R	I	G	R	-	N	P	N	R	D	G	D

Multiple alignment of two highly conserved segments encoded within the NS-3 protein region (single letter code) of HCV (AA1344-AA1356, upper line; and AA1486-AA1500, lower line) respectively, Bovine Diarrhea Virus (BVD, AA2025-AA2037; AA2181-AA2196), Hog Cholera Virus (HOG, AA1886-AA1898; AA-2042-AA2057) and Yellow Fever Virus (YFV AA1800-AA1812; AA1944-AA1958) are aligned for comparison of homology.

Table 1C

5	V V I Y G R Y Y L S G K P A I I P Q R E V L Y R E F D E M	Q H L P Y I E N G M M L A
10	A A E Y L V B L S E L A K E G G E A M D T I S V F L S E E G S	Y I E N G M M L A
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E M F K Q K A L	G L L Q I A S R Q A E Y I	NCV-NS4
E M T I V M L F I L A G L L I	I S G M - - Y I	YFV-NS4

Amino acid sequences (single letter code) derived from a segment of the HCV NS-4 protein and a corresponding segment of the Yellow fever virus NS-4 protein (lower line, AA200-4217A, Ref. 9) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

REFERENCES

1. Houghton M, Choo Q-L, Kuo G: NANBV diagnostics and vaccines. EPO 0318216A1 (1989).
2. Okamoto H, Okada S, Sugiyama S, Yotsumoto S, Tanaka T, Yoshizawa H, Tsuda F, Miyakawa Y, Mayumi M: The 5' terminal sequence of the hepatitis C virus genome. Jpn. J. Exp. Med. 60:167 (1990).
3. Houghton M, Choo Q-L, Kuo G: NANBH diagnostics and vaccines. EPO 0388232A1 (1990).
4. Kato N, Hijikata M, Ootsuyama Y, et al: Molecular cloning of the human hepatitis C virus genome from Japanese patients with non-A, non-B hepatitis. Proc. Natl. Acad. Sci. USA 87:9524 (1990).
5. Miller RH, Purcell RH: Hepatitis C virus shares amino acid sequence similarity with pestiviruses and flaviviruses as well as members of two plant virus supergroups. Proc. Natl. Acad. Sci. USA 87:2057 (1990).
6. Hosein B, Fang CT, Zhang ML, et al: Improved serodiagnosis of hepatitis C virus infection with synthetic peptide antigen from capsid protein. Proc. Natl. Acad. Sci. USA 88:3647 (1991).

7. UBI HCV EIA Product Insert. (1990).
8. Okamoto H, Munekata E, Tsuda F, et al: Enzyme-linked immunosorbent assay for antibodies against the capsid protein of hepatitis C virus with a synthetic oligopeptide. Jap. J. Exp. Med. 60:223 (1990).
9. Rice CM, Lencho EM, Eddy SR, et al: Nucleotide sequence of yellow fever virus: Implications for flavivirus gene expression and evolution. Science 229:726 (1985).
10. Arima T, Takamizawa A, Mori C, et al: A lambda gt11-cDNA clone specific for chronic hepatitis C generated from pooled serum presumably infected by hepatitis C virus. Gastroenterologia Japonica 24:545 (1989).
11. Reyes GR, Purdy MA, Kim J, et al: Isolation of a cDNA from the virus responsible for enterically transmitted non-A, non-B hepatitis. Science 247:1335 (1990).
12. Arima T, Nagashima H, Murakami S, et al: Cloning of a cDNA associated with acute and chronic hepatitis C infection generated from patients serum RNA. Gastroenterologia Japonica 24:540 (1989).
13. Mishiro S, Hoshi Y, Takeda K, et al: Non-A, non-B hepatitis specific antibodies directed at host-derived epitope: Implication for an autoimmune process. Lancet 336:1400 (1990).
14. Brinton MA: in The Togaviridae and Flaviviridae, ed. Schlesinger S and Schlesinger MJ. Plenum Press, NY pp. 327-374 (1986).
15. Mandl CW, Guirakhoo F, Holzmann H, Heinz FX, Kunz C: Antigenic structure of the flavivirus envelope, protein E at the molecular level, using tick-borne encephalitis virus as a model. J. Virol. 63:564 (1989).
16. Bray M, Falgout B, Zhao B, et al: in Vaccines '89. Modern Approaches to New Vaccines Including Prevention of AIDS, ed Lerner RA, Ginsberg H, Chanock RM and Brown F. Cold Spring Harbor Laboratory, NY, pp357-362 (1989).
17. Roehrig JT, Hunt AR, Johnson J, Mathews JH: ibid. pp347-350 (1989).
18. Rothman AL, Kurane J, Ennis FA: ibid. pp363-366 (1989).
19. Roehrig JT: in The Togaviridae and Flaviviridae, ed Schlesinger S and Schlesinger MJ. Plenum Press NY, pp251-278 (1986).
20. Bray M, Meu R, Lai CJ: Meeting on Modern Approaches to New Vaccines Including Prevention of AIDS. Cold Spring Harbor Laboratory, Sept 12-16, 1990. Abst 70.
21. Falgout B, Bray M, Schlesinger JJ, Lai CJ: Immunization of mice with recombinant vaccinia virus expressing authentic dengue virus nonstructural protein NS1 protects against lethal dengue virus encephalitis. J. Virol. 64:4356 (1990).
22. Chambers TJ, Weir RC, Grakoui A, et al: Evidence that the N-terminal domain of nonstructural protein NS-3 from yellow fever virus is a serine protease responsible for site-specific cleavages in the viral polyprotein. Proc. Natl. Acad. Sci. USA 87:8898 (1990).
23. Kuo G., Choo Q-L, Alter HJ, et al: An assay for circulating antibodies to a major etiologic virus of human non-A, non-B hepatitis: Science 244:362 (1989).
24. Maeno M, Kaminaka K, Sugimoto H, et al: A cDNA clone closely associated with non-A, non-B hepatitis. Nucleic Acids Res. 18:2685 (1990).
25. Wang CY: Synthetic-peptide-based immunodiagnosis of retrovirus infections: Current status and future prospects. In: Synthetic Peptides in Biotechnology, ed. Mizrahi A, Advances in Biotechnological Processes, 10:131 (1988).
26. Wang JG, Steel S, Wisniewolski R, Wang CY: Detection of antibodies to HTLV-III using a synthetic peptide of 21 amino acid residues corresponding to a highly antigenic segment of gp41 envelope protein. Proc. Natl. Acad. Sci. USA 83:6159 (1986).
27. Wang CY: European Patent Application Publication: EPO 0328403 (1989). Synthetic peptides related to the HIV-gp120 env protein, and their use.
28. Wang CY, Wang JG: U.S. Patent 4879212 (1989). Peptide composition and method for the detection of antibodies to HTLV-III.
29. Wang CY, Wang JG: U.S. Patent 4735896 (1988). Synthetic peptide and process of using same for the detection and diagnosis of AIDS and pre-AIDS conditions.
30. Wang CY, Wang JG, Walters DW: U.S. Patent 4833071 (1989). Peptide composition as antigen for detection of antibodies to HTLV-I, as a vaccine for ATL, and methods therefore.
31. Wang CY: U.S.S.N. 07/297635. Synthetic peptide compositions with immunoreactivities to antibodies to HTLV.

BRIEF DESCRIPTION OF THE INVENTION

According to the present invention, a series of synthetic peptides representing immunoreactive regions

of the postulated envelope protein and nonstructural proteins NS-1, NS-2, NS-3 and NS-5 of the hepatitis C virus (HCV), each arranged in a specific sequence, has been identified and made by solid phase peptide synthesis. These peptides have been found to be useful for the detection of antibodies to HCV in sera and body fluids and for the diagnosis of non-A, non-B hepatitis (NANBH). Because of their immunoreactivity, it is expected that these peptides are also useful in stimulating production of antibodies to HCV in healthy mammals such as Balb/C mice, and in a vaccine composition to prevent HCV or NANBHV infection.

According to the present invention, a peptide composition useful for the detection of antibodies to HCV and diagnosis of NANBH comprises a peptide from the envelope, NS-1, NS-2, NS-3 and NS-5 regions of the HCV represented by the following sequences:

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(a) Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys-X

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Pep1

(b) Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-
Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys-X

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Pep2

(c) Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Leu-
His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
Asp-Gln-Ala-Glu-Thr-Ala-Gly-X

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Pep3

(d) Asp-Pro-Ser-His-Ile-Thr-Ala-Glu-Ala-Ala-Gly-Arg-Arg-Leu-Ala-
Arg-Gly-Ser-Pro-Pro-Ser-Val-Ala-Ser-Ser-Ser-Ala-Ser-Gln-Leu-
Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-
Pro-X

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Pep4

(e) Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-
Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X

40

45

Pep5

(f) Asp-Pro-Gln-Ala-Arg-Val-Ala-Ile-Lys-Ser-Leu-Thr-Glu-Arg-Leu-
Thr-Val-Gly-Gly-Pro-Leu-Thr-Asn-Ser-Arg-Gly-Glu-Asn-Cys-Gly-
Tyr-Arg-Arg-Cys-Arg-Ala-Ser-X

50

Pep6

55

- (g) Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-Ser-Thr-
Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-
5 Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-
Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-X
Pep7
- 10 (h) Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
15 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala-X
Pep8
- (i) Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-
20 Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-
Ile-Gln-Leu-Ile-Asn-X
Pep9
- 25 (j) Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-
Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Glu-His-Lys-Phe-Asn-Ser-
30 Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-X
Pep10
- (k) Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
35 Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-
40 X
Pep11
- (l) Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
45 Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
50 Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X
Pep12

- (m) Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-
Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
5 Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-
Arg-Arg-X
Pep13
- (n) Cys-Lys-Pro-Leu-Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-
His-Glu-Tyr-Pro-Val-Gly-Ser-Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-
15 Asp-X
Pep14
- (o) Glu-Glu-Tyr-Val-Glu-Ile-Arg-Gln-Val-Gly-Asp-Phe-His-Tyr-Val-
20 Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-Cys-Pro-Cys-Gln-Val-Pro-
Ser-Pro-X
Pep15
- (p) Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-
Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu-
30 X
Pep16
- (q) Gly-Pro-Ala-Asp-Gly-Met-Val-Ser-Lys-Gly-Trp-Arg-Leu-Leu-Ala-
35 Pro-Ile-Thr-Ala-Tyr-Ala-Gln-Gln-Thr-Arg-Gly-Leu-Leu-Gly-Cys-
Ile-Ile-Thr-Ser-Leu-Thr-Gly-Arg-Asp-Lys-Asn-Gln-Val-Glu-Gly-
X
40 Pep17
- (r) Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-
45 Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-
Glu-Leu-Ala-Ala-Lys-Leu-Val-Ala-Leu-X
Pep18
- (s) Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
50 Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys-X
55 Pep19

These 19 peptides are in addition to Peptide VIII E, a peptide from the structural protein region, and Peptides IIH and V, peptides from the non-structural protein region which have also been found to be reactive and useful for the detection of antibodies to HCV and diagnosis of NANBH.

5 Peptide VIII E has the following sequence:

Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thy-Lys-Arg-Asn-Thr-Asn-Arg-
Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gly-Gln-Ile-Val-Gly-Gly-
10 Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-
Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X,

15 Peptide IIH has the following sequence:

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-Glu-
20 Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-Glu-Gln-
Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X

25 Peptide v has the following sequence:

Lys-Gln-Lys-Ala-Leu-Gly-Leu-Leu-Gln-Thr-Ala-Ser-Arg-Gln-Ala-Glu-
Val-Ile-Ala-Pro-Ala-Val-Gln-Thr-Asn-Trp-Gln-Lys-Leu-Glu-Thr-Phe-
30 Trp-Ala-Lys-His-Met-Trp-Asn-Phe-X

wherein X is -OH or -NH₂ and analogues, segments, mixtures, conjugates and polymers thereof.

35 Further, according to the present invention, the peptides by themselves, or when coupled to a protein or a polymeric carrier of homo or hetero dimers or higher oligomers by the use of homo or hetero functional multivalent cross linking reagents, or when directly synthesized and conjugated to a branching polyvalent lysine resin, can be used to elicit the production of antibodies to HCV in healthy mammals, including humans.

40 The method comprises introducing an effective amount of the peptide composition containing each of the individual peptides, analogues or segments or a mixture or a combination thereof, or in a polymeric form, into the body of a healthy mammal by intraperitoneal or subcutaneous injection.

Vaccines containing the peptides according to the present invention as the key immunogen may also be prepared as described above or by known methods. It is expected that such vaccine compositions may be useful to prevent HCV infection or NANBH.

45

BRIEF DESCRIPTION OF DRAWING

Fig. 1 is a photograph of a computer-generated structure of an octameric peptide immunogen.

50 DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, nineteen peptides and their analogues including segments have been selected from the nonstructural regions of HCV and chemically synthesized. These peptides including their analogues are useful for the detection of antibodies to HCV in body fluids, the diagnosis of
55 NANBH, and for the vaccination of healthy mammals by stimulating the production of antibodies to HCV. These peptides are arranged in the following sequences:

- (a) Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
5 Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys-X
Pep1
- (b) Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-
10 Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys-X
Pep2
- (c) Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Leu-
15 His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
Asp-Gln-Ala-Glu-Thr-Ala-Gly-X
20 Pep3
- (d) Asp-Pro-Ser-His-Ile-Thr-Ala-Glu-Ala-Ala-Gly-Arg-Arg-Leu-Ala-
Arg-Gly-Ser-Pro-Pro-Ser-Val-Ala-Ser-Ser-Ser-Ala-Ser-Gln-Leu-
25 Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-
Pro-X
30 Pep4
- (e) Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-
35 Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X
Pep5
- (f) Asp-Pro-Gln-Ala-Arg-Val-Ala-Ile-Lys-Ser-Leu-Thr-Glu-Arg-Leu-
40 Thr-Val-Gly-Gly-Pro-Leu-Thr-Asn-Ser-Arg-Gly-Glu-Asn-Cys-Gly-
Tyr-Arg-Arg-Cys-Arg-Ala-Ser-X
45
50
55

Pep6

(g) Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-Ser-Thr-
 Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-
 5 Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-
 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-X

Pep7

(h) Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
 Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
 10 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala-X

Pep8

(i) Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-
 20 Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-
 Ile-Gln-Leu-Ile-Asn-X

Pep9

(j) Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-
 Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Glu-His-Lys-Phe-Asn-Ser-
 25 Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-X

Pep10

(k) Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
 35 Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
 Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
 Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-
 40 X

Pep11

(l) Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
 45 Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
 Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
 Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X

Pep12

(m) Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-
 55 Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-

5 Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-
Arg-Arg-X

Pep13

10 (n) Cys-Lys-Pro-Leu-Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-
His-Glu-Tyr-Pro-Val-Gly-Ser-Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-
Asp-X

Pep14

15 (o) Glu-Glu-Tyr-Val-Glu-Ile-Arg-Gln-Val-Gly-Asp-Phe-His-Tyr-Val-
Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-Cys-Pro-Cys-Gln-Val-Pro-
20 Ser-Pro-X

Pep15

25 (p) Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-
Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu-
X

Pep16

30 (q) Gly-Pro-Ala-Asp-Gly-Met-Val-Ser-Lys-Gly-Trp-Arg-Leu-Leu-Ala-
Pro-Ile-Thr-Ala-Tyr-Ala-Gln-Gln-Thr-Arg-Gly-Leu-Leu-Gly-Cys-
35 Ile-Ile-Thr-Ser-Leu-Thr-Gly-Arg-Asp-Lys-Asn-Gln-Val-Glu-Gly-
X

Pep17

40 (r) Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-
Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-
45 -Glu-Leu-Ala-Ala-Lys-Leu-Val-Ala-Leu-X

Pep18

50 (s) Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys-X

Pep19

55 These 19 peptides are in addition to Peptide VIII E, a peptide from the structural protein region, and Peptides IIH and V, peptides from the non-structural protein region which have also been found to be reactive and useful for the detection of antibodies to HCV and diagnosis of NANBH.

Peptide VIII E has the following sequence:

5 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-Thr-Asn-Arg-
Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gly-Gln-Ile-Val-Gly-Gly-
Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-
10 Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X,

Peptide II H has the following sequence:

15 Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-Glu-
Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-Glu-Gln-
Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X

Peptide V has the following sequence:

20 Lys-Gln-Lys-Ala-Leu-Gly-Leu-Leu-Gln-Thr-Ala-Ser-Arg-Gln-Ala-Glu-
25 Val-Ile-Ala-Pro-Ala-Val-Gln-Thr-Asn-Trp-Gln-Lys-Leu-Glu-Thr-Phe-
Trp-Ala-Lys-His-Met-Trp-Asn-Phe-X

30 Wherein X is -OH or -NH₂ and analogues, segments, mixtures, conjugates, and polymers thereof.

These peptides may comprise combinations or segments, i.e. longer or shorter peptide chains by having more amino acids added to the terminal amino acids, or by amino acids removed from either terminal end.

35 These peptides may also comprise analogues to accommodate strain-to-strain variations among different isolates of HCV. HCV is indicated to have frequent mutations. Therefore, it is expected that variant strains, such as PT, J, J-1 and J-4 (1-4) exist. Adjustments for conservative substitutions and selection among the alternatives where non-conservative substitutions are involved, may be made in the prescribed sequences (e.g. see Table 1E, Table 8c and Table 11 for possible amino acid substitutions in the hypervariable regions of the envelope and NS-1 proteins). These analogues of the synthetic peptides may
40 therefore comprise substitutions, insertions and/or deletions of the recited amino acids of the above sequence to accommodate the various strains, as long as the immunoreactivity recognizable by the antibodies to HCV is preserved

These peptides may also comprise conjugates, i.e., they may be coupled to carrier proteins such as bovine serum albumin (BSA) or human serum albumin (HSA). Furthermore, these peptides may comprise
45 polymers, i.e., they may be synthesized on a polymeric resin or in dimeric, tetrameric, octameric and decahexyl forms of the peptide or their analogues, such as a branching octameric lysine resin.

The branchine poly-L-lysine can be Lys₈ Lys₄ Lys₂ Lys, Lys₄ Lys₂ Lys, Lys₂ Lys, Lys; the last Lys can be attached to Y as in Lys₄ Lys₂ Lys-Y wherein Y is -OH, -NH₂ or an amino acid containing no side chain functional group, such as alanine, valine, glycine, etc. Y can be inserted to facilitate synthesis onto the 4-
50 methylbenzylhydramine resin. The conjugates and polymers of the peptides are also useful in the present invention.

The amino acid sequences of the polypeptide as described in the invention useful as test reagents for the detection of antibodies to HCV in body fluids and diagnosis of NANBH are selected to correspond to segments of the amino acid sequence of the postulated envelope and non-structural proteins of HCV
55 designated as env, NS-1, NS-2, NS-3 and NS-5 based on amino acid sequence information derived from Houghton et al. (13), Okamoto et al (2) and Kato et al (4).

In selecting regions of the HCV protein for epitope analysis, peptides of about 40 mer size with amino acid sequences covering the complete HCV envelope and non-structural proteins NS-1, NS-2, NS-3 and

NS-5 were synthesized. These were tested for their immunoreactivity with special specimens previously selected through the screening of thousands of patient and normal sera for their unique immunoreactivity with HCV. Nineteen peptides from the postulated envelope and nonstructural protein regions NS-1, NS-2, NS-3 and NS-5 designated as pep1, pep2, pep3, pep4, pep5, pep6, pep7, pep8, pep9, pep10, pep11, pep12, pep13, pep14, pep15, pep16, pep17, pep18 and pep19 and their analogues were identified to have specific immunoreactivity with the positive HCV sera.

At present, available knowledge of protein structure has not enabled the scientist to predict the amino acid sequences that may represent highly immunogenic epitopes. The usefulness of a peptide as an antigen or immunogen must be empirically determined. We have only been able to identify and characterize immuno-reactive epitopes through an extensive process which we call "serological validation". The following example shows how difficult it is to identify immuno-reactive epitopes.

For example, a clone designated as C33c encoded within the NS-3 region was reported to possess immunoreactivity(3). This clone spans 265 amino acid residues. Assuming a useful peptide must be at least 6 amino acids in length and that the upper limit for synthetic peptides in reasonable yield is 120 residues, the number of possible unique peptides from the C33c regions is 23,028. For the entire HCV genome, the figure is about 260,000.

In addition, we have shown that extraction conditions are critical for the expression of the immunopotency of a peptide (Example 4C), so the number of uniquely extracted peptides from this region is in multiples of 23,028. The possibilities for post-extraction modification, such as pH adjustment (Example 4B) further increase the possible selections to $>10^6$. If amino acid substitutions at various positions are taken into consideration, this figure will quickly increase to several millions. In contrast to the HCV core region, in which peptides VIII E and IX D were the optimal analogues, longer peptides are not preferred over shorter analogues in the NS-3/C33c region. For example, the 42 mer 279B shown on Table 4D has only 3% of the reactivity of the 37 mer peptide 3, designated as 279A in Table 4D. Of 30 peptides spanning the C33c region tested, only one was found to be useful. The antigenic index as referred in Houghton et al (3) did not prove to be a useful guide to epitopes, as the profile for peptide 3 is positive for only 30% of its sequence and negative for the remaining 70%.

The strategy for serological validation also depends on the expected characteristics of the target epitopes. Universal immunodominant epitopes, such as the gp41 transmembrane peptide of HIV-1, may be screened by a single representative serum sample from a patient known to be infected with the virus. Epitopes which are not recognized by all infected individuals, or those for which antibody is produced late or only transiently, and especially epitopes which give rise to neutralizing antibodies, must be screened by large panels of sera. For example, peptide 272B shown in Table 4A was initially tested on a panel of eight sera from HCV infected individuals (Panel 1). Only one sample was definitely positive with an absorbance of 880 mA. Three were weakly reactive (<200 mA) and four were negative.

The identification of the immuno-reactive epitopes is also dependent on the panel of sera used. The more closely the panel represents the population most likely to be seropositive for the desired epitope, the greater the chance that the epitope will be identified. For example, peptides synthesized from the NS-1 region, which were hypothesized to be important for generating neutralizing antibodies, gave only weakly reactive or negative results on screening with a very large number ($n > 200$) of samples from individuals who were newly infected and/or chronically infected with HCV. However, a panel of 24 samples from asymptomatic individuals from a known hepatitis virus endemic geographical region, Taiwan and mainland China, yielded two samples with absorbances of >2000 mA against multiple NS-1 peptides.

Finally, if the desired purpose of a targeted peptide/epitope is to extend the range of reactivity of an assay comprised of previously identified epitopes, then a large number of samples from individuals at risk of infection but seronegative against known epitopes must be employed for screening. Unfortunately, the most critical samples from clinically proven and documented cases of infection may be available in quantities insufficient for screening purposes. This is another complication/difficulty encountered in serological validation for determining the immunoreactivity of a peptide.

The process of "serological validation" is particularly difficult when the epitopes to be identified elicit antibodies only in a subpopulation of an infected patient group. When such epitopes become targets for identification, special attention must be paid to synthetic peptides which show very weak reactivity when tested by an enzyme immunoassay.

Fortunately, the low background absorbance of synthetic peptides allows for the precise detection of weak reactivities. In some cases, absorbances of 50 mA versus background reading are of sufficient significance and can lead to the identification of important epitopes through successive refinement of the amino acid sequence of a peptide. The utmost technical skill is required to obtain consistent and reliable results when working in the range of absorbances below 200-300 mA. For example: Peptides 261E and

261F shown on Table 4D were reactive with only one of eight HCV sera panel members (Panel I), with absorbances of 307 and 269 mA, respectively. Yet this weak reactivity led to the eventual identification of pep3 (or 279A), toward which half of the panel is reactive, and toward which some additional reactive samples show absorbances of >2000 mA.

5 Based on the immunoreactivities of the peptides according to the present invention, it is believed that these peptides may also be useful in a vaccine to prevent NANBH. The peptide when coupled to a protein, or synthesized on a polymeric carrier resin (e.g., an octameric lysine resin) or when polymerized to homo or hetero dimers or higher oligomers by cysteine oxidation, or induced disulfide cross linking, or by use of
10 homo or hetero functional multivalent cross linking reagents, can be introduced to normal subjects to stimulate production of antibodies to HCV in healthy mammals.

The advantages of using synthetic peptides are known.

Since the peptides according to the present invention are not derived biologically from the virus, there is no danger of exposing the normal subjects who are to be vaccinated to the disease causing pathogen.

15 The peptides can be chemically synthesized easily. This means that there is no involvement with HCV at any time during the process of making the test reagent or the vaccine. Another problem which can be minimized by the process of the present invention is the false positive results caused by the presence of antigenic material co-purified with the HCV fusion protein. Certain normal individuals have antibodies to E. coli or yeast proteins which are cross reactive with the antigenic materials from the expression system. Sera from these normal individuals may show a positive reaction in the immunoassays.

20 Further, with appropriate amino acid modification or substitutions, it is expected that various peptide analogues based on the prescribed amino acid sequence can be synthesized with properties giving rise to lower background readings or better binding capacity to solid phases useful for HCV antibody screening assays.

Moreover, because the peptide compositions of the present invention are synthetically prepared, the
25 quality can be controlled and as a result, reproducibility of the test results can be assured. Also, since very small amounts of a peptide are required for each test procedure, and because the expense of preparing a peptide is relatively low, the cost of screening body fluids for antibodies to HCV, diagnosis of NANBH infection, and the preparation of a vaccine is relatively low.

30 The peptides prepared in accordance with the present invention can be used to detect HCV infection and diagnose NANBH by using them as the test reagent in an enzyme-linked immunoadsorbent assay (ELISA), an enzyme immunodot assay, an agglutination based assay, or other well-known immunosassay devices. The following examples serve to illustrate the present invention and are not to be used to limit the scope of the invention.

35 EXAMPLE 1

Measurement of Relative (%) Immunoreactivity for HCV synthetic peptides by an Enzyme-Linked Im- munosorbent Assay

40 As an example to illustrate how relative (%) immunoreactivity for HCV synthetic peptides is measured, wells of 96-well plates are coated for 1 hour at 37° C, with each of the following peptides: IIH, V, VIIIE and pep11 at 5 ug/mL at 100 uL per well in 10mM NaHCO₃ buffer, pH 9.5. The peptide coated wells were then incubated with 250 uL of 3% by weight of gelatin in PBS in 37° C for 1 hour to block non-specific protein
45 binding sites, followed by three washes with PBS containing 0.05% by volume of TWEEN 20 and then dried. The test specimens containing a panel of eight well-characterized HCV antibody positive patient sera were diluted with PBS containing 20% by volume normal goat serum, 1% by weight gelatin and 0.05% by volume TWEEN 20 at dilutions of 1:20 volume to volume, respectively. 200 uL of the diluted specimens were added to each of the wells and allowed to react for 15 minutes at 37° C.

50 The wells were then washed six times with 0.05% by volume TWEEN 20 in PBS in order to remove unbound antibodies. Horseradish peroxidase conjugated goat anti-human IgG was used as a second antibody tracer to bind with the HCV antibody-peptide antigen complex formed in positive wells. 100 uL of peroxidase labeled goat anti-human IgG at a dilution of 1:1800 in 1% by volume normal goat serum, 0.05% by volume TWEEN 20 in PBS was added to each well and incubated at 37° C for another 15 minutes.

55 The wells were washed six times with 0.05% by volume TWEEN 20 PBS to remove unbound antibody and reacted with 100uL of the substrate mixture containing 0.04% by weight orthophenylenediamine (OPD) and 0.12% by volume hydrogen peroxide in sodium citrate buffer, pH 5.0.

This substrate mixture was used to detect the peroxidase label by forming a colored product. Reactions were stopped by the addition of 100 uL of 1.0M H₂SO₄ and the A_{492nm} measured. Results of relative

immunoreactivity for each of the peptides obtained from this study are shown in Table A using peptide II H as the reference.

5 **Table A**

Peptide Code	A_{492nm} (Panel I, No. 1 to 8)								Total	%
	1	2	3	4	5	6	7	8		
10 IIH	0.812	0.656	3.114	2.737	1.066	2.254	2.599	3.478	16.712	100
V	0.834	1.060	2.931	0.534	0.137	0.434	0.303	2.787	9.020	54
VIII E	2.745	2.208	2.468	3.032	0.054	2.108	0.730	3.006	16.351	98
15 Pep11	0.241	0.715	3.162	1.020	0.568	2.166	3.330	3.477	14.690	88

EXAMPLE 2

20 Comparison of HCV Immunoreactivities by a Well-characterized 8 Member HCV Serum Panel (Panel I) for % Relative Immunoreactivity with a Group of HCV Capsid Protein Related Peptides by an Enzyme Immunoassay

A 36mer HCV capsid peptide recently disclosed by Okamoto et al. (8) as the basis of an HCV EIA was synthesized for the purpose of comparison of immunoreactivity with peptides VIIIA, VIIIB and VIII E (Table 2A). According to a procedure described in Example 1, peptides were coated at concentrations of 5, 1 and 0.2 $\mu\text{g/mL}$ for immunopotency comparison. This 36mer exhibited only 47.8% of the reactivity of VIII E (Table 2A). More importantly, when tested by our well-characterized HCV serum panel used for serological validation, only 4 out of 8 samples reacted with the 36mer, compared with 7 out of 8 with VIII E. The C terminal end of this 36mer does not appear to contribute to the peptide's HCV immunoreactivity, since IXD is not greater in reactivity than IXC (Table 2A).

In addition, a 61mer peptide and fragments thereof consisting of a 30mer, a 40mer and a 50mer corresponding to sequences from Arima clone 1, which is homologous to the capsid region of the flavivirus yellow fever virus, were synthesized and compared in immunoreactivity with peptide VIII E from the corresponding region of HCV (Table 2B). The 40mer and 61mer of clone 1 exhibited the most reactivity. However these were only 21.1% and 20.7%, respectively, of the immunoreactivity of peptide VIII E.

	Sequence	% Relative Immunoreactivity
Okamoto et al. (8) (36mer)	RRGPRLGVRATRKTSERSOPRGRROP IPKVRPEGR	47.6%
VIII A	GPRLGVRATRKTSERSOPRGR	32.7%
VIII B	VGGYLLPRGPRGPRLGVRATRKTSERSOPRGR	48.9%
VIII E	STIPKPRKTKRNTNRPPQVYKFGGGI VGGYLLPRGPRGPRLGVRATRKTSERSOPRGR	100.0%
IX C	TUAGPGYMPPLYGMEGGGAGGULLSPGSGSPSUGPTOPRRRSNHLG	57.9%
IX D	IPKVRPEGR TUAGPGYMPPLYGMEGGGAGGULLSPGSGSPSUGPTOPRRRSNHLG	58.9%
IX E	GRRGP IPKVRPEGR TUAGPGYMPPLYGMEGGGAGGULLSPGSGSPSUGPTOPRRRSNHLG	50.2%

	Sequence	% Relative Immunoreactivity
30 mer	PGGKXPVGR IKNVREGRIDATYIRKR	0.7%
40mer	KEKETATINPGKHKXPVGR IKNVREGRIDATYIRKR	21.1%
50mer	NDTNKKORRYKEKETATINPGKHKXPVGR IKNVREGRIDATYIRKR	17.8%
61mer	KKCEASHCEAEKNDTHKKORRYKEKETATINPGKHKXPVGR IKNVREGRIDATYIRKR	20.7%

55 EXAMPLE 3

Relative (%) Immunoreactivity for NS-1 Synthetic Peptides by an Enzyme-Linked Immunosorbent Assay

(A) Identification of Immunoreactive NS-1 Peptides.

Wells of 96-well plates were coated for 1 hour at 37° C with each of the 16 peptides (designated as peptides 241A-C, 231A-E, 232A-D, 233C, 234A-C), synthesized according to sequences derived from the NS-1 region (Table 3A), at 5 ug/mL at 100 uL per well in 10 mM NaHCO₃ buffer, pH 9.5. Each peptide's immunoreactivity was measured as previously described (see Example 1), using an 8 member serum panel (Panel I).

All sixteen peptides showed little or no reactivity with serum panel I. The most reactive peptide, pep1 (designated 231c in Table 3A), had an immunopotency index of 13.9%, compared with peptide VIIIE on the same panel. There were isolated examples of epitope recognition; for example, for sample 4, all analogues of the 232 series had absorbances less than or equal to 20 mA except for the longest peptide, 232D, which had an absorbance of 785 mA. However, the remaining 7 panel members were negative when tested with 232D.

After screening these 16 NS-1 region derived peptides with more than 200 additional HCV positive sera with little or no demonstrated immunoreactivities, immunoreactivities of these 16 NS-1 peptides with other sera were sought. A panel of serum samples from individuals coming from regions in which hepatitis C is endemic, namely mainland China and Taiwan, were tested for evidence of reactivity to these NS-1 protein derived peptides. Twenty-four samples were chosen from individuals who had no recognizable symptoms of non-A, non-B hepatitis and for whom the peptide based HCV EIA, Format C, as described in Example 11, was nonreactive. Seven of the 24 samples (29%) were reactive against one or more peptides from the NS-1 region, indicative of the presence of long term protective antibodies responsive to this region. This 7 member panel (designated as Panel II, CH1-CH7) was used to further characterize these NS-1 peptides for their immunoreactivity.

The peptide with the greatest reactivity against the serum Panel II again was pep1 (designated 231c in Table 3A). Using this peptide as a standard, the relative immunoreactivity for each of the other 15 peptides from the NS-1 region are calculated in Table 3A.

Detailed results from the seven member serum Panel II on four of the most immunoreactive analogues (i.e. pep1, or 231C; pep2, or 232A; 233C and 234A) are tabulated in Table 3B. The reactivities of 231C and 232A are complementary in that CH-1 and CH-2 are strongest on 231C, whereas CH-3 through CH-7 are stronger on 232A.

(B) NS-1 Reactivity in Early and Long-term HCV Infection.

In addition, all sixteen NS-1 peptides were tested on panels of samples representing HCV-antibody positive donors (n = 9) in an early stage of infection, namely plasmapheresis donors with the first occurrence of an ALT level >100 i.u./L, and those asymptomatic individuals (n = 14) disqualified from blood donation because of a reactive result for anti-HIV or HBc, for whom the anti-HCV result probably represents a past infection. These select panels were chosen from hundreds of HCV positive sera for their ability to recognize NS-1 antigens. The results of testing the panels with the 16 NS-1 peptides are given in Table 3C. For both groups, peptide designated as 232A (pep2) had the greatest immunoreactivity. Using pep2 as a standard, the relative immunoreactivity of each peptide was calculated (Table 3C).

Synthetic Peptides with their Amino Acid Sequences derived from the HCV NS-1 Protein Region

	Synthetic Peptides with their Amino Acid Sequences derived from the HCV NS-1 Protein Region		
	CPERLASCRPLTDFDGGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSVCGPVTCITPSVWVGITDRSGAPTYSUGENDTDVFLNMTRPPLGNHFGCTLNNSTGFTKVCGAPPC	% IP	
241A	GGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSV	63.9	
241B	CRPLTDFDGGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSV	92.9	
241C	CPERLASCRPLTDFDGGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSV	93.6	
231A	RPYCHNYTPPEPGIIPAKSVCGPVTC	83.2	
231B	ANGSGPDRPYCHNYTPPEPGIIPAKSVCGPVTC	96.4	
231C (Pep1)	GGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSVCGPVTC	100.0	
231D	CRPLTDFDGGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSVCGPVTC	65.3	
231E	CPERLASCRPLTDFDGGGCPISYANGSGPDRPYCHNYTPPEPGIIPAKSVCGPVTC	87.6	
232A (Pep2)			PPLGNHFGCTLNNSTGFTKVCGAPPC
232B			VFVNLNMTRPPLGNHFGCTLNNSTGFTKVCGAPPC
232C			SUGENDTDVFLNMTRPPLGNHFGCTLNNSTGFTKVCGAPPC
232D			DRSGAPTYSUGENDTDVFLNMTRPPLGNHFGCTLNNSTGFTKVCGAPPC
	VIGGAGNNTLNCPTDCFRKHFDATYSRGGSGPVITPRCLVDYPTRLVMPCITNTTIFKIRNTVGGVHRLAACHNTRGERCODEDRSLS		
233C	LHCPTDCFRKHFDATYSRGGSGPVITPRCLVDYPTRLVMPC		
234A			EAACHNTRGERCODEDRSLS
234B			VGGVHRLAACHNTRGERCODEDRSLS
234C			TTFKIRNTVGGVHRLAACHNTRGERCODEDRSLS

Table 3B

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A492nm by EIA (mA)				
Sample No. (Panel II)	231C (Pep1)	232A (Pep2)	233C	234A
CH-1	2237	202	123	118
CH-2	2472	261	174	232
CH-3	171	935	72	64
CH-4	218	1498	238	227
CH-5	311	621	114	206
CH-6	247	1128	175	202
CH-7	206	552	89	151

Table 3C

Panel I.D. Panel Size	Peptide Code	Early HCV Infection n = 9	Late HCV Infection n = 14
		% Relative Immunoreactivity in comparison to Pep2 (232A)	
10	241A	23.9	43.8
	241B	32.7	75.0
	241C	44.7	84.7
15	231A	46.8	48.4
	231B	30.9	46.7
	231C (Pep1)	88.6	62.7
	231D	23.3	43.5
20	231E	70.9	83.4
	232B	91.4	83.5
	232C	21.7	22.0
25	232D	50.0	46.7
	233A	9.9	17.5
	234A	9.5	15.7
30	234B	13.2	20.9
	234C	12.1	44.5

EXAMPLE 4

Relative (%) Immunoreactivity for NS-3 Protein Derived Synthetic Peptides by an Enzyme-Linked Im-
munosorbent Assay

(A) Identification of NS-3 Protein Derived Immunoreactive Peptides.

Wells of 96-well plates were coated for 1 hour at 37° C with each of the 30 peptides (designated as 261A-F, 262A-F, 272A-C, 274A-D, 275A-D, 278A-D and 279A,B,E), synthesized with sequences derived from the NS-3 region, at 5 ug/mL at 100 uL per well in 10 mM NaHCO₃ buffer, pH 9.5. The immunoreactivity of each peptide was measured by an 8 member HCV serum panel (Panel I). The peptide with the greatest immunoreactivity, pep3, designated 279A in Table 4D, had a relative immunoreactivity value of 23.9%, compared with peptide VIIIE (data not shown). When the immunoreactivity of peptide 3 was used as a standard to calculate the relative immunopotency for the other NS-3 peptides (Tables 4A, 4B, 4C and 4D), all other 29 peptides were found to be marginally immunoreactive. More surprisingly, the sequence of pep3 (or 279A), a 37mer, is entirely contained within peptides 261E, 261F, 274B, 274C, 274D, 279B and 279E, yet these seven larger peptides have relative immunoreactivity in the range of only 2.2 to 34%, when compared to their segment pep3. Another surprise was the observation that the mere addition of 5 residues to the N terminus of pep3 completely abrogates the reactivity of the peptide (see the relative immunoreactivity of pep3 vs. peptide 279B, Table 4D).

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x 1 d

272A	PVPQSFQVAHLNAPTGSCKSTKYPAAAYAAGTEVLVLPSPS	2.1
272B	YTMSPVFTONSSPPVPQSFQVAHLNAPTGSCKSTKYPAAAYAAGTEVLVLPSPS	38.0
272C	AVDFIPVENLETIMSPVFTONSSPPVPQSFQVAHLNAPTGSCKSTKYPAAAYAAGTEVLVLPSPS	11.8

8.2
1.4
10.7
11.6

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**Relative
luminescence**

275A	KYLVLNPSVAATLGFAYNSKANGIDPNIRTVRIITGGSPITVSTYCKFLADGGCSGGAYDIIIDCECHS	18.2
275B	KYLVLNPSVAATLGFAYNSKANGIDPNIRTVRIITGGSPITVSTYCKFLADGGCSGGAYDIIIDCECHS	11.1
275C	HLNAPTSGCKSTIKVPLAAYAAQGYKYLVLNPSVAATLGFAYNSKANGIDPNIRTVRIITGGSPITVSTYCKFLADGGCSGGAYDIIIDCECHS	9.6
275D	TNRSPVFTDHSPPVVPQSFQVAHILNAPTSGCKSTIKVPLAAYAAQGYKYLVLNPSVAATLGFAYNSKANGIDPNIRTVRIITGGSPITVSTYCKFLADGGCSGGAYDIIIDCECHS	5.4

275A
275B
275C
275D

Table 4C		
HCV NS-3 PROTEIN DERIVED SYNTHETIC PEPTIDES		
	GYKVLNPSVAATLGFCAVMSKANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVAL	X Relative Immunoreactivity
274A		2.0
274B	TVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVAL	34.0
274C	GCSSGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVAL	3.9
274D	ANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVAL	2.8
	GYKVLNPSVAATLGFCAVMSKANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVAL	
	GYKVLNPSVAATLGFCAVMSKANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDAT	X Relative Immunoreactivity
262A		6.9
262B	YKFLADGGCGGAYDIIICDECHNSTDAT	4.7
262C	ITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDAT	9.3
262D	PNIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDAT	3.6
262E	ATMSKANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDAT	4.7
262F	SVAATLGFCAVMSKANGIDPMIRIGVRTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDAT	5.1
Table 4D		
HCV NS-3 PROTEIN DERIVED SYNTHETIC PEPTIDES		
	RTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVALSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	X Relative Immunoreactivity
261A		7.2
261B	EVALLSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	3.1
261C	SVTPVPHNIEEVALSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	3.2
261D	TVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVALSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	15.8
261E	GGAAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVALSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	14.9
261F	RTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAGARLVWLATATPPGCVTPVPHNIEEVALSTTGEIPFYGKAIPLEVIGKGRHLIFCHSKKKODEL	21.4
279A (pH3)	GCSSGAYDIIICDECHNSTDATSILGIGTVLDOAETAG	100.0
279B	FLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAG	3.0
279E	RTITITGSPITYSTYKFLADGGCGGAYDIIICDECHNSTDATSILGIGTVLDOAETAG	2.2

55 (B) Enhancement of Peptide Immunoreactivity by pH Adjustment.

Although the immunoreactivities of 29 of the 30 NS-3 derived peptides, as originally synthesized and cleaved products, were marginal, the conformation of some peptides could be modulated by pH adjustment

to enhance their immunoreactivity.

Peptides dissolved at 1 mg/mL in H₂O, pH 4, were titrated to pH 11 by addition of diluted NaOH. After 5 min at pH 11, the pH of the peptide solution was brought down to 7.0 using diluted HCl. Immunoreactivity of the peptides thus treated was compared with reactivity prior to pH adjustment (Table 4E). Two- to three-fold increases in A492nm were seen. Some previously non-reactive serum samples were able to react with pH adjusted peptides. For instance, serum sample 1, which is non-reactive to 261C, has an absorbance of 1401 mA when tested with the corresponding pH adjusted peptide. Adjustment of pH increases the relative immunopotency of peptide 261C from 3.2% to 68.5%, compared with the standard pep3 (or 279A).

10 (C) Effect of Extraction Conditions after HF Cleavage on the Immunoreactivities of Peptides.

Peptide extraction conditions after HF cleavage were altered to test for their effect on peptide immunopotency after HF cleavage. Pep3 (or 279A) was extracted with acetic acid at pH 2, whereas pep3' was extracted with ammonium bicarbonate at pH 8. The latter extracted product showed a decrease in its reactivity in all reactive samples tested (Table 4F). The decrease ranged from 77.6% to 99.3%.

Table 4E

		A492nm (mA) by EIA							
		274B		275B		261C		272C	
		Ctrl	pH adj	Ctrl	pH adj	Ctrl	pH adj	Ctrl	pH adj
25	1	628	1604	210	591	6	1401	8	973
	2	148	466	37	159	5	499	74	255
	3	9	625	0	217	24	175	29	141
	4	464	1144	124	311	27	351	17	158

Table 4F

Effect of Extraction Conditions on Synthetic
Peptide's Immunopotency

	A492nm (mA) by EIA		† Decrease
	Pep3 Acetic Acid	Pep3' (NH ₄) ₂ CO ₃	
Blank	0	0	-
NRC	1	1	-
WRC	565	59	89.6
SRC	2213	495	77.6
#1	1550	329	78.8
#2	628	63	90.0
#3	1323	112	91.5
#4	1019	7	99.3
#5	1610	193	88.0

NRC: Negative Control
WRC: Weakly Reactive Control
SRC: Strongly Reactive Control

EXAMPLE 5

Relative (%) Immunoreactivity for NS-5 Protein Derived Synthetic Peptides by an Enzyme-Linked Im-
munosorbent Assay

Wells of 96-well plates were coated for 1 hour at 37° C with each of the three peptides derived from the NS-5 region of HCV (designated as pep4, pep5 and pep6). The results obtained (Table 5) show that all these peptides were immunoreactive with a unique group of 5 HCV positive sera.

Table 5

HCV NS-5 Protein Derived Synthetic Peptides

% Relative
Immunoreactivity

Amino Acid Sequence
 D P S H I T A E A G R R L A R G S P P S V A S S S A S Q L S A P S L K A T C T A N H D S P
 D A E L I E A N L L W R Q E M G N I T R V E S E N K V V I L D S F D P L V A E E D E R
 D P Q A R V A I K S L T E R L T V G G P L T N S R G E N C G Y R R C R A S R A S

28.6
 100.0
 17.0

Code	Sample No.	Pep4	Pep5	Pep6
Pep4	1	0.468	2.942	0.550
Pep5	2	0.659	0.370	0.245
Pep6	3	0.675	0.616	0.043
	4	0.063	1.316	0.162
	5	0.144	1.783	0.192

EXAMPLE 6Detection of Antibodies to HCV By an Agglutination Based Assay

The presently claimed HCV peptides, synthesized according to the Merrifield solid phase method, can be conjugated to bovine serum albumin (BSA) by a simple crosslinking method in the presence of a low percentage of glutaraldehyde solution, or with other crosslinking reagent such as m-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS).

Based on the above mentioned peptide-BSA conjugation process, conjugated peptide was absorbed onto double aldehyde fixed human O erythrocytes at pH 4.0. The peptide-conjugate coated erythrocytes were then treated with NaBH₄ to prevent non-specific protein binding. The peptide-conjugate coated erythrocytes were then washed with PBS and incubated with 5% normal human serum-PBS solution. These

processed cells were then used in an agglutination assay for the detection of HCV antibodies in both serum and plasma specimens. The specimens were diluted 1:10 in a sample diluent buffer and an equal volume of the indicator cells was mixed with the diluted specimens. The agglutination pattern was settled within one hour; and the assay results were read by eye. Serial bleedings from three well-characterized HCV seroconversion panels were tested for antibodies to HCV in the above-described HCV passive hemagglutination assay (PHA) employing Peptide VIII-E-BSA conjugate and Peptide IIH-BSA conjugate as the solid phase. The results were compared with the A492 and S/C of the peptide based HCV EIA (Format C, as described in Example 11) and C100 based HCV EIA (Table 6).

In brief, the PHA assay detected HCV antibodies in all three panels as early as there was an increase in A492 in the peptide based EIA (Format C). rC100 based EIA lagged behind the HCV PHA results by 4-8 weeks.

Table 6

Detection of HCV Specific Antibodies from Seroconversion Panels by Various HCV Antibody Assays

Series	Days	ALT	Format C HCV EIA S/C Ratio	C100 Based HCV EIA	HVC PHA Visual Score
A* (Serologicals Panel B)	0	40	0.108	0.03	-
	7	32	0.045	0.04	-
	14	32	0.025	0.06	++
	21	180	1.037	0.04	++
	50	401	7.193	0.19	++
	92	-	10.185	6.57	++
	105	-	9.770	6.57	++
B* (Serologicals Panel A)	0	39	0	0	-
	10	274	0.058	0	-
	14	346	0.128	0	-
	30	1175	7.835	6.5	++
	51	430	7.811	6.5	++
C* (Serologicals Panel C)	0	63	0.115	0.04	-
	2	81	1.607	0.04	++
	9	183	2.506	0.02	++++
	29	563	9.827	6.57	++++
	57	436	10.630	6.57	++++

* Case presented is a plasma donor from a commercial source. Day 0 designates first sample in the series and does not correspond to date of exposure.

Example 7

Detection of Antibodies to HCV by an Agglutination Assay Utilizing as the Solid Phase Immunosorbent Latex Particles Coated with HCV Peptide

Using the peptide-BSA conjugation process mentioned in the previous example, conjugated peptide VIII-E-BSA, was absorbed to latex particles (0.4 μ size) at pH 9.5. The peptide-conjugate coated latex particles were then treated with BSA to prevent nonspecific protein binding. These coated latex particles were then used in an agglutination assay for the detection of HCV antibodies. The specimens were mixed in a ratio of 1:1 with the latex solution (0.5%). The agglutination pattern was complete in a period of 15 min. Assay results were read by eye and by microscopic examination. The results of serial dilution samples from a well characterized anti-HCV positive plasma sample are summarized in Table 7. A coating concentration of 0.3 mg/mL was found to give optimal sensitivity for antibody detection. As a control for specificity, pooled plasma specimens from normal donors were tested in the peptide VII-BSA conjugate latex assay and were

found clearly negative.

Table 7

5 Rapid Detection of HCV Antibodies using VIIIE-BSA Sensitized Latex Particles and Scoring for Visual Agglutination Pattern

10	HCV Positive Control Dilution	Degree of Agglutination			
		VIIIE-BSA Latex Particle Concentration 2.4 mg/mL	1.2 mg/mL	0.6 mg/mL	0.3 mg/mL
15	1:1	4+	4+	4+	4+
	1:2	4+	4+	4+	4+
	1:5	4+	4+	4+	4+
20	1:10	4+	4+	4+	4+
	1:20	3+	4+	4+	4+
25	1:40	2+	3+	4+	4+
	1:80	+/-	-	+	3+
	1:160	-	-	-	+
30	1:320	-	-	-	+/-
	1:640	-	-	-	+/-
	NP 1:1	-	-	-	-
35	NP: Pooled Normal Plasma				

EXAMPLE 8

40 SYNTHESIS OF OCTAMERIC HCV PEPTIDE ANTIGENS AS KEY COMPONENTS OF IMMUNOGENS/VACCINES

The use of a limited sequential propagation of a trifunctional amino acid (or similar analogues) to form a core that serves as a low molecular weight matrix is the basic underlying principle for the formation of a radially branching multimeric peptide antigen system. The trifunctional amino acid, Boc-Lys(Boc), or di-(Boc)-Lys is most suitable since both N^α- and N^ε- amino acid groups are available as reactive ends. Thus, sequential propagation of di-(Boc)-Lys will generate 2ⁿ reactive ends. For example, the first level coupling of di-(Boc)-Lys will produce two reactive amino ends as a bivalent peptide antigen. Sequential generations of a second, third, and fourth step with di-(Boc)-Lys will therefore generate tetravalent, octavalent, and hexadecavalent peptide antigens respectively. As an example, an octameric HCV peptide immunogen with a structure of [Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys]₃-Lys₄-Lys₂-Lys was synthesized as a prototype immunogen used in our immunization of guinea pigs. This octameric antigen contains a small heptalysyl core (<20%) and the bulk (>80%) is formed by a high density of uniform peptide-antigen layered around the core matrix. This design differs from the conventional peptide-carrier conjugate which contains a large protein carrier such as PPD or

KLH and a low density of peptide antigens randomly distributed on the protein carrier surface in an unidentified form.

For the synthesis of octameric HCV peptide immunogen, a combination of Boc-amino acid resin-bound benzhydrylamide and tBoc-chemistry was used. An octameric heptalysyl core resin was prepared by
 5 coupling di-t-Boc Lys onto an extra low loading 0.14 mmole/g MBHA (4-methyl benzhydrylamine) resin on a Biosearch 9500 instrument. During each of the two coupling cycles, di-(Boc)-Lys was used for single coupling followed by two capping reactions (with 0.3 M acetylimidazole in DMF dimethylformamide).

After two additional di-(Boc)-Lys couplings onto the first di-(NH₂) Lys-resin, the substitution level of synthetic octameric resin was determined by ninhydrin test and found to have an appropriate level of -NH₂
 10 groups, as calculated based on a theoretical coupling yield, and was used thereafter for the synthesis of octameric peptide antigens each with a predefined amino acid sequence according to the standard t-Boc chemistry.

Acid-labile tert-butyloxycarbonyl (t-Boc) was used for the protection of N- α amino acid. The following functional side-chain protecting groups were used: O-benzyl for Thr, Ser, Glu and Tyr; N^t-tosyl for Arg;
 15 BOM(i.e. Boc-N^{im}-Benzyloxymethyl-) for His; N'-dichlorobenzyloxycarbonyl for Lys; S-4-methylbenzyl- for Cys; O-cyclohexyl for Asp and CHO for Trp. Successive amino acids were added as dictated by the sequence. The resultant octameric peptidyl resin was cleaved by anhydrous HF [0°C for 1 hr in the presence of 10% (v/v) anisole]. The released octameric antigen was extracted by acetic acid, after two
 20 cycles of ether washings of the cleaved peptidyl resin, and lyophilized to dryness so as to be ready for use as an immunogen. A computer-generated picture of such an octameric immunogen is shown in Fig. 1.

Example 9

Relative (%) Immunoreactivity for Envelope/NS-1 Protein Derived Synthetic Peptides by an Enzyme-linked 25 Immunosorbent Assay

Wells of 96-well plates were coated for 1 hour at 37°C with each of the 21 peptides (designated as 255 A-C; 244 A,B; 254 A-C; 248 A-C; 247 A-E and 246 A-E, synthesized with sequences derived from the envelope/NS-1 region of HCV, at 5 ug/mL at 100 uL per well in 10 mM NaHCO₃ buffer, pH 9.5. The
 30 immunoreactivity of each peptide was measured by an 8 member HCV serum panel (Panel I). All 21 peptides were lacking in immunoreactivity on this standard screening HCV panel. However, peptide 254B was found to have some weak reactivity with one panel member, and upon further testing it also reacted strongly with a sample derived from an anti-HCV positive (positive with peptides VIIIE and IIH) plas-
 35 mapheresis donor with elevated (100 i.u./L) alanine aminotransferase (ALT) enzyme activity. To select a panel of samples with reactivity to peptides from the envelope/NS-1 region, 97 such samples from anti-HCV positive plasmapheresis donors with elevated ALT levels were tested with peptide 254B. One sample had an absorbance of 3.214, and a second sample, 2.184. 17 samples with the greatest reactivity with peptide
 40 254B were chosen to form a third panel (Panel III) to screen for the immunoreactivity of the other 20 peptides from the envelope/NS-1 region. The relative (%) immunoreactivity, using peptide 254B as a standard, is given in Table 8a. The individual absorbance values of each of the 17 samples on the four peptides with the greatest reactivity, i.e. 255C (pep7), 254B (pep8), 247B (pep9), and 246D (pep10), are listed in Table 8b.

Since a unique immunoreactivity pattern with panel III members is observed for each of the four peptides (see the boxed value), all four peptides or their analogues are therefore found to be useful as
 45 antigens for the development of immunoassays designed for the detection and screening for antibodies to HCV, particularly to the envelope/NS-1 associated proteins. This "unique" yet "complementary" immunoreactivity pattern conferred by the four peptides as illustrated in Table 8b further demonstrates that the utility of the peptides as antigens for HCV antibody detection, as immunogens for the development of antibodies to HCV envelope/NS-1 protein, and as vaccines for the protection of HCV infection.

50 Since all four peptides (pep7, pep8, pep9, and pep10) are derived from the variable regions of the HCV envelope/NS-1 proteins, examples of substitution analogues for these four peptides are given in Table 8c based on the amino acid sequence (single letter code) information derived from three different HCV strains.

In addition to screening on Panel III samples, the envelope/NS-1 peptides were also tested against samples from plasmapheresis donors who had elevated ALT levels but were nonreactive on the HCV from
 55 the core (e.g. peptide VIIIE) and NS-4 (e.g. peptide IIH) regions screening EIA. Six of these samples, which may represent early seroconversion samples, were reactive on one or more envelope/NS-1 peptide (Table 8d). The absorbance values on these HCV EIA nonreactive samples are lower than the values found for Panel III samples. In the case of Pep7 and Pep10, their shorter segments, 225B and 246C, respectively,

gave greater immunoreactivity, in contrast to the performance on Panel III.

Table 8a		
Synthetic Peptides with their Amino Acid Sequences Derived from the HCV Envelope /NS-1 Protein Region		
		% Relative Immunoreactivity
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255A	CLTPASATQVRNSTGLYHVTNDPCPNSSIVYEANDAILHTPGCCVPCVREGNVSRC	9.8
255B	INDCPNSSIVYEANDAILHTPGCCVPCVREGNVSRC	18.7
255C (Pep7)	VRNSTGLYHVTNDPCPNSSIVYEANDAILHTPGCCVPCVREGNVSRC	51.4
244A	CLTPASATQVRNSTGLYHVTNDPCPNSSIVYEANDAILHTPGCCVPCVREGNVSRC	3.8
244B	QVAMTPTVATRDGKLPATQLRRHIDLIVGSATLC	5.8
244C	CVREGNVSRCVAMTPTVATRDGKLPATQLRRHIDLIVGSATLC	
254A	SALTYVDLGGSVFLIGQLFTFSRRHNTTGGNCSTYPGHNITGHRMADWNNWSPTA	21.4
254B (Pep8)	TGGNCSTYPGHNITGHRMADWNNWSPTA	100.0
254C	FTFSRRHNTTGGNCSTYPGHNITGHRMADWNNWSPTA	16.7
246A	ALVMAQLLRIPQAILDHIAGAHGVLGAIYFSHVGHNAX	3.1
246B	QILLRIPQAILDHIAGAHGVLGAIYFSHVGHNAX	2.5
246C	ALVMAQLLRIPQAILDHIAGAHGVLGAIYFSHVGHNAX	5.3
247A	QAAARNSGLVSLFTPGAKQHIQLIN	39.0
247B (Pep9)	VDVETIVSGGDAARANSGLVSLFTPGAKQHIQLIN	41.3
247C	VLVLLLFAGVDAETIVSGGDAARANSGLVSLFTPGAKQHIQLIN	21.9
247D	YFSHVGHNAXVLLVLLFAGVDAETIVSGGDAARANSGLVSLFTPGAKQHIQLIN	22.7
247E	LAGIATFSHVGHNAXVLLVLLFAGVDAETIVSGGDAARANSGLVSLFTPGAKQHIQLIN	26.5
246A	TGMAGLIYQNKFNSSGGERLASC	37.5
246B	CHESLNTGMAGLIYQNKFNSSGGERLASC	19.8
246C	INSTALNCNESLNTGMAGLIYQNKFNSSGGERLASC	97.6
246D (Pep10)	WHINSTALNCNESLNTGMAGLIYQNKFNSSGGERLASC	98.6
246E	KQNTQLINTNGSMINSTALNCNESLNTGMAGLIYQNKFNSSGGERLASC	10.5

Table 8b

Absorbance of Envelope/NS-1 Peptides on Selected
Anti-HCV Positive Samples with Elevated ALT Levels

5	Sample	Pep7	Pep8	Pep9	Pep10
	1	1.520	0.475	0.335	1.085
10	2	0.017	0.612	0.009	0.068
	3	0.235	0.774	0.341	0.090
15	4	0.066	0.279	0.268	1.038
	5	0.711	0.076	1.412	0.077
20	6	0.106	0.058	0.027	1.428
	7	0.784	2.184	0.241	3.468
25	8	0.037	0.120	0.055	2.992
	9	0.019	1.597	0.177	0.334
30	10	0.313	3.214	2.564	1.488
	11	0.035	0.025	0.763	0.045
35	12	2.132	1.497	0.160	0.408
	13	2.266	1.573	0.129	0.451
40	14	0.047	1.155	0.170	0.037
	15	0.012	0.053	0.030	2.280
45	16	0.064	2.200	0.039	0.810
	17	0.077	0.541	0.069	0.111

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Table 8c

PEP7 (255C) (J-1)	CL	T	V	P	A	S	A	Y	Q	V	R	N	S	T	G	L	Y	H	V	T	N	D	C	P	M	S	S	I	V	Y	E	A	N	D	A	I	L	M	T	P	G	C	V	P	C	V	R	E	G	N	V	S	R	C
(J-4)	CL	T	I	P	A	S	A	Y	E	V	R	N	V	S	G	I	Y	H	V	T	N	D	C	S	N	S	S	I	V	Y	E	A	A	D	M	I	M	H	T	P	G	C	V	P	C	V	R	E	D	M	S	S	R	C
(HCV-J)	CL	T	I	P	A	S	A	Y	E	V	R	N	V	S	G	I	Y	H	V	T	N	D	C	S	N	S	S	I	V	Y	E	A	A	D	M	I	M	H	T	P	G	C	V	P	C	V	R	E	S	M	F	S	R	C
PEP8 (2548) (J-1)	F	T	F	S	P	R	R	N	W	T	T	Q	G	C	N	C	S	I	Y	P	G	H	I	T	G	N	R	M	A	V	D	M	M	M	W	S	P	T	A															
(J-4)	F	T	F	S	P	R	R	N	E	T	V	O	D	C	N	C	S	I	Y	P	G	H	L	S	G	N	R	M	A	V	D	M	M	M	W	S	P	T	T															
(HCV-J)	F	T	F	S	P	R	R	Y	E	T	V	O	D	C	N	C	S	I	Y	P	G	H	V	S	G	N	R	M	A	V	D	M	M	M	W	S	P	T	T															
PEP9 (2478) (J-1)	V	D	A	E	T	I	V	S	G	G	Q	A	A	R	A	M	S	G	L	V	S	L	F	T	P	G	A	K	Q	M	I	O	L	I	N																			
(J-4)	V	D	A	E	T	Y	T	S	G	G	A	A	S	H	T	T	S	T	L	A	S	L	F	S	P	G	A	S	Q	R	I	O	L	V	M																			
(HCV-J)	V	D	G	H	T	N	V	T	G	G	R	V	A	S	S	T	Q	S	L	V	S	V	L	S	Q	G	P	S	O	K	I	O	L	V	M																			
PEP10 (2460) (J-1)	V	N	I	N	S	T	A	L	N	C	M	E	S	L	N	T	G	W	L	A	G	L	I	T	Q	N	K	F	N	S	S	G	C	P	E	R	L	A	S	C														
(J-4)	V	N	I	N	R	T	A	L	N	C	M	D	S	L	H	T	G	F	L	A	A	L	F	T	H	R	F	N	S	S	G	C	P	E	R	M	A	S	C															
(HCV-J)	V	N	I	N	R	T	A	L	N	C	M	D	S	L	Q	T	G	F	I	A	A	L	F	A	N	R	F	N	A	S	G	C	P	E	R	M	A	S	C															

Examples of substitution analogues of pep7, pep8, pep9 and pep10 are given above based on the amino acid sequence (single letters code) information derived from three representative HCV strains (J-1, J-4 and J). The shared amino acid residues are boxed for purpose of comparison.

Table 8d

Absorbance of Envelope/NS-1 Peptides on Selected Samples
Nonreactive on HCV Core (VIII E) and NS-4 (IIH) Peptides

Sample	255B	255C (Pep7)	254B (Pep8)	246C	246D (Pep10)
1	0.344	0.098	0.173	0.240	0.068
2	0.419	0.346	0.015	0.015	0.028
3	0.403	0.300	0.0111	0.023	0.029
4	0.021	0.021	0.222	0.046	0.049
5	0.300	0.231	0.014	0.009	0.009
6	0.012	0.017	0.044	0.402	0.102

EXAMPLE 10

SYNTHESIS OF OCTAMERIC HCV ENVELOPE/NS-1 PEPTIDE ANTIGENS AS KEY COMPONENTS OF IMMUNOGENS/VACCINES

Four octameric HCV envelope/NS-1 peptide immunogens with a structure of

[Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-
 Arg-Asn-Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-Ser-
 5 Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-
 Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys]₈Lys₄Lys₂Lys
 (octameric pep7); [Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-
 10 Gln-Gly-Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-
 Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala]₈
 Lys₄Lys₂Lys (octameric pep8); [Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-
 15 Gly-Gly-Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-
 Pro-Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn]₈Lys₄Lys₂Lys
 20 (octameric pep9) and [Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-
 Asn-Glu-Ser-Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-
 Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
 25 Cys]₈Lys₄Lys₂Lys (octameric pep10),

are synthesized respectively according to a general chemical synthesis procedure described in Example 8 and used as immunogens in our immunization of guinea pigs and chimpanzees.

30 These octameric peptides are injected as a mixture into healthy, naive animals both intradermally and subcutaneously at a dosage of 25 ug per mixture per kg body weight using 2% alum as an adjuvant. After the initial immunization, these animals are boosted at the same dose once per month for a period of four months. The animals are bled monthly and the collected immune sera are monitored for their anti-HCV envelope/NS-1 immunoreactivity. Six months after the last boost, the immunized chimpanzees are subse-
 35 quently challenged by experimental inoculation with various dosages (e.g. 50 mL) of a proven infectious Factor VIII concentrate known to contain HCV so as to evaluate the efficacy in using a mixture of these octameric envelope/NS-1 peptides as a vaccine for the prevention of HCV infection.

EXAMPLE 11

40 Detection of Antibodies to HCV by a Peptide Based Enzyme Immunoassay (EIA) Using Format C

A total of 221 well-characterized clinical specimens categorized into four groups, (a) to (d), were tested on a representative HCV peptide based EIA with the plates coated with a mixture of peptides IIH, V and
 45 VIIIE at 5, 3 and 2 ug/mL respectively at 100 uL per well (Format C), containing both the HCV core and nonstructural peptides as shown in Table B.

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Table B		
Clinical Group	n	% positive for HCV antibodies
(a) AIDS/ARC patients	63	55.6
(b) HBsAg positive individuals	50	42.0
(c) HBc antibody positive individuals	22	22.7
(d) Individuals with elevated (>100 i.u./L) alanine amino transferase (ALT) enzyme activity	86	91.5

EXAMPLE 12**Detection of Antibodies to HCV by Peptide Based HCV EIA Using Formats 1 to 6**

The following five groups of serum specimens:

- (a) Plasmapheresis donors with elevated (>100 i.u./L) alanine aminotransferase (ALT) enzyme activity (n = 30);
- (b) Blood donors with elevated (>45 i.u./L) ALT enzyme activity (n = 15);
- (c) Chronic NANBH patients (n = 30);
- (d) Other viral infections (n = 11);
- (e) Autoimmune disease patients (n = 9);

were analyzed on representative HCV peptide based EIA kits according to the present invention, with the plates coated at 100 uL per well either with:

- (i) Format 1: peptides VIII E, II H and pep11 at 0.5, 3 and 1 µg/mL each;
- (ii) Format 2: peptides VIII E and pep11 at 0.5 and 1 µg/mL each;
- (iii) Format 3: peptides VIII E, Pep11 and pep8 at 0.5, 1 and 10 µg/mL each;
- (iv) Format 4: peptides VIII E and pep8 at 0.5 and 10 µg/mL each;
- (v) Format 5: peptides VIII E, pep11 and pep12 at 0.5, 1 and 2 µg/mL each;
- (vi) or Format 6: peptides VIII E and pep12 at 0.5 and 2 µg/mL each.

These kits represent core, NS-4 and NS-5 (Format 1), core and NS-5 (Formats 2, 5 and 6), core, NS-5 and env (Format 3) and core and env (Format 4).

The results of testing these 95 well characterized samples on Formats 1 through 6 are presented in Table 9. The results indicate that (30/30) of the samples in group (a) were reactive by Formats 1, 2 and 3; 90% (27/30) reactive by Format 4 and 97% (29/30) reactive by Formats 5 and 6. All samples in groups (b) and (c) were positive on all 6 formats. Groups (a), (b) and (c) were shown to be reactive by Format C described in Example 11.

Three samples in group (d) were reactive by Formats 1 to 4. In contrast, these samples were indicated as negative by Format C. Serum samples "88" and "124" apparently responded to the presence of pep11, and serum sample "VZV2500" was indicated as positive by the presence of pep8 in Formats 4 and 5.

All serum samples in group (c) were negative on all formats, including Format C.

Table 9

Antibody to HCV Detected By Peptide Based EIA Kits
(Absorbance 492nm)

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Sample ID	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6
NRC	0.065	0.075	0.056	0.061	0.060	0.019
WRC	0.650	0.454	0.953	0.967	0.403	0.340
SRC	2.183	1.791	2.580	2.635	1.589	1.331

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a. Plasmapheresis, ALT > 100 i.u./L

1	-13	3.166	3.419	3.255	3.371	3.291	3.255
	-27	1.555	1.548	1.980	2.904	1.152	0.881
	-31	3.479	3.144	3.220	2.332	3.319	2.665
	-32	3.001	3.035	3.112	2.691	3.076	2.986
	-39	3.063	3.041	3.361	2.886	3.190	3.038
	-42	3.198	3.201	3.050	3.227	3.230	3.118
	-47	3.479	3.110	3.251	3.201	3.229	3.068
	-48	3.142	2.795	3.116	2.934	3.076	2.725
	-49	3.417	3.291	3.525	3.451	3.195	3.592
	-52	3.263	3.329	3.202	0.120	3.262	3.453
	-53	3.225	3.145	3.096	0.062	3.358	3.097
	-54	3.271	3.018	3.267	0.153	3.073	3.211
2	-4	1.012	0.881	1.542	1.767	0.807	0.745
	-6	3.229	2.964	3.169	3.052	3.076	2.897
	-9	2.691	2.416	2.766	2.967	2.119	1.844
	-26	3.222	3.055	3.095	3.167	3.195	2.951
	-32	3.226	3.372	3.368	3.194	3.496	3.417
	-33	3.151	2.918	3.147	3.027	3.108	3.129
	-34	3.059	3.021	3.143	3.167	3.145	3.320
	-38	3.241	3.116	2.967	3.055	3.213	3.137
	-41	2.964	2.593	2.841	2.964	2.469	2.252
	-43	3.146	2.092	2.541	2.627	1.999	1.920
	-46	2.927	2.818	2.998	2.983	2.556	2.415
	-58	3.285	3.444	3.218	3.191	3.355	3.095
	-60	3.094	2.975	3.113	3.167	2.683	2.640
	-61	2.784	2.345	2.501	2.751	2.007	2.212
	-62	3.320	3.076	3.095	3.076	3.003	2.787
	-77	0.815	0.682	1.096	0.418	0.164	0.152
	-82	3.020	2.982	1.826	3.001	3.032	2.820
	-83	3.076	2.914	3.049	2.996	2.928	2.808

b. Elevated ALT blood donors (ALT > i.u./L)

ALT	-1	3.017	3.035	3.116	3.165	3.167	2.920
	-2	3.256	3.166	3.165	2.974	3.292	3.091
	-3	3.153	3.328	3.291	3.105	3.203	3.230
	-4	2.969	2.894	3.096	3.144	2.880	2.866
	-5	3.073	2.956	2.968	2.952	3.376	2.985
	-7	3.218	3.020	3.157	2.980	2.951	3.060
	-8	3.074	2.930	3.094	3.197	3.121	3.012
	-10	3.479	3.228	3.226	3.109	3.432	3.952
	-11	3.398	3.283	3.140	3.035	3.285	3.222
	-53	3.330	3.029	3.253	3.290	2.974	3.070
	-56	3.151	3.086	3.176	3.202	3.107	3.085
	-69	3.021	3.170	3.167	3.318	3.019	2.831
	-70	3.074	3.035	2.951	3.073	3.054	3.184
	-71	2.985	2.901	3.080	3.039	2.902	2.900
	-82	3.230	3.120	3.085	2.977	3.298	3.096

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c. <u>Chronic NANBH</u>								
5	N	-2	3.320	3.052	2.981	3.283	3.032	2.999
		-3	3.285	3.036	3.095	3.167	3.077	3.094
		-4	3.117	3.469	3.590	3.291	2.259	3.141
		-7	3.027	3.008	3.061	3.065	2.962	2.806
		-8	3.285	3.146	3.117	3.194	3.122	3.195
10		-9	2.886	3.001	3.072	2.985	2.848	2.859
		-10	2.606	2.268	2.027	0.423	2.338	1.104
		-14	3.054	2.808	2.856	2.995	2.341	2.041
		-23	3.228	3.050	3.067	3.225	3.152	3.109
		-25	3.891	2.462	3.190	3.165	1.982	2.091
15		-27	3.194	2.926	3.165	3.029	3.143	3.168
		-28	3.027	3.106	3.259	3.175	3.176	3.202
		-34	3.057	3.037	3.035	3.144	2.907	2.892
		-36	3.304	3.213	3.000	3.033	3.075	3.115
		-41	3.217	3.283	3.039	3.248	3.290	3.249
20		-42	2.997	2.858	3.196	3.094	3.097	2.805
		-44	3.391	3.477	3.350	3.254	3.353	3.387
		-45	3.318	3.096	2.964	3.250	3.319	3.036
		-49	3.292	3.371	3.416	3.255	3.292	3.370
		-54	3.329	3.294	3.105	3.105	3.177	3.203
25		-57	3.197	3.169	3.221	3.141	3.120	3.018
		-60	3.115	3.035	3.090	3.072	3.096	2.873
		-65	2.020	1.816	1.898	2.376	1.133	1.284
		-67	2.265	1.776	2.356	2.396	1.319	0.911
		-68	3.178	3.177	3.200	3.176	3.530	3.087
30		-69	3.222	3.167	3.165	3.283	3.399	3.097
		-77	1.438	1.346	2.548	2.397	1.055	1.071
		-78	2.457	2.038	2.251	2.300	1.642	1.494
		-79	3.225	3.197	3.076	3.142	3.224	3.169
		-80	3.138	3.074	3.135	3.054	3.137	2.896
d. <u>Other Viral Infections</u>								
35	HAV	-86	0.558	0.316	0.607	0.054	0.037	0.014
		-88	0.018	0.021	0.062	0.054	0.014	0.018
		-92	0.045	0.061	0.058	0.050	0.043	0.007
		-120	0.057	0.076	0.051	0.032	0.051	0.026
		-121	0.052	0.138	0.094	0.065	0.072	0.026
40		-124	0.816	1.178	0.622	0.062	1.082	0.017
		-125	0.014	0.016	0.050	0.031	0.012	0.010
		-126	0.105	0.134	0.109	0.081	0.117	0.068
	EBV	-2331	0.021	0.021	0.023	0.020	0.012	0.012
	VZV-M002		0.035	0.030	0.154	0.108	0.025	0.012
	VZV -2500		0.090	0.138	0.976	0.923	0.084	0.032
e. <u>Autoimmune</u>								
45		-209	0.102	0.079	0.117	0.097	0.066	0.028
		-210	0.002	0.003	0.018	0.011	0.002	0.005
		-211	0.016	0.019	0.134	0.168	0.022	0.016
		-212	0.016	0.020	0.075	0.080	0.019	0.006
		-213	0.008	0.009	0.055	0.076	0.005	0.002
50		-215	0.118	0.095	0.226	0.282	0.093	0.060
		-216	0.039	0.037	0.100	0.105	0.042	0.022
		-217	0.019	0.021	0.068	0.056	0.023	0.012
		-218	0.032	0.022	0.110	0.086	0.059	0.031

55 EXAMPLE 13

Comparison of Test Results Using the Six Peptide Based HCV EIA Formats (1-6) on Random Blood Donors

Random blood donor samples (n=100) were tested by Formats 1 to 6. All 100 samples were negative on Formats 2, 5 and 6. Sample 14 had an absorbance of 0.680 on Format 1, and sample 34 had an absorbance of 0.601 and 0.551 on Formats 3 and 4, respectively. For the calculation of mean absorbance and standard deviation, absorbance values >0.500 were omitted from analysis. Table 10 lists the mean absorbance and standard deviation of the 100 samples on Formats 1-6.

Table 10

Mean Absorbance (A492nm) \pm SD of
100 Random Blood Donors

	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6
Mean	0.040	0.035	0.068	0.061	0.030	0.017
S.D.	0.036	0.029	0.046	0.046	0.039	0.032

EXAMPLE 14

Peptide Analogues from HCV Variant Strains for Subtyping HCV-Reactive Sera

Immunoreactive peptides pep7, pep8, pep9 and pep19 derived from the ENV and NS-1 regions, and their analogues with sequences taken from HCV strains HC-J1, CDC/HCV 1, H, HC-J4, HCV-JH, HCV-J, BK, HC-J6 and HC-J7 are synthesized to have the amino acid sequences according to Table 11. The immunoreactive peptides are coated at 5 μ g/mL at 100 μ L per well in wells of microtiter plates and are used to assay HCV positive sera from Taiwan, Japan, Europe, Australia and North America to classify their HCV reactivity into subtypes e.g., HCV-J1, HC-J4, HC-J6 and HC-J7. These peptides derived from hypervariable regions of HCV are useful to distinguish the subtypes of HCV responsible for the infection.

Table 11

Immunoreactive Pep⁷, Pep⁸, Pep⁹ and Pep¹⁹ and Their
Substitution Analogues Derived from the HCV ENV/NS-1 Regions

		(Pep ⁷ , 255C, aa 184-238)
10	HC-J1	CLTVPASAYQVRNSTGLYHVTNDCPNSSIVYEAHDAILHTPGCVPCVREGNVSRC
	HCV1	-----A-----A---
	HCV-H	-----S-----A-----A---
	HC-J4	---I---E---VS-I---S---A-M-M---D-S---
	HCV-JH	---I---E---VS-I---S---A-V-M-A---N-S---
	HCV-J	---I---E---VS-I---S---A-M-M---S-F---
15	HCV-BK	---T---E-H-VS-I---S-A---A-L-M---S---
	HCV-J6	-I-T-V--AE-K-ISTG-M---T-D--TWQLQA-V--V---EKV--T---
	HCV-J7	-V--V--VE---ISSS-YA---S-N--TWQLTN-V--L---ENDNGTL--
20		(Pep ⁸ , 254B, aa 291-330)
	HC-J1	FTFSPRRHWTTQGCNCSIYPGHITGHRMAWDMMNWSPTA
	HCV1	-----T
	HCV-H	-----D-----
25	HC-J4	-----E-V-D-----LS-----T
	HCV-JH	-----E-V-D-----VS-----
	HCV-J	-----YE-V-D-----VS-----T
	HCV-BK	-----V-L-D-----VS-----T
	HCV-J6	-IV--QH--FV-D-----T-----
30	HCV-J7	-II--E--NF--E-----Q-----L-----L
35		(Pep ⁹ , 247B, aa 381-415)
	HC-J1	VDAETIVSGGQAARAMSGLVSLFTPGAKQNIQLIN
	HCV-J	--GH-H-T--RV-SSTQS---WLSQ-PS-K---V-
	HCV-BK	--GD-H-T--AQ-KTTNR---M-AS-PS-K-----
40		(Peptide 19, 244B, aa 229-272)
	HC-J1	CVREGNVSRCWVAMTPTVATRDGKLPATQLRRHIDLLVGSATLC
	HCV1	-----A-----
	HCV-H	-----A-----V-----T-----
	HC-J4	----D-S-----L---L-A-NASV-T-TI---V-----A-AF-
	HCV-JH	----N-S-----L---L-A-NASV-T-T---V-----T-AF-
45	HCV-J	----S-F-----L---L-A-NSSI-T-TI---V-----A-A--
	HCV-BK	-----S-----L---L-A-NVTI-T-TI---V-----A-AF-
	HCV-J6	-EKV--T---IPVS-N--VQQPGALTQG--T---MV-M-----
	HCV-J7	-ENDNGTL---IQV--N--VKHRGALTHN--T-V-MI-MA--V-

EXAMPLE 15Comparison of Immunoreactivity for NS-5 Protein Derived Synthetic Peptides

Wells of 96-well plates were coated for 1 hour at 37° C with each of the 23 peptides (designated as 259A-259E, 260A-260C, 309A-309C, 310A-310C, 311A-311C, 312A-312C and 314A-314C) synthesized with sequences derived from the NS-5 region, at 5 µg/mL at 100 µL per well in 10 mM NaHCO₃ buffer, pH 9.5.

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Table 12

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Immunoreactivity of a NS-2 Protein-derived Synthetic Peptide

Wells of 96-well plates were coated for 1 hour at 37 ° C with 9 synthetic peptides derived from the NS-2

region of HCV. The results (Table 13) show that peptide 289B (i.e. pep17) was immunoreactive with selected anti-HCV positive samples with elevated ALT levels.

Table 13

Absorbance of NS-2 Peptides on Selected Anti-HCV
Positive Samples with Elevated ALT Levels

Sample	289B
1	0.263
4	0.311
7	0.266
18	0.751

EXAMPLE 17Immunoreactivity of NS-3 Protein-derived Synthetic Peptide with Sera from Individuals with Early HCV Infection

Wells of 96-well plates were coated for 1 hour at 37° C with synthetic peptide 315D (i.e. pep18) derived from the NS-3 region of HCV. The results (Table 14) show that peptide 315D was strongly reactive with two serial samples from a plasmapheresis donor with elevated ALT levels.

Table 14

Absorbance of NS-3 Peptide on Serial Samples from
Plasmapheresis Donor with Elevated ALT Levels

Sample	315D
A	1.983
B	1.890

EXAMPLE 18Detection of Antibodies to HCV NS-1 and ENV Regions by Peptide Based EIA Using Formats 7 and 8

Plasmapheresis samples with elevated ALT levels were analyzed on representative HCV peptide based EIAs according to the present invention with plates coated either with (i) pep1 and pep10C at 10 and 10 µg/mL each (Format 7, NS-1 kit) or (ii) pep7 and pep8 at 10 and 10 µg/mL each (Format 8, ENV kit). The results on HCV positive samples with elevated ALT levels are shown in Table 15, indicating a subpopulation of HCV infected individuals develop specific humoral immune responses directed at unique regions of the NS-1 and ENV proteins.

Table 16

Absorbance (492nm) of Selected Samples with Elevated
ALT Levels on Formats 7 and 8

5

	Sample	Format 7 NS-1	Format 8 ENV
10	1	0.804	1.499
	2	0.707	2.487
	3	0.441	1.649
	4	2.651	2.868
	5	0.064	1.569
	6	0.244	0.790
15	7	0.382	0.692
	8	1.438	1.226
	9	0.304	0.411
	10	0.160	0.282
	11	0.079	0.599
20	12	0.286	0.302
	13	0.045	0.610
	14	3.058	2.862
Cutoff OD _{492nm} = 0.200			

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EXAMPLE 19

30 Synthesis of Substitution Analogues of Octameric HCV Envelope Peptide Antigen as Components of HCV Immunogens/Vaccines

Substitution analogues of octameric HCV envelope pep7, pep8 and pep19 with a structure of:

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- (a) [Cys-Leu-Thr-Ile-Pro-Ala-Ser-Ala-Tyr-Glu-Val-Arg-Asn-
Val-Ser-Gly-Ile-Tyr-His-Val-Thr-Asn-Asp-Cys-Ser-Asn-
5 Ser-Ser-Ile-Val-Tyr-Glu-Ala-Ala-Asp-Val-Ile-Met-His-
Ala-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Asn-Asn-Ser-
10 Ser-Arg-Cys-]₈K₄K₂K (an analogue of octameric pep7 with
sequence taken from HCV-JH);
- (b) [Cys-Ile-Thr-Thr-Pro-Val-Ser-Ala-Ala-Glu-Val-Lys-Asn-
15 Ile-Ser-Thr-Gly-Tyr-Met-Val-Thr-Asn-Asp-Cys-Thr-Asn-
Asp-Ser-Ile-Thr-Trp-Gln-Leu-Gln-Ala-Ala-Val-Leu-His-
Val-Pro-Gly-Cys-Val-Pro-Cys-Glu-Lys-Val-Gly-Asn-Thr-
20 Ser-Arg-Cys-]₈K₄K₂K (an analogue of octameric pep7 with
sequence taken from HCV-J6);
- (c) [Cys-Val-Thr-Val-Pro-Val-Ser-Ala-Val-Glu-Val-Arg-Asn-
25 Ile-Ser-Ser-Ser-Tyr-Tyr-Ala-Thr-Asn-Asp-Cys-Ser-Asn-
Asn-Ser-Ile-Thr-Trp-Gln-Leu-Thr-Asn-Ala-Val-Leu-His-
30 Leu-Pro-Gly-Cys-Val-Pro-Cys-Glu-Asn-Asp-Asn-Gly-Thr-
Leu-Arg-Cys-]₈K₄K₂K (an analogue of octameric pep7 with
sequence taken from HCV-J6);
- (d) [Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Glu-Thr-Val-Gln-Asp-
35 Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Val-Ser-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
40 Ala-]₈K₄K₂K (an analogue of octameric pep8 with
sequence taken from HCV-JH);
- (e) [Phe-Ile-Val-Ser-Pro-Gln-His-His-His-Phe-Val-Gln-Asp-
45 Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-Thr-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
50

- Ala-]₈K₄K₂K (an analogue of octameric pep8 with sequence taken from HCV-J6);
- 5 (f) [Phe-Ile-Ile-Ser-Pro-Glu-Arg-Asn-Phe-Thr-Gln-Glu-Cys-Asn-Cys-Ser-Ile-Tyr-Gln-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-Asp-Met-Met-Leu-Asn-Trp-Ser-Pro-Thr-Leu-
10]₈K₄K₂K (an analogue of octameric pep8 with sequence taken from HCV-J7);
- 15 (g) [Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys-]₈K₄K₂K (Octameric pep19)
- 20 (h) [Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys-Trp-Val-Ala-Leu-Thr-Pro-Thr-Leu-Ala-Ala-Arg-Asn-Ala-Ser-Val-Pro-Thr-Thr-Thr-Leu-Arg-Arg-His-Val-Asp-Leu-Leu-Val-Gly-Thr-Ala-Ala-Phe-Cys-]₈K₄K₂K (an analogue of octameric pep19 with sequence taken from HCV-JH);
- 25 (i) [Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys-Trp-Ile-Pro-Val-Ser-Pro-Asn-Val-Ala-Val-Gln-Gln-Pro-Gly-Ala-Leu-Thr-Gln-Gly-Leu-Arg-Thr-His-Ile-Asp-Met-Val-Val-Met-Ser-Ala-Thr-Leu-Cys-]₈K₄K₂K (an analogue of octameric pep19 with sequence taken from HCV-J6);
- 30 (j) [Cys-Glu-Asn-Asp-Asn-Gly-Thr-Leu-Arg-Cys-Trp-Ile-Gln-Val-Thr-Pro-Asn-Val-Ala-Val-Lys-His-Arg-Gly-Ala-Leu-Thr-His-Asn-Leu-Arg-Thr-His-Val-Asp-Met-Ile-Val-Met-Ala-Ala-Thr-Val-Cys-]₈K₄K₂K (an analogue of octameric pep19 with sequence taken from HCV-J7);
- 35
40
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50 respectively according to a general chemical synthesis procedure described in Example 7 and used as immunogens in our immunization of guinea pigs and chimpanzees.

These octameric peptides are injected as a mixture into healthy, naive animals both intradermally and subcutaneously at a dosage of 25 ug per mixture per kg body weight using 2% alum as an adjuvant. After the initial immunization, these animals are boosted at the same dose once per month for a period of four months. The animals are bled monthly and the collected immune sera are monitored for their anti-HCV envelope/NS-1 immunoreactivity. Six months after the last boost, the immunized chimpanzees are subsequently challenged by experimental inoculation with various dosages (e.g. 50 mL) of a proven infectious Factor VIII concentrate known to contain HCV so as to evaluate the efficacy in using a mixture of these

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octameric envelope peptides as a vaccine for the prevention of HCV infection, initially by the evaluation of several serological/clinical markers, and subsequently, the observation of the appearance of clinical symptoms of NANBH in these animals.

The present invention has been illustrated in the above examples, which are not to be used to limit the
5 scope of the invention.

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SEQUENCE LISTING

```

SEQ ID No.:      241A
5  Sequence Type:  amino acid (AA)
Sequence Length:  37 AA

10  Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
      5              10              15
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
      20              25              30
15  Val-Pro-Ala-Lys-Ser-Val-Cys
      35

```

```

20      SEQ ID No.:          241B
      Sequence Type:        AA
25     Sequence Length:    45 AA

Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-Gln-Gly-Trp-Gly-Pro-Ile-Ser-
                        5                                10                    15
30 Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-
                        20                                25                    30
Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-
                        35                                40                    45
35

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40 SEQ ID No.:      241C
   Sequence Type:   AA
   Sequence Length: 52 AA

45 Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-
      5              10              15
   Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
      20              25              30
50 Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
      35              40              45
   Val-Pro-Ala-Lys-Ser-Val-Cys
      50
55

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5  SEQ ID No.:      231A
   Sequence Type:   AA
   Sequence Length: 26 AA

```

10 Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-
5 10 15
Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
20 25

15

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20      SEQ ID No.:      231B
      Sequence Type:    AA
      Sequence Length:  34 AA

```

25 Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-
5 10 15
Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-
20 25 30
30 Pro-Val-Tyr-Cys

30

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SEQ ID No.:	231C (Pep 1)
Sequence Type:	AA
Sequence Length:	42 AA

40

Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
5 10 15
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
45 20 25 30
Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
35 40

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SEQ ID No.: 231D
 Sequence Type: AA
 Sequence Length: 50 AA

5

Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-Gln-Gly-Trp-Gly-Pro-Ile-Ser-
 5 10 15
 10 Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-
 20 25 30
 Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-
 35 40 45
 15 Gly-Pro-Val-Tyr-Cys
 50

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SEQ ID No.: 231E
 Sequence Type: AA
 Sequence Length: 57 AA

25

Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-
 5 10 15
 30 Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
 20 25 30
 Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
 35 40 45
 35 Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
 50 55

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SEQ ID No.: 232A (Pep 2)
 Sequence Type: AA
 Sequence Length: 26 AA

45

Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-
 5 10 15
 50 Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys
 20 25

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SEQ ID No.: 232B
Sequence Type: AA
Sequence Length: 34 AA

5

Val-Phe-Val-Leu-Asn-Asn-Thr-Arg-Pro-Pro-Leu-Gly-Asn-Trp-Phe-
5 10 15
Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-Gly-Phe-Thr-Lys-Val-Cys-Gly-
10 20 25 30
Ala-Pro-Pro-Cys

15

SEQ ID No.: 232C
Sequence Type: AA
Sequence Length: 42 AA

20

Ser-Trp-Gly-Glu-Asn-Asp-Thr-Asp-Val-Phe-Val-Leu-Asn-Asn-Thr-
25 5 10 15
Arg-Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-
20 25 30
Thr-Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys
30 35 40

35

SEQ ID No.: 232D
Sequence Type: AA
Sequence Length: 50 AA

40

Asp-Arg-Ser-Gly-Ala-Pro-Thr-Tyr-Ser-Trp-Gly-Glu-Asn-Asp-Thr-
5 10 15
Asp-Val-Phe-Val-Leu-Asn-Asn-Thr-Arg-Pro-Pro-Leu-Gly-Asn-Trp-
45 20 25 30
Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-Gly-Phe-Thr-Lys-Val-Cys-
35 40 45
Gly-Ala-Pro-Pro-Cys
50 50

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SEQ ID No.: 233C
Sequence Type: AA
Sequence Length: 42 AA

5

Leu-His-Cys-Pro-Thr-Asp-Cys-Phe-Arg-Lys-His-Pro-Asp-Ala-Thr-
5 50 15
Tyr-Ser-Arg-Cys-Gly-Ser-Gly-Pro-Trp-Ile-Thr-Pro-Arg-Cys-Leu-
10 20 25 30
Val-Asp-Tyr-Pro-Tyr-Arg-Leu-Trp-His-Trp-Pro-Cys
35 40

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SEQ ID No.: 234A
Sequence Type: AA
Sequence Length: 23 AA

20

25 Glu-Ala-Ala-Cys-Asn-Trp-Thr-Arg-Gly-Glu-Arg-Cys-Asp-Leu-Glu-
5 10 15
Asp-Arg-Asp-Arg-Ser-Glu-Leu-Ser
20

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SEQ ID No.: 234B
Sequence Type: AA
Sequence Length: 31 AA

35

40 Val-Gly-Gly-Val-Glu-His-Arg-Leu-Glu-Ala-Ala-Cys-Asn-Trp-Thr-
5 10 15
Arg-Gly-Glu-Arg-Cys-Asp-Leu-Glu-Asp-Arg-Asp-Arg-Ser-Glu-Leu-
20 25 30
45 Ser

50

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SEQ ID No.: 234C
 5 Sequence Type: AA
 Sequence Length: 39 AA

 10 Thr-Ile-Phe-Lys-Ile-Arg-Met-Tyr-Val-Gly-Gly-Val-Glu-His-Arg-
 5 10 15
 Leu-Glu-Ala-Ala-Cys-Asn-Trp-Thr-Arg-Gly-Glu-Arg-Cys-Asp-Leu-
 20 25 30
 15 Glu-Asp-Arg-Asp-Arg-Ser-Glu-Leu-Ser
 35

20
 SEQ ID No.: 272A
 Sequence Type: AA
 25 Sequence Length: 41 AA

 Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-
 5 10 15
 30 Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-
 20 25 30
 Gln-Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser
 35 40

40 SEQ ID No.: 272B
 Sequence Type: AA
 Sequence Length: 55 AA

 45 Thr-Thr-Met-Arg-Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-
 5 10 15
 Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-
 20 25 30
 50 Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-
 35 40 45
 Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser
 50 55

SEQ ID No.: 272C
 5 Sequence Type: AA
 Sequence Length: 66 AA

10 Ala-Val-Asp-Phe-Ile-Pro-Val-Glu-Asn-Leu-Glu-Thr-Thr-Met-Arg-
 5 10 15
 Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-Val-Pro-Gln-
 20 25 30
 15 Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-
 35 40 45
 Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-Gly-Tyr-Lys-Val-
 50 55 60
 20 Leu-Val-Leu-Asn-Pro-Ser
 65

25
 SEQ ID No.: 278A
 Sequence Type: AA
 30 Sequence Length: 29 AA

Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-
 5 10 15
 35 Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala
 20 25

40
 SEQ ID No.: 278B
 Sequence Type: AA
 45 Sequence Length: 36 AA

Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-
 5 10 15
 50 Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-
 20 25 30
 Val-Pro-Ala-Ala-Tyr-Ala
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SEQ ID No.: 278C
 Sequence Type: AA
 5 Sequence Length: 48 AA

Val-Glu-Asn-Leu-Glu-Thr-Thr-Met-Arg-Ser-Pro-Val-Phe-Thr-Asp-
 5 10 15
 10 Asn-Ser-Ser-Pro-Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-
 20 25 30
 Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-
 35 40 45
 15 Ala-Tyr-Ala

20
 SEQ ID No.: 278D
 Sequence Type: AA
 25 Sequence Length: 54 AA

Ala-Val-Asp-Phe-Ile-Pro-Val-Glu-Asn-Leu-Glu-Thr-Thr-Met-Arg-
 5 10 15
 30 Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-Val-Pro-Gln-
 20 25 30
 Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-
 35 40 45
 35 Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala
 50

40
 SEQ ID No.: 275A
 Sequence Type: AA
 45 Sequence Length: 38 AA

Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
 5 10 15
 50 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
 20 25 30
 Ile-Ile-Cys-Asp-Glu-Cys-His-Ser
 35

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SEQ ID No.: 275B
 5 Sequence Type: AA
 Sequence Length: 71 AA

10 Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-
 5 10 15
 Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-
 20 25 30
 15 Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-
 35 40 45
 Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-
 50 55 60
 20 Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser
 65 70

25
 SEQ ID No.: 275C
 Sequence Type: AA
 30 Sequence Length: 94 AA

His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-
 5 10 15
 35 Ala-Ala-Tyr-Ala-Ala-Gln-Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-
 20 25 30
 Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-
 35 40 45
 40 His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-
 50 55 60
 Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-
 65 70 75
 45 Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-
 80 85 90
 Glu-Cys-His-Ser

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SEQ ID No.: 275D
Sequence Type: AA
Sequence Length: 117 AA

5

Thr-Met-Arg-Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-
5 10 15
Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-
10 20 25 30
Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-Gly-
35 40 45
15 Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-Gly-
50 55 60
Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-
65 70 75
20 Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-
80 85 90
Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-
95 100 105
25 Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser
110 115

30

SEQ ID No.: 274A
Sequence Type: AA
Sequence Length: 37 AA

35

Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
5 10 15
40 Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
20 25 30
Asn-Ile-Glu-Glu-Val-Ala-Leu
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SEQ ID No.: 274B
 Sequence Type: AA
 Sequence Length: 64AA

5 Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-
 5 10 15
 10 His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
 20 25 30
 Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-Leu-Ala-Thr-
 35 40 45
 15 Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-
 50 55 60
 Glu-Val-Ala-Leu

20

SEQ ID No.: 274C
 Sequence Type: AA
 Sequence Length: 97 AA

30 Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-
 5 10 15
 Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-
 20 25 30
 35 Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-
 35 40 45
 Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-
 50 55 60
 40 Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
 65 70 75
 Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
 80 85 90
 45 Asn-Ile-Glu-Glu-Val-Ala-Leu
 95

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SEQ ID No.: 274D
Sequence Type: AA
Sequence Length: 120 AA

5

Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-
5 10 15
Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-
10 20 25 30
Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-
35 40 45
Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-
15 50 55 60
Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-
65 70 75
Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-
20 80 85 90
Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-Leu-Ala-Thr-Ala-Thr-Pro-Pro-
95 100 105
Gly-Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-Leu
25 110 115 120

30

SEQ ID No.: 262A
Sequence Type: AA
Sequence Length: 29 AA

35

Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-
5 10 15
Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr
40 20 25

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SEQ ID No.: 262E
 5 Sequence Type: AA
 Sequence Length: 68 AA

10 Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-
 5 10
 His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-
 20 25 30
 15 Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-
 35 40 45
 Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-
 50 55 60
 20 Glu-Cys-His-Ser-Thr-Asp-Ala-Thr
 65

25 SEQ ID No.: 262F
 Sequence Type: AA
 30 Sequence Length: 77 AA

Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-
 5 10 15
 35 Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-
 20 25 30
 Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-
 35 40 45
 40 Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-
 50 55 60
 Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-
 65 70 75
 45 Ala-Thr

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SEQ ID No.: 261A
Sequence Type: AA
5 Sequence Length: 30 AA

Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-Gly-
5 10 15
10 Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-Leu
20 25 30

15
SEQ ID No.: 261B
Sequence Type: AA
20 Sequence Length: 40 AA

Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-
5 10 15
25 Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-
20 25 30
Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-Leu
35 40

35 SEQ ID No.: 261C
Sequence Type: AA
Sequence Length: 50 AA

40 Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-
5 10 15
Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-
20 25 30
45 Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-
35 40 45
Lys-Cys-Asp-Glu-Leu
50

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SEQ ID No.: 261D
Sequence Type: AA
Sequence Length: 73 AA

Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
5 10 15
10 Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
20 25 30
Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-
35 40 45
15 Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-
50 55 60
Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-Leu
65 70

20

SEQ ID No.: 261E
Sequence Type: AA
Sequence Length: 97 AA

30 Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-
5 10 15
Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-
20 25 30
35 Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-Leu-Ala-Thr-Ala-Thr-Pro-
35 40 45
Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-
50 55 60
40 Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-
65 70 75
Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-
80 85 90
45 Lys-Lys-Lys-Cys-Asp-Glu-Leu
95

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SEQ ID No.: 261F
Sequence Type: AA
5 Sequence Length: 121 AA

Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
5 10 15
10 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
20 25 30
Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-
35 40 45
15 Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-
50 55 60
Leu-Val-Val-Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-
65 70 75
20 Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-
80 85 90
Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-
95 100 105
25 Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-
110 115 120
Leu

30

SEQ ID No.: 279A (Pep 3)
35 Sequence Type: AA
Sequence Length: 37 AA

40 Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-
5 10 15
His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
20 25 30
45 Asp-Gln-Ala-Glu-Thr-Ala-Gly
35

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SEQ ID No.: 279B
 5 Sequence Type: AA
 Sequence Length: 42 AA

Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-
 10 5 10 15
 Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-
 20 25 30
 Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly
 15 35 40

20 SEQ ID No.: 279E
 Sequence Type: AA
 Sequence Length: 58 AA

25 Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
 5 10 15
 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
 30 20 25 30
 Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-
 35 40 45
 Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly
 35 50 55

40 SEQ ID No.: 255A
 Sequence Type: AA
 45 Sequence Length: 35 AA

Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-
 5 10 15
 50 Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-
 20 25 30
 Asn-Val-Ser-Arg-Cys
 35

SEQ ID No.: 255B
 5 Sequence Type: AA
 Sequence Length: 45 AA

10 Val-Arg-Asn-Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-
 5 10 15
 Asn-Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-
 20 25 30
 15 Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-
 35 40 45

20
 SEQ ID No.: 255C (Pep 7)
 Sequence Type: AA
 25 Sequence Length: 55 AA

Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-Ser-Thr-
 5 10 15
 30 Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-
 20 25 30
 Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-
 35 40 45
 35 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys
 50 55

40
 SEQ ID No.: 244A
 Sequence Type: AA
 45 Sequence Length: 35 AA

Cys-Trp-Val-Ala-Met-Thr-Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-
 5 10 15
 50 Leu-Pro-Ala-Thr-Gln-Leu-Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-
 20 25 30
 Ser-Ala-Thr-Leu-Cys
 35

5 SEQ ID No.: 244B
 Sequence Type: AA
 Sequence Length: 44 AA

10 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
 5 10 15
 Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
 20 25 30
 Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys
 15 35 40

20 SEQ ID No.: 254A
 Sequence Type: AA
 Sequence Length: 30 AA

25 Thr-Gln-Gly-Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-
 5 10 15
 His-Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala
 30 20 25 30

35 SEQ ID No.: 254B (Pep 8)
 Sequence Type: AA
 Sequence Length: 40 AA

40 Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
 5 10 15
 Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
 45 20 25 30
 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala
 35 40

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SEQ ID No.: 254C
Sequence Type: AA
5 Sequence Length: 50 AA

Cys-Gly-Ser-Val-Phe-Leu-Ile-Gly-Gln-Leu-Phe-Thr-Phe-Ser-Pro-
5 10 15
10 Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-Cys-Ser-Ile-Tyr-Pro-
20 25 30
Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-
35 40 45
15 Trp-Ser-Pro-Thr-Ala
50

20

SEQ ID No.: 248A
Sequence Type: AA
25 Sequence Length: 25 AA

Asp-Met-Ile-Ala-Gly-Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-
5 10 15
30 Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys
20 25

35

SEQ ID No.: 248B
Sequence Type: AA
40 Sequence Length: 35 AA

Gln-Leu-Leu-Arg-Ile-Pro-Gln-Ala-Ile-Leu-Asp-Met-Ile-Ala-Gly-
5 10 15
45 Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-Tyr-Phe-Ser-Met-Val-
20 25 30
Gly-Asn-Trp-Ala-Lys
50 35

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5      SEQ ID No.:      248C
      Sequence Type:    AA
      Sequence Length:  40 AA

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Ala-Leu-Val-Met-Ala-Gln-Leu-Leu-Arg-Ile-Pro-Gln-Ala-Ile-Leu-
5 10 15
10 Asp-Met-IleAla-Gly-Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-
20 25 30
Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys
35 40

20	SEQ ID No.:	247A
	Sequence Type:	AA
	Sequence Length:	25 AA

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Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-	
5 10 15	
Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn	
20 25	

35 **SEQ ID No.: 247B (Pep 9)**
 Sequence Type: AA
 Sequence Length: 35 AA

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40      Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-
           5                               10                               15
      Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-
           20                               25                               30
45      Ile-Gln-Leu-Ile-Asn
           35

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SEQ ID No.: 247C
Sequence Type: AA
Sequence Length: 45AA

5

Val-Leu-Val-Val-Leu-Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-
5 10 15
Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-
10 20 25 30
Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
35 40 45

15

SEQ ID No.: 247D
Sequence Type: AA
Sequence Length: 55 AA

20

Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys-Val-Leu-Val-Val-Leu-
25 5 10 15
Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-
20 25 30
Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-
30 35 40 45
Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
50 55

35

SEQ ID No.: 247E
Sequence Type: AA
Sequence Length: 60 AA

40

Leu-Ala-Gly-Ile-Ala-Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys-
45 5 10 15
Val-Leu-Val-Val-Leu-Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-
20 25 30
Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-
50 35 40 45
Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
55 50 55 60

55

SEQ ID No.: 246A
 5 Sequence Type: AA
 Sequence Length: 25 AA

10 Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-
 5 10 15
 Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys
 20 25

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20 SEQ ID No.: 246B
 Sequence Type: AA
 Sequence Length: 31 AA

25 Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-
 5 10 15
 Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
 20 25 30
 30 Cys

35 SEQ ID No.: 246C
 Sequence Type: AA
 Sequence Length: 38 AA

40

Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-
 5 10 15
 45 Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-
 20 25 30
 Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys
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SEQ ID No.: 246D (Pep 10)
 Sequence Type: AA
 5 Sequence Length: 40 AA

Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-
 5 10 15
 10 Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-
 20 25 30
 Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys
 35 40

15

20 SEQ ID No.: 246E
 Sequence Type: AA
 Sequence Length: 52 AA

25 Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn-Thr-Asn-Gly-Ser-Trp-His-Ile-
 5 10 15
 Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-Trp-
 20 25 30
 30 Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-Cys-
 35 40 45
 Pro-Glu-Arg-Leu-Ala-Ser-Cys

35

40 SEQ ID No.: 314A (Pep 16)

Sequence Length: 30 AA

45

Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-
 5 10 15
 50 Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu
 20 25 30

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SEQ ID No.: 314B
Sequence Type: AA
Sequence Length: 38 AA

5

Ser-Glu-Cys-Thr-Thr-Pro-Cys-Ser-Gly-Ser-Trp-Leu-Arg-Asp-Ile-
5 10 15
Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-Ser-Asp-Phe-Lys-Thr-Trp-Leu-
10 20 25 30
Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu
35

15

SEQ ID No.: 314C
Sequence Type: AA
Sequence Length: 47 AA

20

Leu-Arg-Arg-Leu-His-Gln-Trp-Ile-Ser-Ser-Glu-Cys-Thr-Thr-Pro-
5 10 15
Cys-Ser-Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-
20 25 30
Val-Leu-Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-
30 35 40 45
Gln-Leu

35

SEQ ID No.: 312A
Sequence Type: AA
Sequence Length: 22 AA

40

Asp-Phe-His-Tyr-Val-Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-Cys-
5 10 15
Pro-Cys-Gln-Val-Pro-Ser-Pro
20

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78

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SEQ ID No.: 311B
Sequence Type: AA
Sequence Length: 42 AA

5

Asp-Gly-Val-Arg-Leu-His-Arg-Phe-Ala-Pro-Pro-Cys-Lys-Pro-Leu-
5 10 15
Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-His-Glu-Tyr-Pro-
10 20 25 30
Val-Gly-Ser-Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-Asp
35 40

15

SEQ ID No.: 311C
Sequence Type: AA
Sequence Length: 54 AA

20

Cys-Gln-Val-Pro-Ser-Pro-Glu-Phe-Phe-Thr-Glu-Leu-Asp-Gly-Val-
5 10 15
Arg-Leu-His-Arg-Phe-Ala-Pro-Pro-Cys-Lys-Pro-Leu-Leu-Arg-Glu-
20 25 30
Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-His-Glu-Tyr-Pro-Val-Gly-Ser-
30 35 40 45
Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-Asp
50

35

SEQ ID No.: 260A
Sequence Type: AA
Sequence Length: 23 AA

40

Ser-Ser-Ser-Ala-Ser-Gln-Leu-Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-
5 10 15
Cys-Thr-Ala-Asn-His-Asp-Ser-Pro
20

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SEQ ID No.: 260B
Sequence Type: AA
5 Sequence Length: 35 AA

Arg-Arg-Leu-Ala-Arg-Gly-Ser-Pro-Pro-Ser-Val-Ala-Ser-Ser-Ser-
5 10 15
10 Ala-Ser-Gln-Leu-Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-Cys-Thr-Ala-
20 25 30
Asn-His-Asp-Ser-Pro
35

15

20 SEQ ID No.: 260C (Pep 4)
Sequence Type: AA
Sequence Length: 46 AA

25 Asp-Pro-Ser-His-Ile-Thr-Ala-Glu-Ala-Ala-Gly-Arg-Arg-Leu-Ala-
5 10 15
Arg-Gly-Ser-Pro-Pro-Ser-Val-Ala-Ser-Ser-Ser-Ala-Ser-Gln-Leu-
20 25 30
30 Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-
35 40 45
Pro

35

40 SEQ ID No.: 259B
Sequence Type: AA
Sequence Length: 35 AA

45 Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-
5 10 15
Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-
20 25 30
50 Glu-Glu-Asp-Glu-Arg
35

55

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SEQ ID No.: 259C (Pep 5)

Sequence Type: AA

Sequence Length: 44 AA

5

Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
5 10 15

10 Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-
20 25 30

Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg
35 40

15

SEQ ID No.: 259D

20

Sequence Type: AA

Sequence Length: 50 AA

25 Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-
5 10 15

Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-
20 25 30

30 Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-
35 40 45

Glu-Glu-Asp-Glu-Arg
50

35

SEQ ID No.: 259E (Pep 12)

40

Sequence Type: AA

Sequence Length: 55 AA

45 Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
5 10 15

Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
20 25 30

50 Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
35 40 45

Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg
50 55

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SEQ ID No.: 310A
 5 Sequence Type: AA
 Sequence Length: 26 AA

Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-
 10 5 10 15
 Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg
 20 25

15

SEQ ID No.: 310B
 20 Sequence Type: AA
 Sequence Length: 35 AA

Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-
 25 5 10 15
 Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-
 20 25 30
 Arg-Lys-Ser-Arg-Arg
 30 35

35

SEQ ID No.: 310C (Pep 13)
 Sequence Type: AA
 40 Sequence Length: 47 AA

Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-
 5 10 15
 45 Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
 20 25 30
 Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-
 35 40 45
 50 Arg-Arg

55

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SEQ ID No.: 309D (Pep 11)
Sequence Type: AA
Sequence Length: 60 AA

5 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
5 10 15
Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
20 25 30
10 Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
35 40 45
Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
50 55 60

15

20 SEQ ID No.: 309E
Sequence Type: AA
Sequence Length: 72 AA

25 Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-
5 10 15
Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-Trp-Ala-Arg-
20 25 30
30 Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-Lys-Pro-Asp-
35 40 45
Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-
50 55 60
35 Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
65 70

40

45 SEQ ID No.: Pep 6
Sequence Type: AA
Sequence Length: 37 AA

50 Asp-Pro-Gln-Ala-Arg-Val-Ala-Ile-Lys-Ser-Leu-Thr-Glu-Arg-Leu-
5 10 15
Thr-Val-Gly-Gly-Pro-Leu-Thr-Asn-Ser-Arg-Gly-Glu-Asn-Cys-Gly-
20 25 30
Tyr-Arg-Arg-Cys-Arg-Ala-Ser
35

55

SEQ ID No.: Pep 17

5 Sequence Type: AA

Sequence Length: 45 AA

10 Gly-Pro-Ala-Asp-Gly-Met-Val-Ser-Lys-Gly-Trp-Arg-Leu-Leu-Ala-
5 10 15
Pro-Ile-Thr-Ala-Tyr-Ala-Gln-Gln-Thr-Arg-Gly-Leu-Leu-Gly-Cys-
20 25 30
15 Ile-Ile-Thr-Ser-Leu-Thr-Gly-Arg-Asp-Lys-Asn-Gln-Val-Glu-Gly
35 40 45

20 SEQ ID No.: Pep 18

Sequence Type: AA

25 Sequence Length: 39 AA

Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-
5 10 15
30 Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-
20 25 30
Glu-Leu-Ala-Ala-Lys-Leu-Val-Ala-Leu
35

40 SEQ ID No.: Pep 19

Sequence Type: AA

Sequence Length: 44 AA

45 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
5 10 15
Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
20 25 30
50 Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys
35 40

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SEQ ID No.: VIIIE
Sequence Type: AA
Sequence Length: 61 AA

5

Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-Thr-Asn-
5 10 15
Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gly-Gln-Ile-Val-
10 20 25 30
Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-
35 40 45
Arg-Ala-Thr-Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-
15 50 55 60
Arg

20

SEQ ID No.: IIH
Sequence Type: AA
25 Sequence Length: 47 AA

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-
30 5 10 15
Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-
20 25 30
Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-
35 40 45
Gly-Leu

40

SEQ ID No.: V
Sequence Type: AA
45 Sequence Length: 40 AA

Lys-Gln-Lys-Ala-Leu-Gly-Leu-Leu-Gln-Thr-Ala-Ser-Arg-Gln-Ala-
5 10 15
50 Glu-Val-Ile-Ala-Pro-Ala-Val-Gln-Thr-Asn-Trp-Gln-Lys-Leu-Glu-
20 25 30
Thr-Phe-Trp-Ala-Lys-His-Met-Trp-Asn-Phe
35 40

55

SEQ ID No.: **Page XX**

Sequence Type: AA

[Peptide]₁₆ Lys₈ Lys₄ Lys₂ Lys-Y
decahexyl peptide

SEQ ID No.: **Pep XXI**

Sequence Type: AA

[Peptide]₈ Lys₄ Lys₂ Lys-Y
octameric peptide

SEQ ID No.: Pep XXII

Sequence Type: AA

[Peptide]₄ Lys₂ Lys-Y
tetrameric peptide

SEQ ID No.: Pep XXIII

Sequence Type: AA

[Peptide]₂ Lys-Y
dimeric peptide

wherein the peptide in Pep XX, Pep XXI, Pep XXII and Pep XIII is selected from the group consisting of (a) to (j):

40 (a) Cys-Leu-Thr-Ile-Pro-Ala-Ser-Ala-Tyr-Glu-Val-Arg-Asn-Val-Ser-
5 10 15
Gly-Ile-Tyr-His-Val-Thr-Asn-Asp-Cys-Ser-Asn-Ser-Ser-Ile-Val-
20 25 30
45 Tyr-Glu-Ala-Ala-Asp-Val-Ile-Met-His-Ala-Pro-Gly-Cys-Val-Pro-
35 40 45
Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys
50 55

(b) Cys-Ile-Thr-Thr-Pro-Val-Ser-Ala-Ala-Glu-Val-Lys-Asn-Ile-Ser-
5 Thr-Gly-Tyr-Met-Val-Thr-Asn-Asp-Cys-Thr-Asn-Asp-Ser-Ile-Thr-
Trp-Gln-Leu-Gln-Ala-Ala-Val-Leu-His-Val-Pro-Gly-Cys-Val-Pro-
Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys

(c) Cys-Val-Thr-Val-Pro-Val-Ser-Ala-Val-Glu-Val-Arg-Asn-Ile-Ser-
15 5 10 15
Ser-Ser-Tyr-Tyr-Ala-Thr-Asn-Asp-Cys-Ser-Asn-Asn-Ser-Ile-Thr-
20 25 30
Trp-Gln-Leu-Thr-Asn-Ala-Val-Leu-His-Leu-Pro-Gly-Cys-Val-Pro-
35 40 45
20 Cys-Glu-Asn-Asp-Asn-Gly-Thr-Leu-Arg-Cys
50 55

25 (d) Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Glu-Thr-Val-Gln-Asp-Cys-Asn-
5 10 15
Cys-Ser-Ile-Tyr-Pro-Gly-His-Val-Ser-Gly-His-Arg-Met-Ala-Trp-
20 25 30
30 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala
35 40

35 (e) Phe-Ile-Val-Ser-Pro-Gln-His-His-His-Phe-Val-Gln-Asp-Cys-Asn-
5 10 15
Cys-Ser-Ile-Tyr-Pro-Gly-Thr-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
20 25 30
40 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala
35 40

45 (f) Phe-Ile-Ile-Ser-Pro-Glu-Arg-Asn-Phe-Thr-Gln-Glu-Cys-Asn-Cys-
5 10 15
Ser-Ile-Tyr-Gln-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-Asp-
20 25 30
50 Met-Met-Leu-Asn-Trp-Ser-Pro-Thr-Leu
35

(g) Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
 5 10 15
Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
5 20 25 30
Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys
 35 40

10 (h) Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys-Trp-Val-Ala-Leu-Thr-
5 10 15
Pro-Thr-Leu-Ala-Ala-Arg-Asn-Ala-Ser-Val-Pro-Thr-Thr-Thr-Leu-
20 25 30
15 Arg-Arg-His-Val-Asp-Leu-Leu-Val-Gly-Thr-Ala-Ala-Phe-Cys
35 40

20 (i) Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys-Trp-Ile-Pro-Val-Ser-
5 10 15
Pro-Asn-Val-Ala-Val-Gln-Gln-Pro-Gly-Ala-Leu-Thr-Gln-Gly-Leu-
20 25 30
25 Arg-Thr-His-Ile-Asp-Met-Val-Val-Met-Ser-Ala-Thr-Leu-Cys
35 40

30 (j) Cys-Glu-Asn-Asp-Asn-Gly-Thr-Leu-Arg-Cys-Trp-Ile-Gln-Val-Thr-
5 10 15
Pro-Asn-Val-Ala-Val-Lys-His-Arg-Gly-Ala-Leu-Thr-His-Asn-Leu-
20 25 30
35 Arg-Thr-His-Val-Asp-Met-Ile-Val-Met-Ala-Ala-Thr-Val-Cys
35 40

40 Claims

1. A peptide composition comprising a peptide having an amino acid sequence selected from the group consisting of:

- (a) Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-
Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-
5 Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-
Val-Tyr-Cys-X;
Pep1
- 10 (b) Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-
Ser-Thr-Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys-X;
Pep2
- 15 (c) Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-
Glu-Leu-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-
20 Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-X;
Pep3
- (d) Asp-Pro-Ser-His-Ile-Thr-Ala-Glu-Ala-Ala-Gly-Arg-Arg-
25 Leu-Ala-Arg-Gly-Ser-Pro-Pro-Ser-Val-Ala-Ser-Ser-Ser-
Ala-Ser-Gln-Leu-Ser-Ala-Pro-Ser-Leu-Lys-Ala-Thr-Cys-
Thr-Ala-Asn-His-Asp-Ser-Pro-X;
30 Pep4
- (e) Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-
35 Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-
Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-
Glu-Glu-Asp-Glu-Arg-X;
40 Pep5
- (f) Asp-Pro-Gln-Ala-Arg-Val-Ala-Ile-Lys-Ser-Leu-Thr-Glu-
Arg-Leu-Thr-Val-Gly-Gly-Pro-Leu-Thr-Asn-Ser-Arg-Gly-
45 Glu-Asn-Cys-Gly-Tyr-Arg-Arg-Cys-Arg-Ala-Ser-X;
Pep6
- 50
- 55

- (g) Cys-Leu-Thr-Val¹-Ala²-Ser-Ala-Tyr-Gln-Val-Arg-Asn-
 Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-
 Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-
 Thr-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-
 Ser-Arg-Cys-X;
 Pep7
- (h) Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
 Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
 Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
 Ala-X;
 Pep8
- (i) Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-
 Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-Gly-
 Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn-X;
 Pep9
- (j) Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-
 Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Glu-His-
 Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
 Cys-X;
 Pep10
- (k) Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
 Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
 Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
 His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
 Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;
 Pep11
- (l) Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-
 Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
 Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-
 Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
 Asp-Glu-Arg-X;
 Pep12

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- (m) Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-
Glu-Asn-Lys-Val¹Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-
Val-Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-
5 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-X;
Pep13
- (n) Cys-Lys-Pro-Leu-Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-
10 Gly-Leu-His-Glu-Tyr-Pro-Val-Gly-Ser-Gln-Leu-Pro-Cys-
Glu-Pro-Glu-Pro-Asp-X;
Pep14
- (o) Glu-Glu-Tyr-Val-Glu-Ile-Arg-Gln-Val-Gly-Asp-Phe-His-
Tyr-Val-Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-Cys-Pro-
20 Cys-Gln-Val-Pro-Ser-Pro-X;
Pep15
- (p) Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp¹-Trp-Ile-Cys-Glu-
25 Val-Leu-Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-
Met-Pro-Gln-Leu-X;
Pep16
- (q) Gly-Pro-Ala-Asp-Gly-Met-Val-Ser-Lys-Gly-Trp-Arg-Leu-
Leu-Ala-Pro-Ile-Thr-Ala-Tyr-Ala-Gln-Gln-Thr-Arg-Gly-
35 Leu-Leu-Gly-Cys-Ile-Ile-Thr-Ser-Leu-Thr-Gly-Arg-Asp-
Lys-Asn-Gln-Val-Glu-Gly-X;
Pep17
- (r) Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-
40 Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-
Lys-Lys-Cys-Asp-Glu-Leu-Ala-Ala-Lys-Leu-Val-Ala-Leu-X;
45 Pep18
- (s) Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-
50 Met-Thr-Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-
Ala-Thr-Gln-Leu-Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-
Ser-Ala-Thr-Leu-Cys-X;
55 Pep19

wherein X is -OH or -NH₂, and analogues, segments, mixtures, conjugates and polymers thereof.

2. A peptide composition according to claim 1 wherein the peptide comprises:

5

Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-
Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-
10 Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-
Thr-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-
Ser-Arg-Cys-X;

15

pep7

wherein X is -OH or -NH₂, and analogues, segments, conjugates and polymers thereof.

- 20 3. A peptide composition according to Claim 1 wherein the peptide comprises:

25

Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

30

pep8

wherein X is -OH or -NH₂, and analogues, segments, conjugates and polymers thereof.

- 35 4. A peptide composition according to Claim 1 wherein the peptide comprises:

40

Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-
Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Glu-His-
Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
Cys-X;

45

pep10

wherein X is -OH or -NH₂, and analogues, segments, conjugates and polymers thereof.

- 50 5. A peptide composition according to Claim 1 wherein the peptide comprises:

55

5 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
10 Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

pep11

wherein X is -OH or -NH₂, and analogues, segments, conjugates and polymers thereof.

15 6. A peptide composition comprising a mixture of Peptides VIIIE and pep11 wherein Peptide VIIIE is:

20 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-
Thr-Asn-Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-
Gly-Gln-Ile-Val-Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-
Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-Arg-Lys-Thr-Ser-
25 Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X;

(VIIIE)

30 and pep11 is:

35 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
40 Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

Pep11

45 wherein X is -OH or NH₂, and analogues thereof.

7. A peptide composition according to claim 6 further comprising Peptide IIH having an amino acid sequence:

50 Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-
Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-
Leu-Pro-Tyr-Ile-Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-
55 Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X;

(IIH)

wherein X -OH or -NH₂ and analogues thereof.

8. A peptide composition according to Claim 6 further comprising pep8 having an amino acid sequence:

5 Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
10 Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

Pep8

15 wherein X is -OH or -NH₂, and analogues thereof.

9. A peptide composition according to Claim 6 further comprising pep12 having an amino acid sequence:

20 Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-
Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
25 Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-
Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
Asp-Glu-Arg-X;

Pep12

wherein X is -OH or -NH₂ and analogues thereof.

- 35 10. A peptide composition comprising a mixture of Peptides VIIIE and pep8 wherein Peptide VIIIE is

Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-
40 Thr-Asn-Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-
Gly-Gln-Ile-Val-Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-
Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-Arg-Lys-Thr-Ser-
45 Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X;

(VIIIE)

50 and pep8 is:

55

Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
5 Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

Pep8

and wherein X is -OH or -NH₂ and analogues thereof.

11. A peptide composition according to Claim 1 comprising a mixture of pep7 and pep8, wherein pep7 is:

Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-
Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-
20 Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-
Thr-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-
Ser-Arg-Cys-X;

Pep7

and pep8 is:

Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
35 Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

Pep8

and wherein X is -OH or -NH₂, and analogues thereof.

12. A peptide composition according to Claim 1 comprising a mixture of pep1 and pep10, wherein pep1 is:

Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-
Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-
50 Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-
Val-Tyr-Cys-X;

Pep1

pep10 is:

Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-
 Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Glu-His-
 Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
 Cys-X;

Pep10

and wherein X is -OH or -NH₂, and analogues thereof.

13. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 1 in an immunoassay procedure.
14. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 6 in an immunoassay procedure.
15. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 7 in an immunoassay procedure.
16. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 8 in an immunoassay procedure.
17. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 9 in an immunoassay procedure.
18. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using on effective amount of a peptide composition according to Claim 10 in an immunoassay procedure.
19. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of a peptide composition according to Claim 11 in an immunoassay procedure.
20. A method of detecting antibodies to HCV or diagnosis of HCV infection or NANBH by using an effective amount of peptide composition according to Claim 12 in an immunoassay procedure.
21. A peptide immunogen comprising a polymeric peptide selected from the group consisting of:
 [Peptide]₁₆ Lys₈ Lys₄ Lys₂ Lys-Y
 [Peptide]₈ Lys₄ Lys₂ Lys-Y
 [Peptide]₄ Lys₂ Lys-Y
 [Peptide]₂ Lys-Y
 wherein Y is -OH₂, -NH₂ or amino acid with no side chain functional group and the peptide is selected from the group consisting of:

- (a) Cys-Leu-Thr-Ile-Pro-Ala-Ser-Ala-Tyr—Glu-Val-Arg-Asn-
Val-Ser-Gly-Ile-Tyr—His-Val-Thr-Asn-Asp-Cys-Ser-Asn-
5 Ser-Ser-Ile-Val-Tyr—Glu-Ala-Ala-Asp-Val-Ile-Met-His-
Ala-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Asn-Asn-Ser-
Ser-Arg-Cys;
10 (b) Cys-Ile-Thr-Thr-Pro-Val-Ser-Ala-Ala-Glu-Val-Lys-Asn-
Ile-Ser-Thr-Gly-Tyr—Met-Val-Thr-Asn-Asp-Cys-Thr-Asn-
15 Asp-Ser-Ile-Thr-Trp-Gln-Leu-Gln-Ala-Ala-Val-Leu-His-
Val-Pro-Gly-Cys-Val-Pro-Cys-Glu-Lys-Val-Gly-Asn-Thr-
Ser-Arg-Cys;
20 (c) Cys-Val-Thr-Val-Pro-Val-Ser-Ala-Val-Glu-Val-Arg-Asn-
Ile-Ser-Ser-Ser-Tyr—Tyr—Ala-Thr-Asn-Asp-Cys-Ser-Asn-
Asn-Ser-Ile-Thr-Trp-Gln-Leu-Thr-Asn-Ala-Val-Leu-His-
25 Leu-Pro-Gly-Cys-Val-Pro-Cys-Glu-Asn-Asp-Asn-Gly-Thr-
Leu-Arg-Cys;

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- (d) Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Glu-Thr-Val-Gln-Asp-
Cys-Asn-Cys-Ser-Ile-Tyr—Pro-Gly-His-Val-Ser-Gly-His-
5 Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala;
- (e) Phe-Ile-Val-Ser-Pro-Gln-His-His-His-Phe-Val-Gln-Asp-
10 Cys-Asn-Cys-Ser-Ile-Tyr—Pro-Gly-Thr-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala;
- (f) Phe-Ile-Ile-Ser-Pro-Glu-Arg-Asn-Phe-Thr-Gln-Glu-Cys-
15 Asn-Cys-Ser-Ile-Tyr—Gln-Gly-His-Ile-Thr-Gly-His-Arg-
20 Met-Ala-Trp-Asp-Met-Met-Leu-Asn-Trp-Ser-Pro-Thr-
Leu;
- (g) Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-
25 Met-Thr-Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-
Ala-Thr-Gln-Leu-Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-
Ser-Ala-Thr-Leu-Cys;
- (h) Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys-Trp-Val-Ala-
30 Leu-Thr-Pro-Thr-Leu-Ala-Ala-Arg-Asn-Ala-Ser-Val-Pro-
35 Thr-Thr-Thr-Leu-Arg-Arg-His-Val-Asp-Leu-Leu-Val-Gly-
Thr-Ala-Ala-Phe-Cys;
- (i) Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys-Trp-Ile-Pro-
40 Val-Ser-Pro-Asn-Val-Ala-Val-Gln-Gln-Pro-Gly-Ala-Leu-
Thr-Gln-Gly-Leu-Arg-Thr-His-Ile-Asp-Met-Val-Val-Met-
Ser-Ala-Thr-Leu-Cys; and
- (j) Cys-Glu-Asn-Asp-Asn-Gly-Thr-Leu-Arg-Cys-Trp-Ile-Gln-
45 Val-Thr-Pro-Asn-Val-Ala-Val-Lys-His-Arg-Gly-Ala-Leu-
50 Thr-His-Asn-Leu-Arg-Thr-His-Val-Asp-Met-Ile-Val-Met-
Ala-Ala-Thr-Val-Cys;

and analogues thereof.

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Fig. 1



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Applicant: **United Biomedical Inc.
2 Nevada Drive
Lake Success New York 11042(US)**

Inventor: **Chang Yi Wang
159 Hill Park Avenue
Great Neck, N.Y.11021(US)**
Inventor: **Hoseln, Barbara
196 E. 75th Street
New York, N.Y.10021(US)**

Representative: **Hansmann, Axel et al
Albert-Rosshaupter-Strasse 65
W-8000 München 70(DE)**

Synthetic peptides specific for the detection of antibodies to HCV, diagnosis of HCV infection and prevention thereof as vaccines.

The present invention relates to peptides which are immunoreactive to antibodies to HCV or NANBHV and a method of detecting the presence of HCV or NANBHV antibodies in body fluids by using the peptides as the antigen. The peptides are selected from both the envelope and non-structural protein regions of the HCV or NANBHV. The detection method includes enzyme linked immunosorbent assay or other immunoassay procedures. The peptides and conjugates or polymers thereof are also useful as immunogens in generating high titer antibodies to HCV or in vaccines.

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Page 1

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
A, D	EP-A-0 318 216 (CHIRON CORPORATION) 31 May 1989 "the whole document"	1-21	C07K7/10 A61K39/12 G01N33/569
A, D	SCIENCE, vol. 244, 21 April 1989, LANCASTER, PA US pages 362 - 364; G KUD ET AL.: 'an assay for circulating antibodies to a major etiologic virus of human non-a, non-b hepatitis' "the whole disclosure"	1-21	
A	SCIENCE, vol. 244, 21 April 1989, LANCASTER, PA US pages 359 - 362; QL CHOO ET AL.: 'isolation of a cDNA clone derived from a blood-borne non-a, non-b viral hepatitis genome' "the whole disclosure"	1-21	
A	NUCLEIC ACIDS RESEARCH, vol. 17, no. 24, 1989, ARLINGTON, VIRGINIA US pages 10367 - 10372; Y KUBO ET AL.: 'a cDNA fragment of hepatitis c virus isolated from an implicated donor of post-transfusion non-a, non-b hepatitis in Japan' "the whole document"	1-21	TECHNICAL FIELDS SEARCHED (Int. Cl.5) C07K A61K G01N
D, P, X	EP-A-0 388 232 (CHIRON CORPORATION) 19 September 1990 "the whole document"	1-21	
D, T	EP-A-0 442 394 (UNITED BIOMEDICAL INC.) 21 August 1991 "the whole document"	1-21	
		-/-	
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 22 JUNE 1992	Examiner ma stupzo
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons A : member of the same patent family, corresponding document			



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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
A	JOURNAL OF IMMUNOLOGICAL METHODS, vol. 124, no. 1, 1989, NEW YORK US pages 53 - 61; J P TAN AND F ZAVALA: 'multiple antigen peptide. A novel approach to increase detection sensitivity of synthetic peptides in solid-phase immunoassays' * the whole document *	21	
			TECHNICAL FIELDS SEARCHED (Int. Cl.5)
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 22 JUNE 1992	Examiner rasturzo
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons A : member of the same patent family, corresponding document			

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(54) **Synthetic peptides specific for the detection of antibodies to HCV, diagnosis of HCV infection and prevention thereof as vaccines**

Synthetische Peptide, spezifisch zum Nachweis von gegen HCV gerichteten Antikörpern zur Diagnose der HCV-Infektion und zum Vorkommen davon als Impfstoffe

Peptides synthétiques, spécifiques pour le dépistage d'anticorps contre HCV, pour la diagnose de l'infection causée par HCV et sa prévention en tant que vaccins

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(73) Proprietor: United Biomedical, Inc.
Hauppauge, New York 11788 (US)

(72) Inventors:
• Chang Yi Wang
Great Neck, N.Y. 11021 (US)
• Hoseln, Barbara
New York, N.Y. 10021 (US)

(74) Representative:
Hansmann, Axel, Dipl.-Wirtsch.-Ing. et al
Patent- und Rechtsanwälte
Hansmann, Vogeser, Dr. Boecker,
Alber, Dr. Strych, Liedl
Albert-Rosshaupter-Strasse 65
81369 München (DE)

(56) References cited:
EP-A- 0 318 216 EP-A- 0 388 232
EP-A- 0 442 394

- SCIENCE. vol. 244, 21 April 1989, LANCASTER, PA US pages 362 - 364; G KUO ET AL.: 'an assay for circulating antibodies to a major etiologic virus of human non-a, non-b hepatitis'
- SCIENCE. vol. 244, 21 April 1989, LANCASTER, PA US pages 359 - 362; QL CHOO ET AL.: 'Isolation of a cDNA clone derived from a blood-borne non-a, non-b viral hepatitis genome'
- NUCLEIC ACIDS RESEARCH. vol. 17, no. 24, 1989, ARLINGTON, VIRGINIA US pages 10367 - 10372; Y KUBO ET AL.: 'a cDNA fragment of hepatitis c virus isolated from an implicated donor of post-transfusion non-a, non-b hepatitis in Japan'
- JOURNAL OF IMMUNOLOGICAL METHODS. vol. 124, no. 1, 1989, NEW YORK US pages 53 - 61; J P TAM AND F ZAVALA: 'multiple antigen peptide. A novel approach to increase detection sensitivity of synthetic peptides in solid-phase immunoassays'

Remarks:

The file contains technical information submitted after the application was filed and not included in this specification

EP 0 468 527 B1

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Description

INTRODUCTION

5 [0001] The present invention relates to peptides specific for the diagnosis and prevention of hepatitis C virus (HCV) infection, or non-A non-B hepatitis (NANBH). More particularly, the present invention is directed to synthetic peptides which are specific for the detection of antibodies to HCV in body fluids and immunoassays using the same. The invention also includes the use of the synthetic peptides in compositions as antigens for eliciting the production of monoclonal and polyclonal antibodies against HCV and as immunogens in vaccines for the prevention of NANBH or HCV infection.

10 [0002] In recent years, non-A, non-B hepatitis (NANBH) has become the most common form of post-transfusion hepatitis. Studies involving the experimental inoculation of chimpanzees provided evidence that the infectious agent was a lipid-containing virus resembling members of the Togaviridae family.

[0003] Recently, this etiological agent, termed hepatitis C virus (HCV) has been shown to be an RNA virus with a genome size of ~ 10 kilobases encoding a single polyprotein which can be further processed into several structural and nonstructural proteins (1-4). Additional computer-assisted protein analysis demonstrates that HCV shares sequence similarity with the polyproteins of animal pestiviruses and flaviviruses as well as members of two plant virus supergroups (5).

[0004] More recently, a number of reports have led to an increasingly coherent understanding of the function of various regions of the virus and of the relationships among genomic fragments isolated from variants or closely related viruses.

20 [0005] A summary of the HCV structure, beginning at the N terminus of the virus, follows. The HCV comprises a structural protein region and nonstructural (NS) protein regions. The structural protein region is further divided into capsid and envelop proteins. The NS protein regions are further divided into NS-1 to NS-5 regions (3).

[0006] The postulated capsid region (AA1-AA120) has been shown to contain highly immunoreactive conserved epitopes with enhanced sensitivity in the detection of hepatitis C infection (6-8). The region appears to consist of two segments of equal length (AA1-61, AA62-AA120), which are homologous to one another, perhaps as a result of a gene duplication, and are also homologous to the N terminal core region of yellow fever virus (9), also a flavivirus (Table 1A). Both halves, as represented by peptides VIIIE (AA2-AA62) and IXD (AA65-AA119), disclosed in application serial No. 558,799, have been shown to be immunoreactive. A genomic fragment of a NANBH virus cloned by Arima et al. (10), designated clone 2, contains a Gly-Pro-Arg-Leu-Gly sequence identical to residues 39-43 in peptide VIIIE (Table 1B), placing this clone 2 fragment in the putative core region of a related virus. Two other sequences from NANBH viruses, cloned by Reyes et al. (11) and by Arima et al. (clone 1) (12), show sequence similarities with the capsid region of yellow fever virus (Table 1C). Thus, there appears to be a number of related viruses, all of which have highly immunogenic capsid regions, as evidenced by the ease of cloning. Variants of hepatitis C (J, J-1, J-4) are also highly conserved in this region (2-4), so the other clones mentioned by Arima et al. may be from different viruses, rather than from variants of HCV.

[0007] Mishiro and colleagues have isolated a cDNA clone from the plasma of a chimpanzee infected with NANBH which codes for a host cellular sequence bearing an epitope which is reactive with sera from individuals who are PCR positive for HCV (13). The sequence of the immunoreactive peptide (GOR epitope) is not encoded by HCV and was reported not to resemble a published sequence of HCV spanning three-quarters of the genome (1) or the 5'-terminal sequence of HCV (2) covering the upstream quarter of the genome. However, inspection of the GOR epitope sequence revealed 47% homology with an N-terminal fragment covered by peptide VIIIE described in UBI Applications Serial No. 558,799. Lesser degrees of homology were obtained from comparison with the N-terminus of the yellow fever virus capsid protein (33%) (9) and the protein segment corresponding to clone 1 of Arima et al. (37.5%) (12) (See Table 1D).

45 [0008] The presence of antibodies which are cross-reactive with the GOR epitope sequence in HCV infected individuals may be explained by structural similarity of the GOR epitope with the corresponding region of the HCV capsid protein. Compared with anti-C100, antibodies to the C100 region, previously identified by Houghton et al.; antibodies to peptide VIIIE share the following characteristics with anti-GOR: they both are present in some but not all anti-C100 positive sera; they can be detected in anti-C100 negative sera from both acute and chronic NANBH patients; they appear earlier than anti-C100 in the seroconversion series; they are detected in more seroconversion panels than anti-C100 (13); and they are present in 1-2% of normal controls and 15-20% of HBsAg positive individuals. Early NANBH assays reported to react with host-determinant cytoplasmic antigens may in fact have detected anti-HCV capsid protein cross-reactivity.

55 [0009] The postulated envelope (env) region consists of amino acids 120 to 400. The env glycoproteins of flaviviruses are key targets for immunization because the env region is a major antigen of free viral particles and plays a central role in flavivirus biology. The env region mediates binding to cell receptors and probably facilitates fusion to membranes. It also induces protective immune responses after vaccination or natural infection with a flavivirus (14, 15) and stimulates cell-mediated immunity (16). The type-specific epitopes on env are the ones most closely associated with protective

immune responses to flaviviruses (17-19). There are a number of hypervariable regions in the HCV env region, based on a comparison of US and Japanese strains (2), which may indicate epitopes for strain specific reactivity.

5 [0010] The non-structural protein NS-1, in addition to the small M protein of the envelope, has been shown to contribute to protective immunity in dengue fever (20,21). Inspection of sequences and hydrophobicity profiles shows that the HCV NS-1 region contains two similar domains (Table 1E). A dominant motif in this region is cysteine pairs separated by five or more amino acids.

[0011] The NS-2 region is of unknown function and little has been reported on its characteristics.

10 [0012] By analogy with yellow fever virus, the HCV NS-3 region may contain protease activity required for viral replication (22). A trypsin-like serine protease active site has been localized in yellow fever virus by means of site-directed mutagenesis of NS-3 to a catalytic triad consisting of His-53, Asp-77 and Ser-138. The corresponding region in HCV is the N-terminal third of NS-3, with the critical residues being His-1103, Asp-1127 and Ser-1188. The remainder of the HCV NS-3 region consists of a region which shows immunoreactivity. This region appears to consist of three subregions homologous to one another (Table 1F) and these subregions bear a distant relationship to the repeated segments of the NS-1 region.

15 [0013] The most widely studied region to date is the NS-4 nonstructural region. Although its function is unknown, it contains highly immunoreactive regions, primarily in the region designated as C100 by Houghton et al. (1), which became the basis for a HCV diagnostic test using recombinant technology. A high degree of structural homology is observed between part of the C100 HCV sequence with a corresponding region in the yellow fever virus (Table 1G). While this region detects antibody to the virus primarily responsible for NANBH (23), experimentally it has been shown 20 in prior United Biomedical Inc.'s application Serial No. 558,799 and numerous recent reports that there are shortcomings in both sensitivity and specificity in the tests relying on the C100 polypeptide as an antigen. However, synthetic peptides from the NS-4 region described in prior application Serial No. 558,799 overcome the problem of non-specific reactivity.

25 [0014] The nonstructural region proximal to the C terminus of HCV is NS-5, the site of polymerase (pol) activity. The Gly-Asp-Asp sequence in this region is conserved across many viruses (11). Maeno et al. have isolated a clone corresponding to a sequence upstream of the pol site in the NS-5 region which is immunoreactive and which reacts specifically with sera from patients in the chronic phase of NANBH (24).

30 [0015] Through an extensive series of experiments involving serological validation using select specimens chosen from the screening of thousands of sera with hundreds of carefully designed synthetic peptides, we have further characterized the capsid protein related immunoreactive peptides and have identified additional immunoreactive epitopes contained within the envelope, NS-1, NS-2, NS-3, and NS-5 protein regions.

35 [0016] Synthetic peptides have been increasingly used to map antigenic or immunogenic sites on the surface of proteins, an approach recently termed "site-directed-serology". We, at United Biomedical, have taken this approach to identify and characterize highly antigenic epitopes on the envelope and core proteins of HIV and to develop sensitive and specific immunoassays for the detection of antibodies to HIV (previously designated HTLV-III) (25-27). See U.S. Patent 4,735,896, issued April 5, 1988 and (U.S. Patent 4,879,212 issued Nov. 7, 1989, the contents of which are, hereby, fully incorporated by reference (28,29). Subsequently, a series of finely mapped and well-characterized HTLV- 40 I/II related synthetic peptides were employed in the development of synthetic peptide-based diagnostic assays for the detection of HTLV-I/II antibodies in infected individuals (30,31). See also U.S. Patent 4,833,071 issued May 23, 1989, U.S.S.N. 07/297,635 filed January 13, 1989 and USSN 07/469,294 filed January 24, 1990. These assays have provided superior sensitivity, excellent specificity, and, in certain cases, an unmatched capability to differentiate infections between two closely related viruses, thus overcoming many of the existing problems associated with biologically-derived tests based on either viral lysates or recombinant DNA-derived proteins.

45 [0017] From EP-A-0 388 232 a recombinant polynucleotide comprising sequences derived from HCV cDNA is known including the application of these sequences and polypeptides in immunoassays, probe diagnostics, anti-HCV antibody production, PCR technology and recombinant DNA technology. Disclosed are also immunogenic polypeptides encoded within clones containing HCV cDNA, methods for purifying an immunogenic HCV polypeptide and antisense polynucleotides derived from HCV cDNA.

[0018] It is, therefore, an objective of the present invention to employ the identified and characterized immunoreactive HCV peptides in the development of a detection or diagnostic procedure to identify and monitor HCV infection.

50 [0019] A further objective is to chemically synthesize a test reagent which can then be used to detect the presence of antibodies to HCV in body fluids and to diagnose NANBH.

[0020] Another objective is to develop a vaccine which, when introduced into healthy mammals, including humans, will stimulate production of efficacious antibodies to HCV, thereby providing protection against HCV infection.

55 [0021] A further objective is to provide a synthetic immunogen which can be used in mammals for the development of monoclonal and polyclonal antibodies to HCV.

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Amino acid sequences (single letter code) derived from the corresponding N-terminal capsid protein of the Yellow fever Virus (AA2-AA68, upper line; Ref. 9) and the Hepatitis C Virus (AA2-AA64, middle line; and AA63-AA119, lower line; Ref. 2) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

STIPKOR
: :
QPIPKV - RRPEGR - TWAQPCYPWPLYGNEGCGWAGULLSP - P
EPTIR - R LCP - RLGRPALMAVE :
P - R - R - GP - RLGR - ATRTY :
RGR - PSW - G - P - YD

Amino acid sequence (single letter code) derived from a segment of Arima et al.'s MAMINV-protein clone 2 (upper line; Ref. 10) is aligned with segments of the N-terminal capsid protein of the Hepatitis C virus (AA2-AAS2, middle line; and AA3-AA111, lower line) for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

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KA:GK:TL-GVNVVRG-V-RSL:SNKIKOKY:KOL:SGVPSRG-VGFI:FM:LTGK:IA:MLK:LV-
K:NM-K:ILN-LRKSAT-KV-SS-KYKIK-KL-SGVA-SVL-V-GAT:F--L-GSTA:A:ASDEQLADKO:
K:GE:SNCEAENDTIN:KOR--R:YKEREK:YAT:IN:PGK:K:KPAV-GRI-K:NM:NR:EG:KD-A:YOR-KR-RE

Amino acid sequence (single letter code) derived from the N-terminal capsid protein of the Yellow fever virus (AAS-AA69, upper line; Ref. 9), another MAMBV sequence cloned by Reyes et al. (AA1-AA55, middle line; Fig. 3, Ref. 11) and a third MAMBV sequence cloned by Arino et al. (AAS-AA66, lower line; Ref. 12) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

Table 10

GOR Epitope Sequence (Ref.13)

AA4-AA19 Segment of MCV Capsid Peptide VIIIIE
of prior application serial no. 558,799
Arism et al. Clone 1 (AA22-AA37, Ref.12)

Yellow Fever Virus (AA3-AA19, Ref.9)

Amino acid sequences (single letter code) derived from the GOR epitope (upper line; Ref.139), a segment of the MCV capsid peptide VIII_E representing MCV AA4-AA19 of prior application (second line), A422-A437 of the MATHNV sequence (clone 1) reported by Arias et al (third line; Ref. 12) and a segment of the Yellow Fever Virus N-terminal capsid protein (AA2-AA19, Ref.9) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

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Table 1E

MCV-MS1(J-1)	C R R - L -	T D F D Q G W G P I S Y A N G S G P D O R P Y C -	W M Y P P K P C G - I V - P A - -	K S V C G P V Y C
	D R S G A P T - Y - - S W G E N D T D V F V L M N T R P P L G M U - F G - -	C T U M N S T G F T E - V C G A P P C		

Amino acid sequences (single letter code) derived from two segments of the MCV MS-1 protein (upper line, AA59-AA508; and lower line, AA520-AA569); are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

MCV-MS-1(J-1)	C R R L T D F D Q G W G P I S Y A N G S G P D O R P Y C M N Y P P K P C G I V P A K S V C G P V Y C
MCV-MS-1(J-4)	C R P I D W F A Q G W G P I T Y T E P D S P D O R P Y C M N Y A P R P C G I V P A S Q V C G P V Y C
MCV-MS-1(J)	C R P I D E F A Q G W G P I T M D M P E S D O R P Y C M N Y A P R P C G I V P A S Q V C G P V Y C

Amino acid sequences (single letter code) derived from three MCV strains (J-1, J-4 and J) for a segment of the MS-1 protein (AA59-AA508); are aligned for comparison of homology.

MCV-MS-1(PT)	D R S G A P T Y S U G E N D T V D F V L M N T R P P L G M U F G C T M N N S T G F T E K V C G A P P C
MCV-MS-1(J)	D R F G A P T Y S U G E N E T D V L L L S N T R P P O G M U F G C T M N N S T G F T E K T C G G P P C

Amino acid sequences (single letter code) derived from two MCV strains (PT and J) for a segment of the MS-1 protein (AA520-AA569) are aligned for comparison of homology.

MCV-MS-1(PT)	D R S G A P T Y S U G E N D T V D F V L M N T R P P L G M U F G C T M N N S T G F T E K V C G A P P C
MCV-MS-1(J)	D R F G A P T Y S U G E N E T D V L L L S N T R P P O G M U F G C T M N N S T G F T E K T C G G P P C

Amino acid sequences (single letter code) derived from two MCV strains (PT and J) for a segment of the MS-1 protein (AA520-AA569) are aligned for comparison of homology.

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Table 1f

MCV-NS3	D	F	I	P	-	-	V	E	M	L	E	T	M	R	S	P	V	F	T	D	M	S	-	P	P	V	-	-	V	-	P	Q	-	S	-	F	Q	V	A	H	L	M	A	P	T	G	S	G	K	S	T	-	-	K	V	P			
	P	M	I	R	T	G	V	R	T	I	-	T	T	G	-	S	P	I	-	T	T	-	S	T	Y	G	K	F	L	A	D	-	G	G	C	S	G	G	A	Y	D	-	-	I	I	I	C	D	E	C	M	S	T	U	A	T			
	P	M	I	E	E	-	V	A	-	L	S	T	T	G	E	I	P	-	F	-	Y	G	K	A	-	I	P	-	L	E	V	I	E	G	G	R	N	L	I	F	C	-	-	H	S	K	E	K	C	D	E	L	-	A	-	-	A	K	L

Amino acid sequences (single letter code) derived from three segments of the HCV NS-3 protein (AA1194-1241, upper line; AA1276-AA1324, middle line; and AA1360-AA1407, lower line) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

MCV	V	V	L	A	T	A	T	P	P	G	S	V	T
BVD	V	V	A	M	T	A	T	P	A	G	S	V	T
MOG	V	V	A	M	T	A	T	P	A	G	T	V	T
YFV	T	I	L	N	T	A	T	P	P	G	T	S	D

MCV	Q	R	G	R	T	G	R	G	K	P	G	I	-	Y	R
BVD	Q	R	G	R	V	G	R	V	K	P	G	R	Y	R	
MOG	Q	R	G	R	V	G	R	V	K	P	G	R	Y	R	
YFV	Q	R	G	R	I	G	R	-	M	P	M	R	D	G	D

Multiple alignment of two highly conserved segments encoded within the NS-3 protein region (single letter code) of HCV (AA1344-AA1356, upper Table; and AA1486-AA1500, lower Table respectively), Bovine Diarrhea Virus (BVD, AJ2025-AA2037; AA2181-AA2196), Hog Cholera Virus (MOG, AA1866-AA1898; AA-2042-AA2057) and Yellow Fever Virus (YFV AA1800-AA1812; AA1944-AA1956) are aligned for comparison of homology.

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Table 1G

V	I	Y	G	R	Y	V	L	S	C	K	P	A	I	P	R	R	E	V	L	Y	B	E	F	D	E	M	Q	N	L	P	Y	I	E	N	G	M	L	A	..		
:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	:	
A	A	E	Y	L	Y	B	L	S	E	L	P	R	F	..	L	A	K	G	G	E	A	R	D	T	I	S	V	F	L	S	E	E	G	S	R	A	..
E	N	F	K	K	A	L	..	G	L	L	G	I	A	S	R	G	A	E	Y	I	
E	N	T	I	V	N	L	F	I	L	A	G	L	L	..	I	..	S	G	N	

Amino acid sequences (single letter code) derived from a segment of the HCV NS-4 protein and a corresponding segment of the Yellow Fever Virus NS-4 protein (lower line, AA2109-AA2176, Ref.9) are aligned for comparison of homology. Identical amino acid matches are boxed with a solid line, while matches scored as similar by the PAM-250 matrix are connected with a colon. Dashes represent spaces between adjacent amino acids that have been inserted to optimize the alignment.

REFERENCES

[0022]

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1. Houghton M, Choo Q-L, Kuo G: NANBV diagnostics and vaccines. EPO 0318216A1 (1989).
2. Okamoto H, Okada S, Sugiyama S, Yotsumoto S, Tanaka T, Yoshizawa H, Tsuda F, Miyakawa Y, Mayumi M: The 5' terminal sequence of the hepatitis C virus genome. *Jpn. J. Exp. Med.* 60:167 (1990).
3. Houghton M, Choo Q-L, Kuo G: NANBH diagnostics and vaccines. EPO 0388232A1 (1990).
4. Kato N, Hijikata M, Ootsuyama Y, et al: Molecular cloning of the human hepatitis C virus genome from Japanese patients with non-A, non-B hepatitis. *Proc. Natl. Acad. Sci. USA* 87:9524 (1990).
5. Miller RH, Purcell RH: Hepatitis C virus shares amino acid sequence similarity with pestiviruses and flaviviruses as well as members of two plant virus supergroups. *Proc. Natl. Acad. Sci. USA* 87:2057 (1990).
6. Hosein B, Fang CT, Zhang ML, et al: Improved serodiagnosis of hepatitis C virus infection with synthetic peptide

- antigen from capsid protein. Proc. Natl. Acad. Sci. USA 88:3647 (1991).
7. UBI HCV EIA Product Insert. (1990).
8. Okamoto H, Munekata E, Tsuda F, et al: Enzyme-linked immunosorbent assay for antibodies against the capsid protein of hepatitis C virus with a synthetic oligopeptide. Jap. J. Exp. Med. 60:223 (1990).
- 5 9. Rice CM, Lencheo EM, Eddy SR, et al: Nucleotide sequence of yellow fever virus: Implications for flavivirus gene expression and evolution. Science 229:726 (1985).
10. Arima T, Takamizawa A, Mori C, et al: A lambda gt11-cDNA clone specific for chronic hepatitis C generated from pooled serum presumably infected by hepatitis C virus. Gastroenterologia Japonica 24:545 (1989).
11. Reyes GR, Purdy MA, Kim J, et al: Isolation of a cDNA from the virus responsible for enterically transmitted non-A, non-B hepatitis. Science 247:1335 (1990).
12. Arima T, Nagashima H, Murakami S, et al: Cloning of a cDNA associated with acute and chronic hepatitis C infection generated from patients serum RNA. Gastroenterologia Japonica 24:540 (1989).
13. Mishiro S, Hoshi Y, Takeda K, et al: Non-A, non-B hepatitis specific antibodies directed at host-derived epitope: Implication for an autoimmune process. Lancet 336:1400 (1990).
- 15 14. Brinton MA: in The Togaviridae and Flaviviridae, ed. Schlesinger S and Schlesinger MJ. Plenum Press, NY pp. 327-374 (1986).
15. Mandl CW, Guirakhoo F, Holzmann H, Heinz FX, Kunz C: Antigenic structure of the flavivirus envelope protein E at the molecular level, using tick-borne encephalitis virus as a model. J. Virol. 63:564 (1989).
16. Bray M, Falgout B, Zhao B, et al: in Vaccines '89. Modern Approaches to New Vaccines Including Prevention of AIDS, ed Lerner RA, Ginsberg H, Chanock RM and Brown F. Cold Spring Harbor Laboratory, NY, pp357-362 (1989).
- 20 17. Roehrig JT, Hunt AR, Johnson J, Mathews JH: ibid. pp347-350 (1989).
18. Rothman AL, Kurane J, Ennis FA: ibid. pp363-366 (1989).
19. Roehrig JT: in The Togaviridae and Flaviviridae, ed Schlesinger S and Schlesinger MJ. Plenum Press NY, pp251-278 (1986).
- 25 20. Bray M, Meu R, Lai CJ: Meeting on Modern Approaches to New Vaccines Including Prevention of AIDS. Cold Spring Harbor Laboratory, Sept 12-16, 1990. Abst 70.
21. Falgout B, Bray M, Schlesinger JJ, Lai CJ: Immunization of mice with recombinant vaccinia virus expressing authentic dengue virus nonstructural protein NS1 protects against lethal dengue virus encephalitis. J. Virol. 64: 4356 (1990).
- 30 22. Chambers TJ, Weir RC, Grakoui A, et al: Evidence that the N-terminal domain of nonstructural protein NS-3 from yellow fever virus is a serine protease responsible for site-specific cleavages in the viral polyprotein. Proc. Natl. Acad. Sci. USA 87:8898 (1990).
23. Kuo G., Choo Q-L, Alter HJ, et al: An assay for circulating antibodies to a major etiologic virus of human non-A, non-B hepatitis. Science 244:362 (1989).
- 35 24. Maeno M, Kaminaka K, Sugimoto H, et al: A cDNA clone closely associated with non-A, non-B hepatitis. Nucleic Acids Res. 18:2685 (1990).
25. Wang CY: Synthetic-peptide-based immunodiagnosis of retrovirus infections: Current status and future prospects. In: Synthetic Peptides in Biotechnology, ed. Mizrahi A, Advances in Biotechnological Processes, 10:131 (1988).
- 40 26. Wang JG, Steel S, Wisniewolski R, Wang CY: Detection of antibodies to HTLV-III using a synthetic peptide of 21 amino acid residues corresponding to a highly antigenic segment of gp41 envelope protein. Proc. Natl. Acad. Sci. USA 83:6159 (1986).
27. Wang CY: European Patent Application Publication: EPO 0328403 (1989). Synthetic peptides related to the HIV-gp120 env protein, and their use.
- 45 28. Wang CY, Wang JG: U.S. Patent 4879212 (1989). Peptide composition and method for the detection of antibodies to HTLV-III.
29. Wang CY, Wang JG: U.S. Patent 4735896 (1988). Synthetic peptide and process of using same for the detection and diagnosis of AIDS and pre-AIDS conditions.
- 50 30. Wang CY, Wang JG, Walters DW: U.S. Patent 4833071 (1989). Peptide composition as antigen for detection of antibodies to HTLV-I, as a vaccine for ATL, and methods therefore.
31. Wang CY: U.S.S.N. 07/297635. Synthetic peptide compositions with immunoreactivities to antibodies to HTLV.

BRIEF DESCRIPTION OF THE INVENTION

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[0023] According to the present invention, a series of synthetic peptides representing immunoreactive regions of the postulated envelope protein and nonstructural proteins NS-1, NS-2, NS-3 and NS-5 of the hepatitis C virus (HCV), each arranged in a specific sequence, has been identified and made by solid phase peptide synthesis. These peptides

have been found to be useful for the detection of antibodies to HCV in sera and body fluids and for the diagnosis of non-A, non-B hepatitis (NANBH). Because of their immunoreactivity, it is expected that these peptides are also useful in stimulating production of antibodies to HCV in healthy mammals such as Balb/C mice, and in a vaccine composition to prevent HCV or NANBHV infection.

5 [0024] According to the present invention, a peptide composition useful for the detection of antibodies to HCV and diagnosis of NANBH comprises a peptide from the envelope, NS-1, NS-2, NS-3 and NS-5 regions of the HCV represented by the following sequences:

10 Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala-X

15 Pep8

20 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
25 Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-
X

Pep11

30 Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
35 Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X

40 Pep12

[0025] These 3 peptides are in addition to Peptide VIIIE, a peptide from the structural protein region, and Peptide IIH, peptides from the non-structural protein region which have also been found to be reactive and useful for the de-
45 tection of antibodies to HCV and diagnosis of NANBH.

[0026] Peptide VIIIE has the following sequence:

50 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thy-Lys-Arg-Asn-Thr-Asn-Arg-
Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gly-Gln-Ile-Val-Gly-Gly-
Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-
55 Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X,

[0027] Peptide IIH has the following sequence:

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-Glu-
 Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-Glu-Gln-
 Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X

wherein X is -OH or -NH₂ and analogues, segments, mixtures, conjugates and polymers thereof.

[0028] Further, according to the present invention, the peptides by themselves, or when coupled to a protein or a polymeric carrier of homo or hetero dimers or higher oligomers by the use of homo or hetero functional multivalent cross linking reagents, or when directly synthesized and conjugated to a branching polyvalent lysine resin, can be used to elicit the production of antibodies to HCV in healthy mammals, including humans.

[0029] The method comprises introducing an effective amount of the peptide composition containing each of the individual peptides, analogues or segments or a mixture or a combination thereof, or in a polymeric form, into the body of a healthy mammal by intraperitoneal or subcutaneous injection.

[0030] Vaccines containing the peptides according to the present invention as the key immunogen may also be prepared as described above or by known methods. It is expected that such vaccine compositions may be useful to prevent HCV infection or NANBH.

20 BRIEF DESCRIPTION OF DRAWING

[0031] Fig. 1 is a photograph of a computer-generated structure of an octameric peptide immunogen.

25 DETAILED DESCRIPTION OF THE INVENTION

[0032] In accordance with the present invention, three peptides and their analogues have been selected from the nonstructural regions of HCV and chemically synthesized. These peptides including their analogues are useful for the detection of antibodies to HCV in body fluids, the diagnosis of NANBH, and for the vaccination of healthy mammals by stimulating the production of antibodies to HCV. These peptides are arranged in the following sequences:

Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
 Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala-X

Pep8

Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
 Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
 Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
 Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-
 X

Pep11

Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-X

Pep12

[0033] These 3 peptides are in addition to Peptide VIIIE, a peptide from the structural protein region, and Peptide IIH, peptides from the non-structural protein region which have also been found to be reactive and useful for the detection of antibodies to HCV and diagnosis of NANBH.

[0034] Peptide VIIIE has the following sequence:

Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-Thr-Asn-Arg-
Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gly-Gln-Ile-Val-Gly-Gly-
Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-
Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X,

[0035] Peptide IIH has the following sequence:

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-Glu-
Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-Glu-Gln-
Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X

Wherein X is -OH or -NH₂ and analogues, segments, mixtures, conjugates, and polymers thereof.

[0036] These peptides may comprise combinations or segments, i.e. longer or shorter peptide chains by having more amino acids added to the terminal amino acids, or by amino acids removed from either terminal end.

[0037] These peptides may also comprise analogues to accommodate strain-to-strain variations among different isolates of HCV. HCV is indicated to have frequent mutations. Therefore, it is expected that variant strains, such as PT, J, J-1 and J-4 (1-4) exist. Adjustments for conservative substitutions and selection among the alternatives where non-conservative substitutions are involved, may be made in the prescribed sequences (e.g. see Table 1E for possible amino acid substitutions in the hypervariable regions of the envelope and NS-1 proteins). These analogues of the synthetic peptides may therefore comprise substitutions, insertions and/or deletions of the recited amino acids of the above sequence to accommodate the various strains, as long as the immuno-reactivity recognizable by the antibodies to HCV is preserved.

[0038] These peptides may also comprise conjugates, i.e., they may be coupled to carrier proteins such as bovine serum albumin (BSA) or human serum albumin (HSA). Furthermore, these peptides may comprise polymers, i.e., they may be synthesized on a polymeric resin or in dimeric, tetrameric, octameric and decahexyl forms of the peptide or their analogues, such as a branching octameric lysine resin.

[0039] The branchine poly-L-lysine can be Lys₈ Lys₄ Lys₂ Lys, Lys₄ Lys₂ Lys, Lys₂ Lys, Lys; the last Lys can be attached to Y as in Lys₄ Lys₂ Lys-Y wherein Y is -OH, -NH₂ or an amino acid containing no side chain functional group, such as alanine, valine, glycine, etc. Y can be inserted to facilitate synthesis onto the 4-methylbenzylhydramine resin. The conjugates and polymers of the peptides are also useful in the present invention.

[0040] The amino acid sequences of the polypeptide as described in the invention useful as test reagents for the detection of antibodies to HCV in body fluids and diagnosis of NANBH are selected to correspond to segments of the amino acid sequence of the postulated envelope and non-structural proteins of HCV designated as env, NS-1, NS-2, NS-3 and NS-5 based on amino acid sequence information derived from Houghton et al. (13), Okamoto et al (2) and

Kato et al (4).

[0041] In selecting regions of the HCV protein for epitope analysis, peptides of about 40 mer size with amino acid sequences covering the complete HCV envelope and non-structural proteins NS-1, NS-2, NS-3 and NS-5 were synthesized. These were tested for their immunoreactivity with special specimens previously selected through the screening of thousands of patient and normal sera for their unique immunoreactivity with HCV. Three peptides from the postulated envelope and nonstructural protein regions NS-1, NS-2, NS-3 and NS-5 designated as pep8, pep11 and pep12 and their analogues were identified to have specific immunoreactivity with the positive HCV sera.

[0042] At present, available knowledge of protein structure has not enabled the scientist to predict the amino acid sequences that may represent highly immunogenic epitopes. The usefulness of a peptide as an antigen or immunogen must be empirically determined. We have only been able to identify and characterize immuno-reactive epitopes through an extensive process which we call "serological validation". The following example shows how difficult it is to identify immuno-reactive epitopes.

[0043] For example, a clone designated as C33c encoded within the NS-3 region was reported to possess immunoreactivity(3). This clone spans 265 amino acid residues. Assuming a useful peptide must be at least 6 amino acids in length and that the upper limit for synthetic peptides in reasonable yield is 120 residues, the number of possible unique peptides from the C33c regions is 23,028. For the entire HCV genome, the figure is about 260,000.

[0044] In addition, we have shown that extraction conditions are critical for the expression of the immunopotency of a peptide (Example 4C), so the number of uniquely extracted peptides from this region is in multiples of 23,028. The possibilities for post-extraction modification, such as pH adjustment (Example 4B) further increase the possible selections to $>10^6$. If amino acid substitutions at various positions are taken into consideration, this figure will quickly increase to several millions. In contrast to the HCV core region, in which peptides VIII E and IX D were the optimal analogues, longer peptides are not preferred over shorter analogues in the NS-3/C33c region. For example, the 42 mer 279B shown on Table 4D has only 3% of the reactivity of the 37 mer peptide 3, designated as 279A in Table 4D. Of 30 peptides spanning the C33c region tested, only one was found to be useful. The antigenic index as referred in Houghton et al (3) did not prove to be a useful guide to epitopes, as the profile for peptide 3 is positive for only 30% of its sequence and negative for the remaining 70%.

[0045] The strategy for serological validation also depends on the expected characteristics of the target epitopes. Universal immunodominant epitopes, such as the gp41 transmembrane peptide of HIV-1, may be screened by a single representative serum sample from a patient known to be infected with the virus. Epitopes which are not recognized by all infected individuals, or those for which antibody is produced late or only transiently, and especially epitopes which give rise to neutralizing antibodies, must be screened by large panels of sera. For example, peptide 272B shown in Table 4A was initially tested on a panel of eight sera from HCV infected individuals (Panel 1). Only one sample was definitely positive with an absorbance of 880 mA. Three were weakly reactive (<200 mA) and four were negative.

[0046] The identification of the immuno-reactive epitopes is also dependent on the panel of sera used. The more closely the panel represents the population most likely to be seropositive for the desired epitope, the greater the chance that the epitope will be identified. For example, peptides synthesized from the NS-1 region, which were hypothesized to be important for generating neutralizing antibodies, gave only weakly reactive or negative results on screening with a very large number ($n > 200$) of samples from individuals who were newly infected and/or chronically infected with HCV. However, a panel of 24 samples from asymptomatic individuals from a known hepatitis virus endemic geographical region, Taiwan and mainland China, yielded two samples with absorbances of >2000 mA against multiple NS-1 peptides.

[0047] Finally, if the desired purpose of a targeted peptide/epitope is to extend the range of reactivity of an assay comprised of previously identified epitopes, then a large number of samples from individuals at risk of infection but seronegative against known epitopes must be employed for screening. Unfortunately, the most critical samples from clinically proven and documented cases of infection may be available in quantities insufficient for screening purposes. This is another complication/difficulty encountered in serological validation for determining the immunoreactivity of a peptide.

[0048] The process of "serological validation" is particularly difficult when the epitopes to be identified elicit antibodies only in a subpopulation of an infected patient group. When such epitopes become targets for identification, special attention must be paid to synthetic peptides which show very weak reactivity when tested by an enzyme immunoassay.

[0049] Fortunately, the low background absorbance of synthetic peptides allows for the precise detection of weak reactivities. In some cases, absorbances of 50 mA versus background reading are of sufficient significance and can lead to the identification of important epitopes through successive refinement of the amino acid sequence of a peptide. The utmost technical skill is required to obtain consistent and reliable results when working in the range of absorbances below 200-300 mA. For example: Peptides 261E and 261F shown on Table 4D were reactive with only one of eight HCV sera panel members (Panel I), with absorbances of 307 and 269 mA, respectively. Yet this weak reactivity led to the eventual identification of pep3 (or 279A), toward which half of the panel is reactive, and toward which some additional reactive samples show absorbances of >2000 mA.

[0050] Based on the immunoreactivities of the peptides according to the present invention, it is believed that these peptides may also be useful in a vaccine to prevent NANBH. The peptide when coupled to a protein, or synthesized on a polymeric carrier resin (e.g., an octameric lysine resin) or when polymerized to homo or hetero dimers or higher oligomers by cysteine oxidation, or induced disulfide cross linking, or by use of homo or hetero functional multivalent cross linking reagents, can be introduced to normal subjects to stimulate production of antibodies to HCV in healthy mammals.

[0051] The advantages of using synthetic peptides are known.

[0052] Since the peptides according to the present invention are not derived biologically from the virus, there is no danger of exposing the normal subjects who are to be vaccinated to the disease causing pathogen.

[0053] The peptides can be chemically synthesized easily. This means that there is no involvement with HCV at any time during the process of making the test reagent or the vaccine. Another problem which can be minimized by the process of the present invention is the false positive results caused by the presence of antigenic material co-purified with the HCV fusion protein. Certain normal individuals have antibodies to E. coli or yeast proteins which are cross reactive with the antigenic materials from the expression system. Sera from these normal individuals may show a positive reaction in the immunoassays.

[0054] Further, with appropriate amino acid modification or substitutions, it is expected that various peptide analogues based on the prescribed amino acid sequence can be synthesized with properties giving rise to lower background readings or better binding capacity to solid phases useful for HCV antibody screening assays.

[0055] Moreover, because the peptide compositions of the present invention are synthetically prepared, the quality can be controlled and as a result, reproducibility of the test results can be assured. Also, since very small amounts of a peptide are required for each test procedure, and because the expense of preparing a peptide is relatively low, the cost of screening body fluids for antibodies to HCV, diagnosis of NANBH infection, and the preparation of a vaccine is relatively low.

[0056] The peptides prepared in accordance with the present invention can be used to detect HCV infection and diagnose NANBH by using them as the test reagent in an enzyme-linked immunoadsorbent assay (ELISA), an enzyme immunodot assay, an agglutination based assay, or other well-known immunoassay devices. The following examples serve to illustrate the present invention and are not to be used to limit the scope of the invention.

EXAMPLE 1

Measurement of Relative (%) Immunoreactivity for HCV synthetic peptides by an Enzyme-Linked Immunosorbent Assay

[0057] As an example to illustrate how relative (%) immunoreactivity for HCV synthetic peptides is measured, wells of 96-well plates are coated for 1 hour at 37°C, with each of the following peptides: IIH, V, VIIIE and pep11 at 5 ug/mL at 100 uL per well in 10mM NaHCO₃ buffer, pH 9.5. The peptide coated wells were then incubated with 250 uL of 3% by weight of gelatin in PBS in 37°C for 1 hour to block non-specific protein binding sites, followed by three washes with PBS containing 0.05% by volume of TWEEN 20 and then dried. The test specimens containing a panel of eight well-characterized HCV antibody positive patient sera were diluted with PBS containing 20% by volume normal goat serum, 1% by weight gelatin and 0.05% by volume TWEEN 20 at dilutions of 1:20 volume to volume, respectively. 200 uL of the diluted specimens were added to each of the wells and allowed to react for 15 minutes at 37°C.

[0058] The wells were then washed six times with 0.05% by volume TWEEN 20 in PBS in order to remove unbound antibodies. Horseradish peroxidase conjugated goat anti-human IgG was used as a second antibody tracer to bind with the HCV antibody-peptide antigen complex formed in positive wells. 100 uL of peroxidase labeled goat anti-human IgG at a dilution of 1:1800 in 1% by volume normal goat serum, 0.05% by volume TWEEN 20 in PBS was added to each well and incubated at 37°C for another 15 minutes.

[0059] The wells were washed six times with 0.05% by volume TWEEN 20 PBS to remove unbound antibody and reacted with 100uL of the substrate mixture containing 0.04% by weight orthophenylenediamine (OPD) and 0.12% by volume hydrogen peroxide in sodium citrate buffer, pH 5.0.

[0060] This substrate mixture was used to detect the peroxidase label by forming a colored product. Reactions were stopped by the addition of 100 uL of 1.0M H₂SO₄ and the A₄₉₂mm measured. Results of relative immunoreactivity for each of the peptides obtained from this study are shown in Table A using peptide II H as the reference.

Table A

Peptide Code	A _{492nm} (Panel I, No. 1 to 8)								Total	%
	1	2	3	4	5	6	7	8		
IIH	0.812	0.656	3.114	2.737	1.066	2.254	2.599	3.478	16.712	100
V	0.834	1.060	2.931	0.534	0.137	0.434	0.303	2.787	9.020	54
VIII E	2.745	2.208	2.468	3.032	0.054	2.108	0.730	3.006	16.351	98
Pep11	0.241	0.715	3.162	1.020	0.568	2.166	3.330	3.477	14.690	88

EXAMPLE 2**Comparison of HCV Immunoreactivities by a Well-characterized 8 Member HCV Serum Panel (Panel I) for % Relative Immunoreactivity with a Group of HCV Capsid Protein Related Peptides by an Enzyme Immunoassay**

[0061] A 36mer HCV capsid peptide recently disclosed by Okamoto et al. (8) as the basis of an HCV EIA was synthesized for the purpose of comparison of immunoreactivity with peptides VIIIA, VIIIB and VIIIE (Table 2A). According to a procedure described in Example 1, peptides were coated at concentrations of 5, 1 and 0.2 µg/mL for immunopotency comparison. This 36mer exhibited only 47.8% of the reactivity of VIIIE (Table 2A). More importantly, when tested by our well-characterized HCV serum panel used for serological validation, only 4 out of 8 samples reacted with the 36mer, compared with 7 out of 8 with VIIIE. The C terminal end of this 36mer does not appear to contribute to the peptide's HCV immunoreactivity, since IXD is not greater in reactivity than IXC (Table 2A).

[0062] In addition, a 61mer peptide and fragments thereof consisting of a 30mer, a 40mer and a 50mer corresponding to sequences from Arima clone 1, which is homologous to the capsid region of the flavivirus yellow fever virus, were synthesized and compared in immunoreactivity with peptide VIIIE from the corresponding region of HCV (Table 2B). The 40mer and 61mer of clone 1 exhibited the most reactivity. However these were only 21.1% and 20.7%, respectively, of the immunoreactivity of peptide VIIIE.

	Table 2A	X Relative Immunoreactivity
Otsamoto et al.(8)(3dmer)	RKCPRLGVRAITSEISQPRGIRDP I PKVRIPPEGR	67.8%
VIII A	CPRLGVRAITSEISQPRGAR	32.7%
VIII B	VGCVTLPRGPCPLGVRAITSEISQPGCAR	48.9%
VIII E	STIPIEQPKTICNTINRPPDVKPFGCGRI VGCVTLPRGPCPLGVRAITSEISQPGCAR	100.0%
IX C	TVAOPGTVPVLPYGNEGCGIAQA L SPKCSRPVSQPTOPRRSRNLG	57.9%
IX D	I PKVRIPPEGR TVAOPGTVPVLPYGNEGCGIAQA L SPKCSRPVSQPTOPRRSRNLG	56.9%
IX E	GARQP I PKVRIPPEGR TVAOPGTVPVLPYGNEGCGIAQA L SPKCSRPVSQPTOPRRSRNLG	50.2%

Table 2b		% Relative Immunoreactivity
30 mer	PCGCGCGCPVGR CGAMRECGGDAVQ RGR	0.7%
40mer	KEKKTATIMPQCGCPVGR CGAMRECGGDAVQ RGR	21.1%
50mer	NDTNGDGRYTKKTAITMPQCGCPVGR CGAMRECGGDAVQ RGR	17.8%
61mer	KKCGASNGEAEQNTNGDGRYTKKTAITIMPQCGCPVGR CGAMRECGGDAVQ RGR	20.7%

EXAMPLE 3**Detection of Antibodies to HCV By an Agglutination Based Assay**

5 [0063] The presently claimed HCV peptides, synthesized according to the Merrifield solid phase method, can be conjugated to bovine serum albumin (BSA) by a simple crosslinking method in the presence of a low percentage of glutaraldehyde solution, or with other crosslinking reagent such as m-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS).

10 [0064] Based on the above mentioned peptide-BSA conjugation process, conjugated peptide was absorbed onto double aldehyde fixed human O erythrocytes at pH 4.0. The peptide-conjugate coated erythrocytes were then treated with NaBH₄ to prevent non-specific protein binding. The peptide-conjugate coated erythrocytes were then washed with PBS and incubated with 5% normal human serum-PBS solution. These processed cells were then used in an agglutination assay for the detection of HCV antibodies in both serum and plasma specimens. The specimens were diluted 1:10 in a sample diluent buffer and an equal volume of the indicator cells was mixed with the diluted specimens. The agglutination pattern was settled within one hour; and the assay results were read by eye. Serial bleedings from three well-characterized HCV seroconversion panels were tested for antibodies to HCV in the above-described HCV passive hemagglutination assay (PHA) employing Peptide VIII-E-BSA conjugate and Peptide IIH-BSA conjugate as the solid phase. The results were compared with the A492 a S/C of the peptide based HCV EIA (Format C, a combination of peptides IIH, V and VIII-E) and C100 based HCV EIA (Table 3).

20 [0065] In brief, the PHA assay detected HCV antibodies in all three panels as early as there was an increase in A492 in the peptide based EIA (Format C). rC100 based EIA lagged behind the HCV PHA results by 4-8 weeks.

Table 3

Detection of HCV Specific Antibodies from Seroconversion Panels by Various HCV Antibody Assays					
Series	Days	ALT	Format C HCV EIA S/C Ratio	C100 Based HCV EIA	HVC PHA Visual Score
A* (Serologicals Panel B)	0	4b	0.108	0.03	-
	7	32	0.045	0.04	-
	14	32	0.025	0.06	++
	21	180	1.037	0.04	++
	50	401	7.193	0.19	++
	92	-	10.185	6.57	++
	105	-	9.770	6.57	++
B* (Serologicals Panel A)	0	39	0	0	-
	10	274	0.058	0	-
	14	346	0.128	0	-
	30	1175	7.835	6.5	++
	51	430	7.811	6.5	++
C* (Serologicals Panel C)	0	63	0.115	0.04	-
	2	81	1.607	0.04	++
	9	183	2.506	0.02	++++
	29	563	9.827	6.57	++++
	57	436	10.630	6.57	++++

* Case presented is a plasma donor from a commercial source. Day 0 designates first sample in the series and does not correspond to date of exposure.

Example 4Detection of Antibodies to HCV by an Agglutination Assay Utilizing as the Solid Phase Immunosorbent Latex Particles Coated with HCV Peptide

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[0066] Using the peptide-BSA conjugation process mentioned in the previous example, conjugated peptide VIII-E-BSA, was absorbed to latex particles (0.4 μ size) at pH 9.5. The peptide-conjugate coated latex particles were then treated with BSA to prevent nonspecific protein binding. These coated latex particles were then used in an agglutination assay for the detection of HCV antibodies. The specimens were mixed in a ratio of 1:1 with the latex solution (0.5%).

10 The agglutination pattern was complete in a period of 15 min. Assay results were read by eye and by microscopic examination. The results of serial dilution samples from a well characterized anti-HCV positive plasma sample are summarized in Table 4. A coating concentration of 0.3 mg/mL was found to give optimal sensitivity for antibody detection. As a control for specificity, pooled plasma specimens from normal donors were tested in the peptide VIII-BSA conjugate latex assay and were found clearly negative.

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Table 4

Rapid Detection of HCV Antibodies using VIII-E-BSA Sensitized Latex Particles and Scoring for Visual Agglutination Pattern				
HCV Positive Control Dilution	Degree of Agglutination			
	VIII-E-BSA 2.4 mg/mL	Latex Particle 1.2 mg/mL	Coating 0.6 mg/mL	Concentration 0.3 mg/mL
1:1	4+	4+	4+	4+
1:2	4+	4+	4+	4+
1:5	4+	4+	4+	4+
1:10	4+	4+	4+	4+
1:20	3+	4+	4+	4+
1:40	2+	3+	4+	4+
1:80	+/-	-	+	3+
1:160	-	-	-	+
1:320	-	-	-	+/-
1:640	-	-	-	+/-
NP 1:1	-	-	-	-
NP: Pooled Normal Plasma				

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EXAMPLE 540 SYNTHESIS OF OCTAMERIC HCV PEPTIDE ANTIGENS AS KEY COMPONENTS OF IMMUNOGENS/VACCINES

[0067] The use of a limited sequential propagation of a trifunctional amino acid (or similar analogues) to form a core that serves as a low molecular weight matrix is the basic underlying principle for the formation of a radially branching multimeric peptide antigen system. The trifunctional amino acid, Boc-Lys(Boc), or di-(Boc)-Lys is most suitable since

45 both N α - and N ϵ - amino acid groups are available as reactive ends. Thus, sequential propagation of di-(Boc)-Lys will generate 2 n reactive ends. For example, the first level coupling of di-(Boc)-Lys will produce two reactive amino ends as a bivalent peptide antigen. Sequential generations of a second, third, and fourth step with di-(Boc)-Lys will therefore generate tetravalent, octavalent, and hexadecavalent peptide antigens respectively. As an example, an octameric HCV peptide immunogen with a structure of [Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys]₈-Lys₄-Lys₂-Lys

50 was synthesized as a prototype immunogen used in our immunization of guinea pigs. This octameric antigen contains a small heptalysyl core (<20%) and the bulk (>80%) is formed by a high density of uniform peptide-antigen layered around the core matrix. This design differs from the conventional peptide-carrier conjugate which contains a large protein carrier such as PPD or KLH and a low density of peptide antigens randomly distributed on the protein carrier surface in an unidentified form.

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[0068] For the synthesis of octameric HCV peptide immunogen, a combination of Boc-amino acid resin-bound benzhydrylamide and tBoc-chemistry was used. An octameric heptalysyl core resin was prepared by coupling di-t-Boc Lys onto an extra low loading 0.14 mmole/g MBHA (4-methyl benzhydrylamine) resin on a Biosearch 9500 instrument.

During each of the two coupling cycles, di-(Boc)-Lys was used for single coupling followed by two capping reactions (with 0.3 M acetylimidazole in DMF dimethylformamide).

[0069] After two additional di-(Boc)-Lys couplings onto the first di-(NH₂) Lys-resin, the substitution level of synthetic octameric resin was determined by ninhydrin test and found to have an appropriate level of -NH₂ groups, as calculated based on a theoretical coupling yield, and was used thereafter for the synthesis of octameric peptide antigens each with a predefined amino acid sequence according to the standard t-Boc chemistry.

[0070] Acid-labile tert-butyloxycarbonyl (t-Boc) was used for the protection of N- α amino acid. The following functional side-chain protecting groups were used: O-benzyl for Thr, Ser, Glu and Tyr; N⁶-tosyl for Arg; BOM (i.e. Boc-N^{im}-Benzoyloxymethyl-) for His; N²-dichlorobenzoyloxycarbonyl for Lys; S-4-methylbenzyl- for Cys; O-cyclohexyl for Asp and CHO for Trp. Successive amino acids were added as dictated by the sequence. The resultant octameric peptidyl resin was cleaved by anhydrous HF [0°C for 1 hr in the presence of 10% (v/v) anisole]. The released octomeric antigen was extracted by acetic acid, after two cycles of ether washings of the cleaved peptidyl resin, and lyophilized to dryness so as to be ready for use as an immunogen. A computer-generated picture of such an octameric immunogen is shown in Fig. 1.

EXAMPLE 6

Detection of Antibodies to HCV by Peptide Based HCV EIA Using Formats 1 to 6

[0071] The following five groups of serum specimens:

- (a) Plasmapheresis donors with elevated (>100 i.u./L) alanine aminotransferase (ALT) enzyme activity (n=30);
- (b) Blood donors with elevated (>45 i.u./L) ALT enzyme activity (n=15);
- (c) Chronic NANBH patients (n=30);
- (d) Other viral infections (n=11);
- (e) Autoimmune disease patients (n=9);

were analyzed on representative HCV peptide based EIA kits according to the present invention, with the plates coated at 100 μ L per well either with:

- (i) Format 1: peptides VIII E, II H and pepII at 0.5, 3 and 1 μ g/mL each;
- (ii) Format 2: peptides VIII E and pepII at 0.5 and 1 μ g/mL each;
- (iii) Format 3: peptides VIII E, pepII and peps at 0.5, 1 and 10 μ g/mL each;
- (iv) Format 4: peptides VIII E and pep8 at 0.5 and 10 μ g/mL each;
- (v) Format 5: peptides VIII E, pepII and pep12 at 0.5, 1 and 2 μ g/mL each;
- (vi) or Format 6: peptides VIII E and pep12 at 0.5 and 2 μ g/mL each.

[0072] These kits represent core, NS-4 and NS-5 (Format 1), core and NS-5 (Formats 2, 5 and 6), core, NS-5 and env (Format 3) and core and env (Format 4).

[0073] The results of testing these 95 well characterized samples on Formats 1 through 6 are presented in Table 5. The results indicate that (30/30) of the samples in group (a) were reactive by Formats 1, 2 and 3; 90% (27/30) reactive by Format 4 and 97% (29/30) reactive by Formats 5 and 6. All samples in groups (b) and (c) were positive on all 6 formats. Groups (a), (b) and (c) were shown to be reactive by Format C, a mixture of peptides II H, V and VIII E.

[0074] Three samples in group (d) were reactive by Formats 1 to 4. In contrast, these samples were indicated as negative by Format C. Serum samples "86" and "124" apparently responded to the presence of pepII, and serum sample "VZV2500" was indicated as positive by the presence of pep8 in Formats 4 and 5.

[0075] All serum samples in group (c) were negative on all formats, including Format C.

Table 5

Antibody to HCV Detected By Peptide Based EIA Kits (Absorbance 492nm)						
Sample ID	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6
NRC	0.065	0.075	0.056	0.061	0.060	0.019
WRC	0.650	0.454	0.953	0.967	0.403	0.340
SRC	2.183	1.791	2.580	2.635	1.589	1.331

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Table 5 (continued)

Antibody to HCV Detected By Peptide Based EIA Kits (Absorbance 492nm)							
Sample ID		Format 1	Format 2	Format 3	Format 4	Format 5	Format 6
a. Plasmapheresis, ALT > 100 i.u.L							
1	-13	3.166	3.419	3.255	3.371	3.291	3.255
	-27	1.555	1.548	1.980	2.904	1.152	0.881
	-31	3.479	3.144	3.220	2.332	3.319	2.665
	-32	3.001	3.035	3.112	2.691	3.076	2.986
	-39	3.063	3.041	3.361	2.886	3.190	3.038
	-42	3.198	3.201	3.050	3.227	3.230	3.118
	-47	3.479	3.110	3.251	3.201	3.229	3.068
	-48	3.142	2.795	3.116	2.934	3.076	2.725
	-49	3.417	3.291	3.525	3.451	3.195	3.592
	-52	3.263	3.329	3.202	0.120	3.262	3.453
	-53	3.225	3.145	3.096	0.062	3.358	3.097
	-54	3.271	3.018	3.267	0.153	3.073	3.211
2	-4	1.012	0.881	1.542	1.767	0.807	0.745
	-6	3.229	2.964	3.169	3.052	3.076	2.897
	-9	2.691	2.416	2.766	2.967	2.119	1.844
	-26	3.222	3.055	3.095	3.167	3.195	2.951
	-32	3.226	3.372	3.368	3.194	3.496	3.417
	-33	3.151	2.918	3.147	3.027	3.108	3.129
	-34	3.059	3.021	3.143	3.167	3.145	3.320
	-38	3.241	3.116	2.967	3.055	3.213	3.137
	-41	2.964	2.593	2.841	2.964	2.469	2.252
	-43	3.146	2.092	2.541	2.627	1.999	1.920
	-46	2.927	2.818	2.998	2.983	2.556	2.415
	-58	3.285	3.444	3.218	3.191	3.355	3.095
	-60	3.094	2.975	3.113	3.167	2.683	2.640
	-61	2.784	2.345	2.501	2.751	2.007	2.212
	-62	3.320	3.076	3.095	3.076	3.003	2.787
	-77	0.815	0.682	1.096	0.418	0.164	0.152
	-82	3.020	2.982	1.826	3.001	3.032	2.820
	-83	3.076	2.914	3.049	2.996	2.928	2.808
b. Elevated ALT blood donors (ALT > i.u./L)							
ALT	-1	3.017	3.035	3.116	3.165	3.167	2.920
	-2	3.256	3.166	3.165	2.974	3.292	3.091
	-3	3.153	3.328	3.291	3.105	3.203	3.230
	-4	2.969	2.894	3.096	3.144	2.880	2.866
	-5	3.073	2.956	2.968	2.952	3.376	2.985
	-7	3.218	3.020	3.157	2.980	2.951	3.060
	-8	3.074	2.930	3.094	3.197	3.121	3.012
	-10	3.479	3.228	3.226	3.109	3.432	3.952
	-11	3.398	3.283	3.140	3.035	3.285	3.222
	-53	3.330	3.029	3.253	3.290	2.974	3.070
	-56	3.151	3.086	3.176	3.202	3.107	3.085
	-69	3.021	3.170	3.167	3.318	3.019	2.831
	-70	3.074	3.035	2.951	3.073	3.054	3.184
	-71	2.985	2.901	3.080	3.039	2.902	2.900
	-82	3.230	3.120	3.085	2.977	3.298	3.096

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Table 5 (continued)

Antibody to HCV Detected By Peptide Based EIA Kits (Absorbance 492nm)							
Sample ID	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6	
c. Chronic NANBH							
N	-2	3.320	3.052	2.981	3.283	3.032	2.999
	-3	3.285	3.036	3.095	3.167	3.077	3.094
	-4	3.117	3.469	3.590	3.291	2.259	3.141
	-7	3.027	3.008	3.061	3.065	2.962	2.806
	-8	3.285	3.146	3.117	3.194	3.122	3.195
	-9	2.886	3.001	3.072	2.985	2.848	2.859
	-10	2.606	2.268	2.027	0.423	2.338	1.104
	-14	3.054	2.808	2.856	2.995	2.341	2.041
	-23	3.228	3.050	3.067	3.225	3.152	3.109
	-25	3.891	2.462	3.190	3.165	1.982	2.091
	-27	3.194	2.926	3.165	3.029	3.143	3.168
	-28	3.027	3.106	3.259	3.175	3.176	3.202
	-34	3.057	3.037	3.035	3.144	2.907	2.892
	-36	3.304	3.213	3.000	3.033	3.075	3.115
	-41	3.217	3.283	3.039	3.248	3.290	3.249
	-42	2.997	2.858	3.196	3.094	3.097	2.805
	-44	3.391	3.477	3.350	3.254	3.353	3.387
	-45	3.318	3.096	2.964	3.250	3.319	3.036
	-49	3.292	3.371	3.416	3.255	3.292	3.370
	-54	3.329	3.294	3.105	3.105	3.177	3.203
30	-57	3.197	3.169	3.221	3.141	3.120	3.018
	-60	3.115	3.035	3.090	3.072	3.096	2.873
	-65	2.020	1.816	1.898	2.376	1.133	1.284
	-67	2.265	1.776	2.356	2.396	1.319	0.911
	-68	3.178	3.177	3.200	3.176	3.530	3.087
	-69	3.222	3.167	3.165	3.283	3.399	3.097
	-77	1.438	1.346	2.548	2.397	1.055	1.071
	-78	2.457	2.038	2.251	2.300	1.642	1.494
35	-79	3.225	3.197	3.076	3.142	3.224	3.169
	-80	3.138	3.074	3.135	3.054	3.137	2.896
d. Other Viral Infections							
HAV	-86	0.558	0.316	0.607	0.054	0.037	0.014
	-88	0.018	0.021	0.062	0.054	0.014	0.018
	-92	0.045	0.061	0.058	0.050	0.043	0.007
	-120	0.057	0.076	0.051	0.032	0.051	0.026
	-121	0.052	0.138	0.094	0.065	0.072	0.026
	-124	0.816	1.178	0.622	0.062	1.082	0.017
	-125	0.014	0.016	0.050	0.031	0.012	0.010
	-126	0.105	0.134	0.109	0.081	0.117	0.068
EBV	-2331	0.021	0.021	0.023	0.020	0.012	0.012
VZV	-M002	0.035	0.030	0.154	0.108	0.025	0.012
VZV	-2500	0.090	0.138	0.976	0.923	0.084	0.032
e. Autoimmune							
55	-209	0.102	0.079	0.117	0.097	0.066	0.028
	-210	0.002	0.003	0.018	0.011	0.002	0.005

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Table 5 (continued)

Antibody to HCV Detected By Peptide Based EIA Kits (Absorbance 492nm)							
Sample ID	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6	
e. Autoimmune							
-211	0.016	0.019	0.134	0.168	0.022	0.016	
-212	0.016	0.020	0.075	0.080	0.019	0.006	
-213	0.008	0.009	0.055	0.076	0.005	0.002	
-215	0.118	0.095	0.226	0.282	0.093	0.060	
-216	0.039	0.037	0.100	0.105	0.042	0.022	
-217	0.019	0.021	0.068	0.056	0.023	0.012	
-218	0.032	0.022	0.110	0.086	0.059	0.031	

EXAMPLE 7

Comparison of Test Results Using the Six Peptide Based HCV EIA Formats (1-6) on Random Blood Donors

[0076] Random blood donor samples (n=100) were tested by Formats 1 to 6. All 100 samples were negative on Formats 2, 5 and 6. Sample 14 had an absorbance of 0.680 on Format 1, and sample 34 had an absorbance of 0.601 and 0.551 on Formats 3 and 4, respectively. For the calculation of mean absorbance and standard deviation, absorbance values >0.500 were omitted from analysis. Table 6 lists the mean absorbance and standard deviation of the 100 samples on Formats 1-6.

Table 6

Mean Absorbance (A492nm) \pm SD of 100 Random Blood Donors						
	Format 1	Format 2	Format 3	Format 4	Format 5	Format 6
Mean	0.040	0.035	0.068	0.061	0.030	0.017
S.D.	0.036	0.029	0.046	0.046	0.039	0.032

EXAMPLE 8

Comparison of Immunoreactivity for NS-5 Protein Derived Synthetic Peptides

[0077] Wells of 96-well plates were coated for 1 hour at 37°C with each of the 23 peptides (designated as 259A-259E, 260A-260C, 309A-309C, 310A-310C, 311A-311C, 312A-312C and 314A-314C) synthesized with sequences derived from the NS-5 region, at 5 μ g/mL at 100 μ L per well in 10 mM NaHCO₃ buffer, pH 9.5. The immunoreactivity of each peptide was measured by an 8 member HCV serum panel (Panel I). The peptide with the greatest immunoreactivity was pep11, designated 309C in Table 7. When the immunoreactivity of pep11 was used as a standard to calculate the relative immunopotency for the other NS-5 peptides, the peptides in series 309-314 were seen to be equal to or more reactive than pep4 and pep5. The extension of pep5 to include an additional 10 residues (259E, i.e. pep12) increased the relative immunopotency from 47.6% to 70.1%.

[0078]

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Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
5 10 15
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
20 25 30
Val-Pro-Ala-Lys-Ser-Val-Cys
35

15 SEQ ID No.: 241B
Sequence Type: AA
Sequence Length: 45 AA

20 Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-Gln-Gly-Trp-Gly-Pro-Ile-Ser-
5 10 15
Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-
20 25 30
Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-
35 40 45

30 SEQ ID No.: 241C
Sequence Type: AA
Sequence Length: 52 AA

35 Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-
5 10 15
Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
20 25 30
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
35 40 45
40 Val-Pro-Ala-Lys-Ser-Val-Cys
50

SEQ ID No.: 231A.
Sequence Type: AA
Sequence Length: 26 AA

Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-
5 10 15
Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
20 25

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SEQ ID No.: 231B
Sequence Type: AA
Sequence Length: 34 AA

5 Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-
5 10 15
Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-
20 25 30
10 Pro-Val-Tyr-Cys

15 SEQ ID No.: 231C (Pep 1)
Sequence Type: AA
Sequence Length: 42 AA

20 Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
5 10 15
Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
20 25 30
25 Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
35 40

30 SEQ ID No.: 231D
Sequence Type: AA
Sequence Length: 50 AA

35 Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-Gln-Gly-Trp-Gly-Pro-Ile-Ser-
5 10 15
Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-Gln-Arg-Pro-Tyr-Cys-Trp-His-
20 25 30
40 Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-Val-Pro-Ala-Lys-Ser-Val-Cys-
35 40 45
Gly-Pro-Val-Tyr-Cys
45 50

50 SEQ ID No.: 231E
Sequence Type: AA
Sequence Length: 57 AA

55

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Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys-Arg-Pro-Leu-Thr-Asp-Phe-Asp-
 5 10 15
 5 Gln-Gly-Trp-Gly-Pro-Ile-Ser-Tyr-Ala-Asn-Gly-Ser-Gly-Pro-Asp-
 20 25 30
 Gln-Arg-Pro-Tyr-Cys-Trp-His-Tyr-Pro-Pro-Lys-Pro-Cys-Gly-Ile-
 35 40 45
 10 Val-Pro-Ala-Lys-Ser-Val-Cys-Gly-Pro-Val-Tyr-Cys
 50 55

15 SEQ ID No.: 232A (Pep 2)
 Sequence Type: AA
 Sequence Length: 26 AA

20 Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-
 5 10 15
 Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys
 20 25

30 SEQ ID No.: 232B
 Sequence Type: AA
 Sequence Length: 34 AA

35 Val-Phe-Val-Leu-Asn-Asn-Thr-Arg-Pro-Pro-Leu-Gly-Asn-Trp-Phe-
 5 10 15
 Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-Gly-Phe-Thr-Lys-Val-Cys-Gly-
 20 25 30
 40 Ala-Pro-Pro-Cys

45 SEQ ID No.: 232C
 Sequence Type: AA
 Sequence Length: 42 AA

50 Ser-Trp-Gly-Glu-Asn-Asp-Thr-Asp-Val-Phe-Val-Leu-Asn-Asn-Thr-
 5 10 15
 Arg-Pro-Pro-Leu-Gly-Asn-Trp-Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-
 20 25 30
 Thr-Gly-Phe-Thr-Lys-Val-Cys-Gly-Ala-Pro-Pro-Cys
 35 40

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SEQ ID No.: 232D
Sequence Type: AA
Sequence Length: 50 AA

5 Asp-Arg-Ser-Gly-Ala-Pro-Thr-Tyr-Ser-Trp-Gly-Glu-Asn-Asp-Thr-
5 10 15
Asp-Val-Phe-Val-Leu-Asn-Asn-Thr-Arg-Pro-Pro-Leu-Gly-Asn-Trp-
20 25 30
10 Phe-Gly-Cys-Thr-Trp-Met-Asn-Ser-Thr-Gly-Phe-Thr-Lys-Val-Cys-
35 40 45
Gly-Ala-Pro-Pro-Cys
50

15

SEQ ID No.: 233C
Sequence Type: AA
Sequence Length: 42 AA

20

25 Leu-His-Cys-Pro-Thr-Asp-Cys-Phe-Arg-Lys-His-Pro-Asp-Ala-Thr-
5 50 15
Tyr-Ser-Arg-Cys-Gly-Ser-Gly-Pro-Trp-Ile-Thr-Pro-Arg-Cys-Leu-
20 25 30
30 Val-Asp-Tyr-Pro-Tyr-Arg-Leu-Trp-His-Trp-Pro-Cys
35 40

SEQ ID No.: 234A
Sequence Type: AA
Sequence Length: 23 AA

35

40 Glu-Ala-Ala-Cys-Asn-Trp-Thr-Arg-Gly-Glu-Arg-Cys-Asp-Leu-Glu-
5 10 15
Asp-Arg-Asp-Arg-Ser-Glu-Leu-Ser
20

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SEQ ID No.: 234B
Sequence Type: AA
Sequence Length: 31 AA

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Val-Gly-Gly-Val-Glu-His-Arg-Leu-Glu-Ala-Ala-Cys-Asn-Trp-Thr-
5 Arg-Gly-Glu-Arg-Cys-Asp-Leu-Glu-Asp-Arg-Asp-Arg-Ser-Glu-Leu-
Ser

10

SEQ ID No.: 234C
Sequence Type: AA
Sequence Length: 39 AA

Thr-Ile-Phe-Lys-Ile-Arg-Met-Tyr-Val-Gly-Gly-Val-Glu-His-Arg-
20 Leu-Glu-Ala-Ala-Cys-Asn-Trp-Thr-Arg-Gly-Glu-Arg-Cys-Asp-Leu-
Glu-Asp-Arg-Asp-Arg-Ser-Glu-Leu-Ser

25

SEQ ID No.: 272A
Sequence Type: AA
Sequence Length: 41 AA

Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-
35 Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-
Gln-Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser

40

SEQ ID No.: 272B
Sequence Type: AA
Sequence Length: 55 AA

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Thr-Thr-Met-Arg-Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-
 5 10 15
 5 Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-
 20 25 30
 Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-
 35 40 45
 10 Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser
 50 55

15 SEQ ID No.: 272C
 Sequence Type: AA
 Sequence Length: 66 AA

20 Ala-Val-Asp-Phe-Ile-Pro-Val-Glu-Asn-Leu-Glu-Thr-Thr-Met-Arg-
 5 10 15
 Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-Val-Pro-Gln-
 20 25 30
 25 Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-
 35 40 45
 Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-Gly-Tyr-Lys-Val-
 50 55 60
 30 Leu-Val-Leu-Asn-Pro-Ser
 65

35 SEQ ID No.: 278A
 Sequence Type: AA
 Sequence Length: 29 AA

40 Pro-Val-Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-
 5 10 15
 Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala
 20 25

50 SEQ ID No.: 278B
 Sequence Type: AA
 Sequence Length: 36 AA

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Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
5 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
Ile-Ile-Cys-Asp-Glu-Cys-His-Ser

10

SEQ ID No.: 275B
Sequence Type: AA
Sequence Length: 71 AA

15

Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-
20 Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-
Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-
25 Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-
Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser

30

SEQ ID No.: 275C
Sequence Type: AA
Sequence Length: 94 AA

35

His-Leu-His-Ala-Pro-Thr-Gly-Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-
40 Ala-Ala-Tyr-Ala-Ala-Gln-Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-
Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-
45 His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-
Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-
50 Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-
Glu-Cys-His-Ser

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SEQ ID No.: 275D
Sequence Type: AA
Sequence Length: 117 AA

5 Thr-Met-Arg-Ser-Pro-Val-Phe-Thr-Asp-Asn-Ser-Ser-Pro-Pro-Val-
5 10 15
Val-Pro-Gln-Ser-Phe-Gln-Val-Ala-His-Leu-His-Ala-Pro-Thr-Gly-
10 20 25 30
Ser-Gly-Lys-Ser-Thr-Lys-Val-Pro-Ala-Ala-Tyr-Ala-Ala-Gln-Gly-
35 40 45
Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-Gly-
15 50 55 60
Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-
65 70 75
Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-
20 80 85 90
Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-
95 100 105
Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser
110 115

SEQ ID No.: 274A
Sequence Type: AA
Sequence Length: 37 AA

35 Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
5 10 15
Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
20 25 30
Asn-Ile-Glu-Glu-Val-Ala-Leu
35

SEQ ID No.: 274B
Sequence Type: AA
Sequence Length: 64AA

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Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-
5 5 10 15
His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
 20 25 30
Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-Leu-Ala-Thr-
 35 40 45
Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-
10 50 55 60
Glu-Val-Ala-Leu

15

SEQ ID No.: 274C
Sequence Type: AA
Sequence Length: 97 AA

20

Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-
5 10 15
25 Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-
20 25 30
Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-
35 40 45
30 Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-
50 55 60
Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
65 70 75
35 Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
80 85 90
Asn-Ile-Glu-Glu-Val-Ala-Leu
95

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SEQ ID No.: 274D
Sequence Type: AA
Sequence Length: 120 AA

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25 SEQ ID No.: 262A
Sequence Type: AA
Sequence Length: 29 AA

40 SEQ ID No.: 262B
Sequence Type: AA
Sequence Length: 39 AA

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SEQ ID No.: 262C
Sequence Type: AA
Sequence Length: 49 AA

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Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-
5 Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-
Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-
10 Thr-Asp-Ala-Thr

15 SEQ ID No.: 262D
Sequence Type: AA
Sequence Length: 59 AA

20 Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-
Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-
25 Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-
Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr

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35 SEQ ID No.: 262E
Sequence Type: AA
Sequence Length: 68 AA

40 Ser-Val-Ala-Ala-Thr-Leu-Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-
His-Gly-Ile-Asp-Pro-Asn-Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-
45 Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-
Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-
50 Glu-Cys-His-Ser-Thr-Asp-Ala-Thr

55 SEQ ID No.: 262F
Sequence Type: AA
Sequence Length: 77 AA

Gly-Tyr-Lys-Val-Leu-Val-Leu-Asn-Pro-Ser-Val-Ala-Ala-Thr-Leu-
5 5 10 15
Gly-Phe-Gly-Ala-Tyr-Met-Ser-Lys-Ala-His-Gly-Ile-Asp-Pro-Asn-
 20 25 30
Ile-Arg-Thr-Gly-Val-Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-
 35 40 45
Tyr-Ser-Thr-Tyr-Gly-Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-
10 50 55 60
Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-
 65 70 75
Ala-Thr

15

20 SEQ ID No.: 261A
Sequence Type: AA
Sequence Length: 30 AA

[illegible]

SEQ ID No.: 261B
Sequence Type: AA
Sequence Length: 40 AA

Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-
5 10 15
Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-
40 20 25 30
Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-Leu
35 40

50 SEQ ID No.: 261C
Sequence Type: AA
Sequence Length: 50 AA

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Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-
 5 10 15
 5 Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-
 20 25 30
 Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-
 35 40 45
 10 Lys-Cys-Asp-Glu-Leu
 50

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SEQ ID No.: 261D
 Sequence Type: AA
 Sequence Length: 73 AA

20

Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-
 5 10 15
 25 Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-
 20 25 30
 Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-
 35 40 45
 30 Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-
 50 55 60
 Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-Leu
 65 70

35

SEQ ID No.: 261E
 Sequence Type: AA
 Sequence Length: 97 AA

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Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-
 5 10 15
 5 Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-
 20 25 30
 Glu-Thr-Ala-Gly-Ala-Arg-Leu-Val-Val-Leu-Ala-Thr-Ala-Thr-Pro-
 35 40 45
 10 Pro-Gly-Ser-Val-Thr-Val-Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-
 50 55 60
 Leu-Ser-Thr-Thr-Gly-Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-
 65 70 75
 15 Leu-Glu-Val-Ile-Lys-Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-
 80 85 90
 Lys-Lys-Lys-Cys-Asp-Glu-Leu
 95

20

25 SEQ ID No.: 261F
 Sequence Type: AA
 Sequence Length: 121 AA

30 Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
 5 10 15
 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
 20 25 30
 35 Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-
 35 40 45
 Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly-Ala-Arg-
 50 55 60
 40 Leu-Val-Val-Leu-Ala-Thr-Ala-Thr-Pro-Pro-Gly-Ser-Val-Thr-Val-
 65 70 75
 Pro-His-Pro-Asn-Ile-Glu-Glu-Val-Ala-Leu-Ser-Thr-Thr-Gly-Glu-
 80 85 90
 45 Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-Gly-
 95 100 105
 Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-Glu-
 110 115 120
 50 Leu

55 SEQ ID No.: 279A (Pep 3)
 Sequence Type: AA
 Sequence Length: 37 AA

EP 0 468 527 B1

5 Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-Ile-Cys-Asp-Glu-Cys-
 10 5 10 15
 His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-Ile-Gly-Thr-Val-Leu-
 20 25 30
 Asp-Gln-Ala-Glu-Thr-Ala-Gly
 35

10

15

SEQ ID No.: 279B
 Sequence Type: AA
 Sequence Length: 42 AA

20

Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-Ile-
 5 10 15
 Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-Gly-
 20 25 30
 Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly
 35 40

30

SEQ ID No.: 279E
 Sequence Type: AA
 Sequence Length: 58 AA

35

Arg-Thr-Ile-Thr-Thr-Gly-Ser-Pro-Ile-Thr-Tyr-Ser-Thr-Tyr-Gly-
 5 10 15
 Lys-Phe-Leu-Ala-Asp-Gly-Gly-Cys-Ser-Gly-Gly-Ala-Tyr-Asp-Ile-
 20 25 30
 Ile-Ile-Cys-Asp-Glu-Cys-His-Ser-Thr-Asp-Ala-Thr-Ser-Ile-Leu-
 35 40 45
 Gly-Ile-Gly-Thr-Val-Leu-Asp-Gln-Ala-Glu-Thr-Ala-Gly
 50 55

50

SEQ ID No.: 255A
 Sequence Type: AA
 Sequence Length: 35 AA

55

EP 0 468 527 B1

Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-
5 Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-
Asn-Val-Ser-Arg-Cys
35

10

SEQ ID No.: 255B
Sequence Type: AA
Sequence Length: 45 AA

15

Val-Arg-Asn-Ser-Thr-Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-
20 Asn-Ser-Ser-Ile-Val-Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-
Pro-Gly-Cys-Val-Pro-Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-
35 40 45

25

SEQ ID No.: 255C (Pep 7)
Sequence Type: AA
Sequence Length: 55 AA

30

Cys-Leu-Thr-Val-Pro-Ala-Ser-Ala-Tyr-Gln-Val-Arg-Asn-Ser-Thr-
35 Gly-Leu-Tyr-His-Val-Thr-Asn-Asp-Cys-Pro-Asn-Ser-Ser-Ile-Val-
Tyr-Glu-Ala-His-Asp-Ala-Ile-Leu-His-Thr-Pro-Gly-Cys-Val-Pro-
40 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys
50 55

45

SEQ ID No.: 244A
Sequence Type: AA
Sequence Length: 35 AA

50

55

EP 0 468 527 B1

5 Cys-Trp-Val-Ala-Met-Thr-Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-
 10 Leu-Pro-Ala-Thr-Gln-Leu-Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-
 15 Ser-Ala-Thr-Leu-Cys

10

15 SEQ ID No.: 244B
 Sequence Type: AA
 Sequence Length: 44 AA

20 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
 25 Pro-Thr-Val-Ala-Thr-Arg-Asp-Gly-Lys-Leu-Pro-Ala-Thr-Gln-Leu-
 30 Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys

30 SEQ ID No.: 254A
 Sequence Type: AA
 Sequence Length: 30 AA

35 Thr-Gln-Gly-Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-
 40 His-Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala

40

45 SEQ ID No.: 254B (Pep 8)
 Sequence Type: AA
 Sequence Length: 40 AA

50 Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-
 55 Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-
 Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-Ala

EP 0 468 527 B1

SEQ ID No.: 254C
Sequence Type: AA
Sequence Length: 50 AA

5 Cys-Gly-Ser-Val-Phe-Leu-Ile-Gly-Gln-Leu-Phe-Thr-Phe-Ser-Pro-
5 10 15
Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-Cys-Asn-Cys-Ser-Ile-Tyr-Pro-
20 25 30
10 Gly-His-Ile-Thr-Gly-His-Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-
35 40 45
Trp-Ser-Pro-Thr-Ala
50
15

SEQ ID No.: 248A
Sequence Type: AA
Sequence Length: 25 AA

20
25 Asp-Met-Ile-Ala-Gly-Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-
5 10 15
Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys
20 25
30

SEQ ID No.: 248B
Sequence Type: AA
Sequence Length: 35 AA

35
40 Gln-Leu-Leu-Arg-Ile-Pro-Gln-Ala-Ile-Leu-Asp-Met-Ile-Ala-Gly-
5 10 15
Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-Tyr-Phe-Ser-Met-Val-
20 25 30
Gly-Asn-Trp-Ala-Lys
35
45

SEQ ID No.: 248C
Sequence Type: AA
Sequence Length: 40 AA

50
55

EP 0 468 527 B1

Ala-Leu-Val-Met-Ala-Gln-Leu-Leu-Arg-Ile-Pro-Gln-Ala-Ile-Leu-
 5 10 15

Asp-Met-IleAla-Gly-Ala-His-Trp-Gly-Val-Leu-Ala-Gly-Ile-Ala-
 20 25 30

Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys
 35 40

SEQ ID No.: 247A
Sequence Type: AA
Sequence Length: 25 AA

Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-
5 10 15
20 Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
20 25

SEQ ID No.: 247B (Pap 9)
Sequence Type: AA
Sequence Length: 35 AA

Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-
5 10 15
Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-
20 25 30
35 Ile-Gln-Leu-Ile-Asn
35

45 SEQ ID No.: 247C
Sequence Type: AA
Sequence Length: 45AA

50 Val-Leu-Val-Val-Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-
5 10 15
Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-
20 25 30
55 Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
35 40 45

EP 0 468 527 B1

SEQ ID No.: 247D
Sequence Type: AA
Sequence Length: 55 AA

5 Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys-Val-Leu-Val-Val-Leu-
5 10 15
Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-Ile-Val-Ser-Gly-Gly-
20 25 30
10 Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-Ser-Leu-Phe-Thr-Pro-
35 40 45
Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
50 55
15

SEQ ID No.: 247E
Sequence Type: AA
Sequence Length: 60 AA

20
25 Leu-Ala-Gly-Ile-Ala-Tyr-Phe-Ser-Met-Val-Gly-Asn-Trp-Ala-Lys-
5 10 15
Val-Leu-Val-Val-Leu-Leu-Leu-Phe-Ala-Gly-Val-Asp-Ala-Glu-Thr-
20 25 30
30 Ile-Val-Ser-Gly-Gly-Gln-Ala-Ala-Arg-Ala-Met-Ser-Gly-Leu-Val-
35 40 45
Ser-Leu-Phe-Thr-Pro-Gly-Ala-Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn
50 55 60
35

SEQ ID No.: 246A
Sequence Type: AA
Sequence Length: 25 AA

40
Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-
5 10 15
45 Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys
20 25

SEQ ID No.: 246B
Sequence Type: AA
Sequence Length: 31 AA

55

EP 0 468 527 B1

5 Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-
5 Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-
Cys

10 SEQ ID No.: 246C
Sequence Type: AA
Sequence Length: 38 AA

15 Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-
20 Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-
Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys

25 SEQ ID No.: 246D (Pep 10)
Sequence Type: AA
Sequence Length: 40 AA

30 Trp-His-Ile-Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-
35 Thr-Gly-Trp-Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-
Ser-Gly-Cys-Pro-Glu-Arg-Leu-Ala-Ser-Cys

40 SEQ ID No.: 246E
Sequence Type: AA
Sequence Length: 52 AA

50

55

EP 0 468 527 B1

5 Lys-Gln-Asn-Ile-Gln-Leu-Ile-Asn-Thr-Asn-Gly-Ser-Trp-His-Ile-
 5 10 15
 Asn-Ser-Thr-Ala-Leu-Asn-Cys-Asn-Glu-Ser-Leu-Asn-Thr-Gly-Trp-
 20 25 30
 Leu-Ala-Gly-Leu-Ile-Tyr-Gln-His-Lys-Phe-Asn-Ser-Ser-Gly-Cys-
 35 40 45
 10 Pro-Glu-Arg-Leu-Ala-Ser-Cys
 50

15
 SEQ ID No.: 314A (Pep 16)
 Sequence Length: 30 AA

20 Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-
 5 10 15
 Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu
 20 25 30
 25

30
 SEQ ID No.: 314B
 Sequence Type: AA
 Sequence Length: 38 AA

35 Ser-Glu-Cys-Thr-Thr-Pro-Cys-Ser-Gly-Ser-Trp-Leu-Arg-Asp-Ile-
 5 10 15
 Trp-Asp-Trp-Ile-Cys-Glu-Val-Leu-Ser-Asp-Phe-Lys-Thr-Trp-Leu-
 20 25 30
 40 Lys-Ala-Lys-Leu-Met-Pro-Gln-Leu
 35

45
 SEQ ID No.: 314C
 Sequence Type: AA
 Sequence Length: 47 AA

50

55

```

Leu-Arg-Arg-Leu-His-Gln-Trp-Ile-Ser-Ser-Glu-Cys-Thr-Thr-Pro-
                    5                                10                                15
5 Cys-Ser-Gly-Ser-Trp-Leu-Arg-Asp-Ile-Trp-Asp-Trp-Ile-Cys-Glu-
                    20                                25                                30
Val-Leu-Ser-Asp-Phe-Lys-Thr-Trp-Leu-Lys-Ala-Lys-Leu-Met-Pro-
                    35                                40                                45
10 Gln-Leu

```

15 SEQ ID No.: 312A
Sequence Type: AA
Sequence Length: 22 AA

20 Asp-Phe-His-Tyr-Val-Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-Cys-
5 10 15
Pro-Cys-Gln-Val-Pro-Ser-Pro
20

30 SEQ ID No.: 312B (Pep 15)
Sequence Type: AA
Sequence Length: 32 AA

[illegible]

45 SEQ ID No.: 312C
Sequence Type: AA
Sequence Length: 38 AA

Leu-Trp-Arg-Val-Ser-Ala-Glu-Glu-Tyr-Val-Glu-Ile-Arg-Gln-Val-
 5 10 15
⁵⁰ Gly-Asp-Phe-His-Tyr-Val-Thr-Gly-Met-Thr-Thr-Asp-Asn-Leu-Lys-
 20 25 30
Cys-Pro-Cys-Gln-Val-Pro-Ser-Pro
 35

SEQ ID No.: 311A (Pop14)

EP 0 468 527 B1

Sequence Type: AA
Sequence Length: 31 AA

5 Cys-Lys-Pro-Leu-Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-
5 10 15
His-Glu-Tyr-Pro-Val-Gly-Ser-Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-
20 25 30
10 Asp

15

SEQ ID No.: 311B
Sequence Type: AA
Sequence Length: 42 AA

20

Asp-Gly-Val-Arg-Leu-His-Arg-Phe-Ala-Pro-Pro-Cys-Lys-Pro-Leu-
5 10 15
25 Leu-Arg-Glu-Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-His-Glu-Tyr-Pro-
20 25 30
Val-Gly-Ser-Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-Asp
35 40

30

SEQ ID No.: 311C
Sequence Type: AA
Sequence Length: 54 AA

35

Cys-Gln-Val-Pro-Ser-Pro-Glu-Phe-Phe-Thr-Glu-Leu-Asp-Gly-Val-
5 10 15
40 Arg-Leu-His-Arg-Phe-Ala-Pro-Pro-Cys-Lys-Pro-Leu-Leu-Arg-Glu-
20 25 30
Glu-Val-Ser-Phe-Arg-Val-Gly-Leu-His-Glu-Tyr-Pro-Val-Gly-Ser-
35 40 45
45 Gln-Leu-Pro-Cys-Glu-Pro-Glu-Pro-Asp
50

50

SEQ ID No.: 260A
Sequence Type: AA
Sequence Length: 23 AA

55



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EP 0 468 527 B1

Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-ser-
5 10 15
5 Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-
20 25 30
Glu-Glu-Asp-Glu-Arg
35

10

15 **SEQ ID No.: 259C (Pep 5)**
Sequence Type: AA
Sequence Length: 44 AA

20 Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
5 10 15
Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-
20 25 30
25 Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg
35 40

30 SEQ ID No.: 259D
Sequence Type: AA
Sequence Length: 50 AA

35 Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-Ile-Glu-Ala-Asn-Leu-
5 10 15
Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-
20 25 30
40 Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-
35 40 45
Glu-Glu-Asp-Clu-Arg
50

45

50

SEQ ID No.: 259E (Pep 12)
Sequence Type: AA
Sequence Length: 55 AA

55

EP 0 468 527 B1

Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-Glu-Leu-
 5 10 15
 5 Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-
 20 25 30
 Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-
 35 40 45
 10 Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg
 50 55

15 SEQ ID No.: 310A
 Sequence Type: AA
 Sequence Length: 26 AA

20 Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-
 5 10 15
 Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg
 20 25

30 SEQ ID No.: 310B
 Sequence Type: AA
 Sequence Length: 35 AA

35 Ser-Glu-Asn-Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-
 5 10 15
 Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-
 20 25 30
 40 Arg-Lys-Ser-Arg-Arg
 35

45 SEQ ID No.: 310C (Pep 13)
 Sequence Type: AA
 Sequence Length: 47 AA

50

55

EP 0 468 527 B1

Arg-Gln-Glu-Met-Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-
 5 10 15
Lys-Val-Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
5 20 25 30
Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-Arg-Lys-Ser-
 35 40 45
Arg-Arg

15 SEQ ID No.: 309A
Sequence Type: AA
Sequence Length: 27 AA

20 Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-
5 10 15
Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
20 25

30 SEQ ID No.: 309B
Sequence Type: AA
Sequence Length: 35 AA

35 Val-Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-
5 10 15
Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-
20 25 30
40 Arg-Lys-Lys-Arg-Thr
35

45 SEQ ID No.: 309C
Sequence Type: AA
Sequence Length: 44 AA

EP 0 468 527 B1

Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-Lys-
5 10 15
Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-Pro-
20 25 30
Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
35 40

SEQ ID No.: 309D (Pep 11)
Sequence Type: AA
Sequence Length: 60 AA

Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-
5 10 15
Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-
20 25 30
Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-
35 40 45
Pro-Pro-Lys-Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
50 55 60

SEQ ID No.: 309E
Sequence Type: AA
Sequence Length: 72 AA

Ala-Glu-Glu-Asp-Glu-Arg-Glu-Ile-Ser-Val-Pro-Ala-Glu-Ile-Leu-
5 10 15
Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-Pro-Val-Trp-Ala-Arg-
20 25 30
Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-Glu-Thr-Trp-Lys-Lys-Pro-Asp-
35 40 45
Tyr-Glu-Pro-Pro-Val-Val-His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-
50 55 60
Ser-Pro-Pro-Val-Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr
65 70

SEQ ID No.: **Pep 6**
Sequence Type: **AA**
Sequence Length: **37 AA**

Asp-Pro-Gln-Ala-Arg-Val-Ala-Ile-Lys-Ser-Leu-Thr-Glu-Arg-Leu-
5 10 15
s Thr-Val-Gly-Gly-Pro-Leu-Thr-Asn-Ser-Arg-Gly-Glu-Asn-Cys-Gly-
20 25 30
Tyr-Arg-Arg-Cys-Arg-Ala-Ser
35

10

15 SEQ ID No.: **Pep 17**
Sequence Type: **AA**
Sequence Length: **45 AA**

20 Gly-Pro-Ala-Asp-Gly-Met-Val-Ser-Lys-Gly-Trp-Arg-Leu-Leu-Ala-
5 10 15
Pro-Ile-Thr-Ala-Tyr-Ala-Gln-Gln-Thr-Arg-Gly-Leu-Leu-Gly-Cys-
20 25 30
Ile-Ile-Thr-Ser-Leu-Thr-Gly-Arg-Asp-Lys-Asn-Gln-Val-Glu-Gly
35 40 45

25

30 SEQ ID No.: **Pop 18**
Sequence Type: **AA**
Sequence Length: **39 AA**

35 Glu-Ile-Pro-Phe-Tyr-Gly-Lys-Ala-Ile-Pro-Leu-Glu-Val-Ile-Lys-
 5 10 15
Gly-Gly-Arg-His-Leu-Ile-Phe-Cys-His-Ser-Lys-Lys-Lys-Cys-Asp-
 20 25 30
40 Glu-Leu-Ala-Ala-Lys-Leu-Val-Ala-Leu
 35

40

45 SEQ ID No.: **Pop 19**
Sequence Type: **AA**
Sequence Length: **44 AA**

50

55

EP 0 468 527 B1

5 Cys-Val-Arg-Glu-Gly-Asn-Val-Ser-Arg-Cys-Trp-Val-Ala-Met-Thr-
 5 20 25 30
 Arg-Arg-His-Ile-Asp-Leu-Leu-Val-Gly-Ser-Ala-Thr-Leu-Cys
 35 40

10

15 SEQ ID No.: VIII
 Sequence Type: AA
 Sequence Length: 61 AA

20 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-Thr-Asn-
 5 10 15
 Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-Gln-Ile-Val-
 20 25 30
 Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-Gly-Pro-Arg-Leu-Gly-Val-
 25 35 40 45
 Arg-Ala-Thr-Arg-Lys-Thr-Ser-Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-
 50 55 60
 Arg

30

35 SEQ ID No.: IIH
 Sequence Type: AA
 Sequence Length: 47 AA

40 Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-Tyr-Arg-
 5 10 15
 Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-Leu-Pro-Tyr-Ile-
 20 25 30
 Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-Phe-Lys-Gln-Lys-Ala-Leu-
 35 40 45
 Gly-Leu

50 SEQ ID No.: V
 Sequence Type: AA
 Sequence Length: 40 AA

55

Lys-Gln-Lys-Ala-Leu-Gly-Leu-Leu-Gln-Thr-Ala-Ser-Arg-Gln-Ala-
5 10 15
s Glu-Val-Ile-Ala-Pro-Ala-Val-Gln-Thr-Asn-Trp-Gln-Lys-Leu-Glu-
20 25 30
Thr-Phe-Trp-Ala-Lys-His-Met-Trp-Asn-Phe
35 40

15 SEQ ID No.: **Pep XX**
Sequence Type: AA
[Peptide]₁₆ Lys₈ Lys₄ Lys₂ Lys-Y decahexyl peptide
SEQ ID No.: **Pep XXI**
Sequence Type: AA
[Peptide]₈ Lys₄ Lys₂ Lys-Y octameric peptide
SEQ ID No.: **Pep XXII**
Sequence Type: AA
20 [Peptide]₄ Lys₂ Lys-Y tetrameric peptide
SEQ ID No.: **Pep XXIII**
Sequence Type: AA
[Peptide]₂ Lys-Y dimeric peptide

wherein the peptide in Pep XX, Pep XXI, Pep XXII and Pep XIII is selected from the group consisting of (a) to (j):

30 (a) Cys-Leu-Thr-Ile-Pro-Ala-Ser-Ala-Tyr-Glu-Val-Arg-Asn-Val-Ser-
5 10 15
Gly-Ile-Tyr-His-Val-Thr-Asn-Asp-Cys-Ser-Asn-Ser-Ser-Ile-Val-
20 25 30
35 Tyr-Glu-Ala-Ala-Asp-Val-Ile-Met-His-Ala-Pro-Gly-Cys-Val-Pro-
35 40 45
Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys
50 55

(b) Cys-Ile-Thr-Thr-Pro-Val-Ser-Ala-Ala-Glu-Val-Lys-Asn-Ile-Ser-
5 10 15
Thr-Gly-Tyr-Met-Val-Thr-Asn-Asp-Cys-Thr-Asn-Asp-Ser-Ile-Thr-
20 25 30
Trp-Gln-Leu-Gln-Ala-Ala-Val-Leu-His-Val-Pro-Gly-Cys-Val-Pro-
35 40 45
50 Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys
50 55

57

(h) Cys-Val-Arg-Glu-Asn-Asn-Ser-Ser-Arg-Cys-Trp-Val-Ala-Leu-Thr-
5 Pro-Thr-Leu-Ala-Ala-Arg-Asn-Ala-Ser-Val-Pro-Thr-Thr-Thr-Leu-
Arg-Arg-His-Val-Asp-Leu-Leu-Val-Gly-Thr-Ala-Ala-Phe-Cys

10

(i) Cys-Glu-Lys-Val-Gly-Asn-Thr-Ser-Arg-Cys-Trp-Ile-Pro-Val-Ser-
 5 10 15
 Pro-Asn-Val-Ala-Val-Gln-Gln-Pro-Gly-Ala-Leu-Thr-Gln-Gly-Leu-
 20 25 30
 Arg-Thr-His-Ile-Asp-Met-Val-Val-Met-Ser-Ala-Thr-Leu-Cys
 35 40

20

(j) Cys-Glu-Asn-Asp-Asn-Gly-Thr-Leu-Arg-Cys-Trp-Ile-Gln-Val-Thr-
 5 10 15
 Pro-Asn-Val-Ala-Val-Lys-His-Arg-Gly-Ala-Leu-Thr-His-Asn-Leu-
 20 25 30
 Arg-Thr-His-Val-Asp-Met-Ile-Val-Met-Ala-Ala-Thr-Val-Cys
 35 40

30

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Claims

1. A peptide composition comprising a peptide having an amino acid sequence:

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Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

50

Pep11

wherein X is -OH or -NH₂, or

55

an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV,
a conjugate of the above peptide or said analogue peptide with a carrier protein, the conjugate having specific immunoreactivity to antibodies to HCV, and
a polymer of the above peptide, or said analogue peptide having specific immunoreactivity to antibodies to

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HCV said polymer being in dimeric, tetrameric, octameric and decahexyl forms of the peptide and said polymer being synthesised on a branching poly-L-lysine resin which can be Lys₈; Lys₄; Lys₂; Lys, Lys₄; Lys₂; Lys, Lys₂; Lys, Lys.

- 5 2. A peptide composition comprising a mixture of peptides VIIIE and pep 11 wherein peptide VIIIE is:

10 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-
Thr-Asn-Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-
Gly-Gln-Ile-Val-Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-
Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-Arg-Lys-Thr-Ser-
15 Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X;
(VIIIE)

and pep11 is:

20 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
25 Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;
30 Pep11

wherein X is -OH or -NH₂, or

35 an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV.

3. A peptide composition according to claim 2 further comprising Peptide IIH having an amino acid sequence:

40 Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-
Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-
Leu-Pro-Tyr-Ile-Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-
Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X;
45 (IIH)

wherein X is -OH or -NH₂, or

50 an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV.

4. A peptide composition according to claim 2 further comprising pep8 having an amino acid sequence:

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5 Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

pep8

10 wherein X is -OH or -NH₂, or

an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV.

15 5. A peptide composition according to claim 2 further comprising pep12 having an amino acid sequence:

20 Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-
Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-
Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
Asp-Glu-Arg-X;

pep12

wherein X is -OH or NH₂; or

30 an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV.

35 6. A method of detecting antibodies to HCV or NANBHCV by using an effective amount of a peptide composition according to any one of claims 1 to 5 in an immunoassay procedure.

7. A peptide immunogen comprising an octomeric peptide having the structure:

40 [peptide]₈Lys₄Lys₂Lys

wherein the peptide is:

45 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
50 His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

Pep11

55 wherein X is -OH or -NH₂, or

an analogue peptide of the above peptide having an amino acid sequence from a strain/isolate of HCV in a region corresponding to the peptide and having specific immunoreactivity to antibodies to HCV,

a segment of the above peptide or said analogue peptide having specific immunoreactivity to antibodies to HCV, and

a conjugate of the above peptide, said analogue peptide or said segment with a carrier protein, the conjugate having specific immunoreactivity to antibodies to HCV.

5

Patentansprüche

1. Peptidzusammensetzung enthaltend ein Peptid mit einer Aminosäuresequenz:

10

**Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;**

15

20

Pep11

worin-X -OH oder NH₂ ist, oder

25

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz aus dem Stamm/Isolat von HCV in einem Bereich, der dem Peptid entspricht, und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV,

ein Konjugat des vorstehenden Peptids oder des analogen Peptids mit einem Trägerprotein, wobei das Konjugat eine spezifische Immunreaktivität gegenüber HCV hat, und

30

ein Polymer des vorstehenden Peptids oder des analogen Peptids mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV, wobei das Polymer eine dimere, tetramere, octamere und decahexyle Form des Peptids ist, und das Polymer auf einem verzweigten Poly-L-lysin Harz synthetisiert wird, welches Lys₈; Lys₄; Lys₂; Lys, Lys₄; Lys₂; Lys, Lys₂; Lys, Lys sein kann.

35

2. Peptidzusammensetzung enthaltend eine Mischung der Peptide VIIIE und Pep 11, worin Peptid VIIIE:

**Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-
Thr-Asn-Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-
Gly-Gln-Ile-Val-Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-
Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-Arg-Lys-Thr-Ser-
Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X**

45

(VIIIE)

und pep11

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55

Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Por-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X

Pep11

sind, worin X -OH oder -NH₂ ist, oder

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz von einem Stamm/Isolat von HCV in einem Bereich entsprechend dem Peptid und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV.

- 3. Peptidzusammensetzung nach Anspruch 2, enthaltend weiter Peptid IHH mit einer Aminosäuresequenz:**

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-
Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-
Leu-Pro-Tyr-Ile-Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-
Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X,

(IIR)

worin X -OH oder NH_2 ist, oder

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz aus einem Stamm/Isolat von HCV in einem Bereich entsprechend dem Peptid und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV.

- 4. Peptidzusammensetzung nach Anspruch 2, weiter enthaltend Pep8 mit einer Aminosäuresequenz:**

**Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X**

Pep8

worin X -OH oder NH_2 ist, oder

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz aus einem Stamm/Isolat von HCV in einem Bereich entsprechend dem Peptid und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV ist.

- 5. Peptidzusammensetzung nach Anspruch 2, weiter enthaltend Pep12 mit einer Aminosäuresequenz:**

Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-
 Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
 Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-
 Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
 Asp-Glu-Arg-X

Pep12

worin X -OH oder NH₂ ist, oder

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz aus einem Stamm/Isolat von HCV in einem Bereich entsprechend dem Peptid und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV ist.

6. Verfahren zur Bestimmung von Antikörpern gegen HCV oder NANBHV unter Verwendung einer wirksamen Menge einer Peptidzusammensetzung nach einem der Ansprüche 1 bis 5 in einem Immunoassay-Verfahren.

7. Peptidimmunogen enthaltend ein octamer Peptid mit der Struktur:

[Peptid]₈Lys₄Lys₂Lys

worin das Peptid

Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
 Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
 Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
 His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
 Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X

Pep11

worin X -OH oder NH₂ ist, oder

ein analoges Peptid des vorstehenden Peptids mit einer Aminosäuresequenz aus einem Stamm/Isolat von HCV in einem Bereich entsprechend dem Peptid und mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV,

ein Segment des vorstehenden Peptids oder des analogen Peptids mit spezifischer Immunreaktivität gegenüber Antikörpern gegen HCV, und

ein Konjugat des vorstehenden Peptids, des analogen Peptids oder des Segmentes mit einem Trägerprotein, wobei das Konjugat eine spezifische Immunreaktivität gegenüber Antikörpern gegen HCV hat.

Revendications

1. Composition peptidique comprenant un peptide ayant la séquence d'acides aminés

5 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

10

Pep11

dans laquelle X est -OH ou -NH₂, ou

- 15 un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de
HCV (virus de l'hépatite C) dans une région correspondant au peptide et ayant une immunoréactivité spécifique
à l'égard des anticorps anti-HCV,
un conjugué du peptide ci-dessus ou dudit peptide analogue avec une protéine support, le conjugué ayant
une immunoréactivité spécifique à l'égard des anticorps anti-HCV, et
20 un polymère du peptide ci-dessus ou dudit peptide analogue ayant une immunoréactivité spécifique à l'égard
des anticorps anti-HCV, ledit polymère représentant une forme dimère, tétramère, octamère et décahexamère
du peptide et ledit polymère étant synthétisé sur une résine poly-L-lysine de ramification qui peut être Lys₆,
Lys₄, Lys₂, Lys, Lys₄, Lys₂, Lys, Lys₂, Lys, Lys.

- 25 2. Composition peptidique comprenant un mélange de peptides VIII E et pep11, dans lequel le peptide VIII E est

25

30 Ser-Thr-Ile-Pro-Lys-Pro-Gln-Arg-Lys-Thr-Lys-Arg-Asn-
Thr-Asn-Arg-Arg-Pro-Gln-Asp-Val-Lys-Phe-Pro-Gly-Gly-
Gly-Gln-Ile-Val-Gly-Gly-Val-Tyr-Leu-Leu-Pro-Arg-Arg-
Gly-Pro-Arg-Leu-Gly-Val-Arg-Ala-Thr-Arg-Lys-Thr-Ser-
Glu-Arg-Ser-Gln-Pro-Arg-Gly-Arg-Arg-X;

35

(VIII E)

et pep11 est

40

45 Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

50

Pep11

dans laquelle X est -OH ou -NH₂, ou

- 55 un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de
HCV dans une région correspondant au peptide et ayant une immunoréactivité spécifique à l'égard des anti-
corps anti-HCV.

3. Composition peptidique selon la revendication 2, comprenant en outre le peptide I IH ayant la séquence d'acides
amino-

Ser-Gly-Lys-Pro-Ala-Ile-Ile-Pro-Asp-Arg-Glu-Val-Leu-
Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu-Glu-Cys-Ser-Gln-His-
Leu-Pro-Tyr-Ile-Glu-Gln-Gly-Met-Met-Leu-Ala-Glu-Gln-
Phe-Lys-Gln-Lys-Ala-Leu-Gly-Leu-X;

(III)

dans laquelle X est -OH ou -NH₂, ou

un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de HCV dans une région correspondant au peptide et ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV.

4. Composition peptidique selon la revendication 2, comprenant en outre le pep8 ayant la séquence d'acides aminés:

Phe-Thr-Phe-Ser-Pro-Arg-Arg-His-Trp-Thr-Thr-Gln-Gly-
Cys-Asn-Cys-Ser-Ile-Tyr-Pro-Gly-His-Ile-Thr-Gly-His-
Arg-Met-Ala-Trp-Asp-Met-Met-Met-Asn-Trp-Ser-Pro-Thr-
Ala-X;

pep8

dans laquelle X est -OH ou -NH₂, ou

un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de HCV dans une région correspondant au peptide et ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV.

5. Composition peptidique selon la revendication 2, comprenant en outre le pep12 ayant la séquence d'acides aminés:

Lys-Ala-Thr-Cys-Thr-Ala-Asn-His-Asp-Ser-Pro-Asp-Ala-
Glu-Leu-Ile-Glu-Ala-Asn-Leu-Leu-Trp-Arg-Gln-Glu-Met-
Gly-Gly-Asn-Ile-Thr-Arg-Val-Glu-Ser-Glu-Asn-Lys-Val-
Val-Ile-Leu-Asp-Ser-Phe-Asp-Pro-Leu-Val-Ala-Glu-Glu-
Asp-Glu-Arg-X;

pep12

dans laquelle X est -OH ou -NH₂, ou

un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de HCV dans une région correspondant au peptide et ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV.

6. Procédé de détection d'anticorps anti-HCV ou anti-NANBHV, comprenant l'utilisation d'une quantité efficace d'une composition peptidique selon l'une quelconque des revendications 1 à 5 dans une technique d'analyse immunologique.

7. Immunogène peptidique comprenant un peptide octamère ayant la structure:

[peptide]₈Lys₄Lys₂Lys

5

dans laquelle le peptide est:

10

Glu-Ile-Leu-Arg-Lys-Ser-Arg-Arg-Phe-Ala-Gln-Ala-Leu-
Pro-Val-Trp-Ala-Arg-Pro-Asp-Tyr-Asn-Pro-Pro-Leu-Val-
Glu-Thr-Trp-Lys-Lys-Pro-Asp-Tyr-Glu-Pro-Pro-Val-Val-
15 His-Gly-Cys-Pro-Leu-Pro-Pro-Pro-Lys-Ser-Pro-Pro-Val-
Pro-Pro-Pro-Arg-Lys-Lys-Arg-Thr-X;

Pep11

20

où X est -OH ou -NH₂, ou

un peptide analogue du peptide ci-dessus ayant une séquence d'acides aminés issue d'une souche/isolat de HCV dans une région correspondant au peptide et ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV,

25

un segment du peptide ci-dessus ou dudit peptide analogue ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV, et

un conjugué du peptide ci-dessus, dudit peptide analogue ou dudit segment avec une protéine support, le conjugué ayant une immunoréactivité spécifique à l'égard des anticorps anti-HCV.

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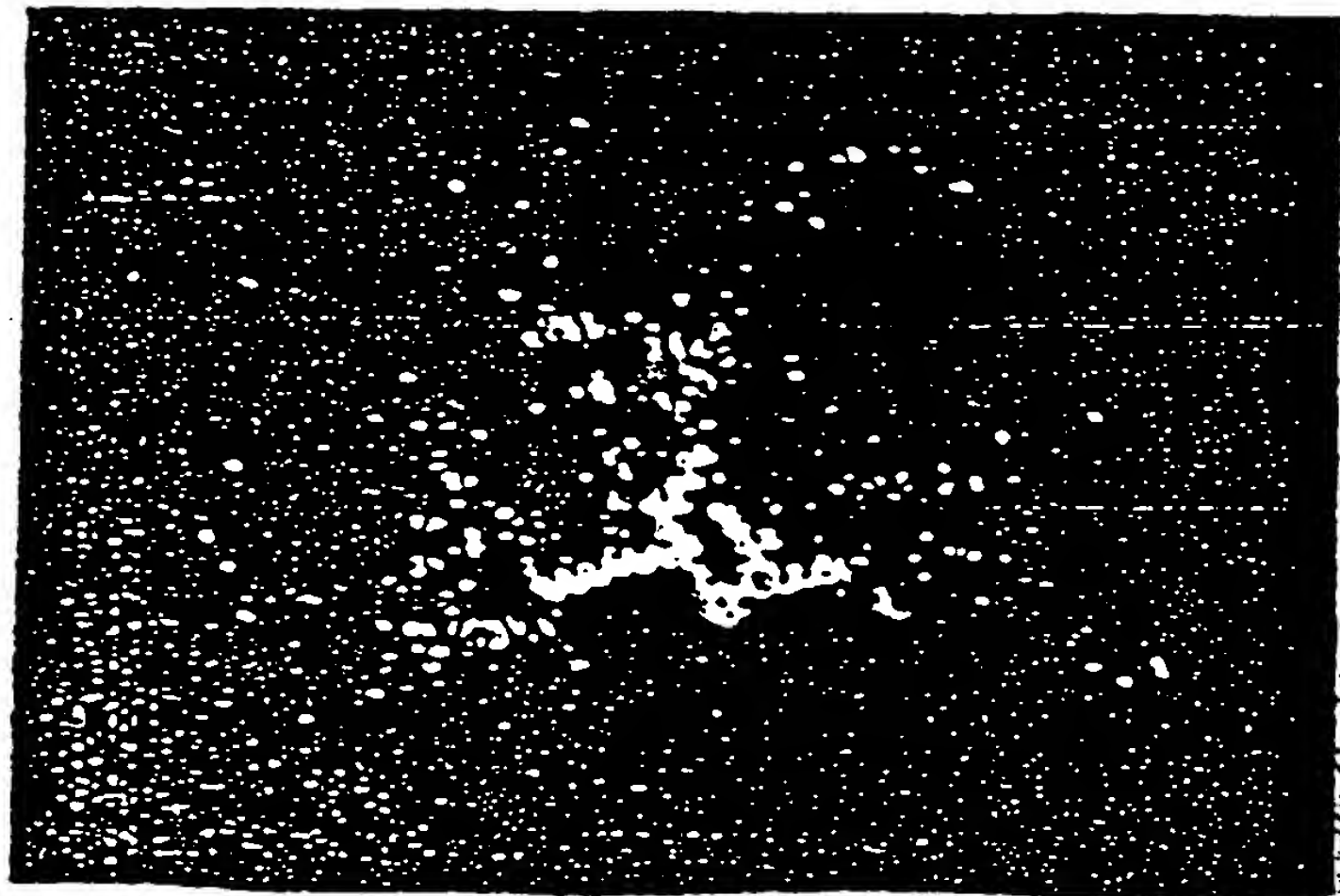


Fig. 1